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Statistical Optimization of Copper Oxide Nanoparticles Using Response Surface Methodology and Box–Behnken Design Towards *In Vitro* and *In Vivo* Toxicity Assessment

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Abstract: The current study focusses on the optimization of Copper oxide nanoparticles (CuO NPs) biosynthesis with *Alternanthera sessilis* (L.) extract using response surface methodology (RSM). The effect of time, pH, and extract to metal concentration ratio on the yield of synthesized nanoparticles (NPs) were estimated using Box–Behnken design. The influence of each of the parameters, as mentioned earlier, was determined by synthesizing nanoparticles under different conditions. A total of 29 experimental runs were carried out to estimate the crucial parameters. Extract to the metal ratio was found to be the vital parameter for yield optimization based on the p-values (p-value < 0.05). The physicochemical property of NPs, like size, was estimated to be in the range of 10-20 nm. In zebrafish, 48 hpf and 72 hpf were measured at 90 μ M to reduce dysfunction and mortality during organ development. These results can have a valuable impact on eco-toxicological effects.

Keywords: box-Behnken design; copper oxide NPs; Alternanthera sessilis; antioxidant; zebrafish.

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1. Introduction

Nano-biotechnology, a field that connects both nanotechnology and biotechnology, is dedicated to product improvement and technology creation. Nanoparticles are employed in the manufacture of several materials, including pigments, cosmetics, biomedical devices, etc [1,2]. Nanomaterials such as copper oxide nanoparticles are recently being employed in pharmaceutical fields, food packaging, and catalyst manufacturing [3,4]. Nanoparticles have been synthesized through numerous methods [5]; conventional strategies use toxic chemicals as reducing or stabilizing agents in order to prevent agglomeration. Also, CuO NPs, which is synthesized through chemical routes, are known to cause acute and chronic toxicity in aquatic organisms [6].

The reduction of copper chloride to copper oxide nanoparticles is carried out by the high content of diterpenoid lactones possessing polyol groups present in the *Alternanthera sessilis* (dwarf copper leaf) leaves at room temperature [7]. The importance of green chemistry has been raised in recent decades to prevent the usage of toxic materials [8]. Green synthesis or the biological method involves the use of plant extracts [9], microorganisms [10], and enzymes [11] is based on the principle of the utilization of non-toxic materials or environment-friendly agents, which also deals with the diminished production of harmful substances [12]. However, there is inadequate evidence to prove the actual toxicity of nanomaterials. The nanotoxicology has focused on *in vitro* assays, using the variety of antioxidant systems. The

advantage of this approach is that it considers the various level of physiological reactivity in the whole organism [13], and so, zebrafish (*Danio rerio*) has rapidly evolved as an ideal model organism for biotoxicity evaluation of nanomaterials. Zebrafish are used as a vertebrate model owing to its properties like small size, high fecundity, transparent embryos, rapid embryonic development cycle, and easy maintenance. Moreover, zebrafish resembles the human at gene metabolism level [14].

The toxicological studies on living organisms of nanoparticles suggested differential toxicities based on physiological properties like size, shape, agglomeration, concentration, dissolution, and application method [15]. Further, parameters such as morphology, concentration, and coating can also significantly contribute to its toxicity. In the present study, CuONPs were synthesized using *Alternanthera sessilis* leaf extract, a single-pot process, with no use of toxic chemicals or reagents. The optimized AS-CuO was then used to evaluate the physicochemical and antioxidant activity. *In vivo* studies using embryonic and larval zebrafish model was used for evaluating the toxicological effects.

2. Materials and Methods

2.1. Sample collection and preparation.

The plant *Alternanthera sessilis* is available throughout tropical Africa, Australia, eastern and southern Asia. Fresh leaves of *Alternanthera sessilis*, free from diseases, were collected from VIT School of Agricultural Innovations and Advanced Learning (VAIAL). The plant was documented by the Plant anatomy research center, Chennai (Reference number: pg:191:1987) [16]. The fresh 250g of leaves was chopped, dipped into 100 mL of distilled water. The mixture was cooled and stored for experimental usage.

2.2. Green synthesis of CuO NPs.

To prepare 50 mM of copper chloride solution, typically, 0.672 g was dissolved in 30 mL of aqueous extract. The solution was kept on an overnight shaker at 150 rpm. The aqueous extract was mixed with the copper chloride stock solution. The solution was kept for overnight shaking at 150 rpm. The formation of a deep green colored solution indicated a synthesis of CuO NPs Figure 1. The solution was transferred into a watch glass and heated at 80°C for 24 h resulting in black powder, which was further used for experimental analysis.



Figure 1. Schematic representation of the green synthesis of CuO NPs from the leaf extract of *Alternanthera sessilis*.

2.3. Design of experiments and optimization.

To optimize the process variables, thereby probing their relative or interactive effects on the synthesis of CuONPs, Box–Behnken design (BBD) was employed. It comprises three steps: (1) design of experimental sets and their execution, (2) regression-based response surface modeling, and (3) optimization and validation of the model. The effective process parameters which substantially contribute to the synthesis of CuONPs were investigated using statistical software called Design-Expert 11. Equation 1 was implemented to find out the number of experiments that would be required for the Box-Behnken design.

Where k and C₀ refer to the number of experimental factors and central points, respectively, four experimental factors, which are independent, were subjected to analysis. The design created 29 experimental runs. The experimental parameters were denoted as A [Concentration of Copper Chloride mM (25, 50, 75 mM)], B [pH of reaction (4, 5, 6)], C [volume of *Alternanthera sessilis* in mL (20, 30, 40)] and D [reaction time (7, 8, 9 hrs)] respectively and coded as -1, 0, 1 (low, central point, high. The relationship between these variables and its interactive responses was fitted using the below mentioned second-order polynomial quadratic equation. The two most crucial aspects, statistical significance, and proficiency of the experimental model, were determined with the help of analysis of variance (ANOVA). This statistical analysis was used in order to ensure that responses related to the levels of each variable could be categorized vigorously by enunciating the aforementioned equation, which had been fitted and represented as surface and contour plots.

2.4. Physiochemical characterizations of CuO NPs.

UV-Vis, DLS, FTIR, XRD, SEM with EDS were employed to understand the composition and physicochemical properties of CuO nanoparticles. The size of particles was estimated by SEM (Zeiss EVO18) attached to EDS and TEM. For SEM analysis, of CuO nanoparticles in solution were dried in a hot air oven, and images were captured at 10KV. The stability and size of CuO nanoparticles in the aqueous medium were estimated through zeta potential and hydrodynamic diameter measurement using DLS (HORIBA SZ100). UV-Visible spectra of CuO nanoparticles were obtained in the range of 200–600 nm at different time intervals using a double beam UV-Visible spectrophotometer (Cary 5000, Agilent, USA). XRD spectra were obtained in the $2\theta^{\circ}$ range of 20° to 80° to comprehend the crystallinity and size of CuO nanoparticles using X-ray diffractometer (XRD Brukerd 8 Advance). FTIR spectroscopy, in the range of 400-4000 cm⁻¹, was used to identify the compounds involved in the reduction and capping of nanoparticles.

2.5. DPPH activity.

A similar protocol was followed for the antioxidant activity of *Magnolia champaca* extracts mediated CuO nanoparticles was evaluated by the DPPH method [17]. Various concentrations of CuO nanoparticles (100-500 μ g/ml) were mixed with a constant range of (500 μ l) DPPH solution. Finally, the solution was made with 3 mL of methanol. The same concentration of ascorbic acid was prepared used as a positive control. This entire mixture was shaken and for 30 min under the dark condition at room temperature. The absorbance was recorded at 517.

Scavenging activity (%) = control-test/control
$$\times 100$$
 (1)

2.6. Radical ABTS radical scavenging assay.

The *in-vitro* antioxidant activity of ABTS has formed. 2 mM ABTS solution with 17 mM, (0.3ml) potassium persulfate was kept in the dark condition for 12-16 h at 28°C. This

solution was diluted in ethanol at 30°C. Then, the ABTS solution was added to various concentrations of 100- 500 μ g/mL CuO nanoparticles and kept in the dark condition at room temperature for 30 min. Ascorbic acid of the same concentration was used as a positive control. Absorbance was then measured at 734 nm.

ABTS (%) = control-test/control $\times 100$ (2)

2.7. Nitric oxide radical scavenging activity.

Nitric oxide radical capturing analysis was calculated using the UV-Visible spectrophotometer method. Test samples at different concentrations (100-500 μ g/mL) were dissolved in DMSO. 1ml of test solution was mixed with 1ml of sodium nitroprusside (5 mM) in phosphate buffer saline and incubated at 25°C for 30 min. Medium without a test solution was used as control. 1ml of Griess reagent was then added to an equal volume of incubated solution. The chromophore produced for the period of the diazotization of the nitrile with sulphanilamide was estimated at 546 nm.

NO (%) = control-test/control
$$\times 100$$
 (3)

2.8. In-vivo toxicity of green synthesized CuO NPs on zebrafish.

All experiments were performed according to relevant animal practice guidelines and regulations of OECD. Adult zebrafish were maintained at a temperature of 28.5°C, with a pH of 7 ± 0.2 , under 14:10 h light / dark photoperiod. They were given live salt shrimp once a day and dry meals twice a day. A natural random mating procedure was used to obtain zebrafish embryos, which were further raised in embryonic water (0.2 g/L saline in double-distilled water) at 28.5°C. Zebrafish fertilized eggs were stored at 24 ± 3 °C until hatching. The morphological changes and developments were monitored using microscopy. The rate of hatching was determined by hatching 72 hpf compared to the untreated group of multiple embryos. Mortality was depicted as many dead embryos after 72 hpf compared with the group that was left untreated. The experiment was performed thrice.

2.9. Zebrafish embryotoxicity (mortality and hatchability).

Disclosures were conducted with minor modifications following OECD Guideline 236 (OECD, 2013). Briefly, fertilized zebrafish embryos (<6 hpf) were tested for viability, and healthy embryos were collected for estimating the mortality and hatchability in almost equivalent growth conditions. Tested for each sample used different concentrations at 30, 60, 90, 120, and 150 μ M. Mortality and hatching were observed during the exposure period of 5 days under extreme care. The mortality was recorded after 1, 2, 3, 4 days of the incubation period, and the percentage of mobility was reported. The success of the hatching process was confirmed after 96 h of hatching.

2.10. Statistical analyses.

All results are represented as the mean for experiments repeated in triplicated and presented with standard deviation. Statistical analysis of the results was carried out by using Graph pad 6.1 software package and considered significant at <0.05 p values.

3. Results and Discussion

3.1. Design of experiments and optimization.

The regression equation, which was represented graphically using the threedimensional response surface plots and the corresponding contour plots. Figure 2a. Shows the contour plot of two factors (volume of Andrographis paniculate extract and amount of copper nitrate), while Figure 2b shows the corresponding 3D surface plot for the CuO NPs yield. Table 1 contains the list of experimental runs and the similar responses obtained from the experiments projected by BBD. A regression coefficient R2 value of 0.9802 was obtained with a secondorder quadratic equation generated for the green synthesis of CuO NPs.

The statistical significance of the model was ascertained based on the results obtained from the analysis of variance. As observed from the results, the model F-value was 49.47, which implied that the model was significant. The predicted R2 (0.8886) was found to agree with the adjusted R2, which was calculated to be 0.9604. The computed value of C.V% was found to be 2.63, which is relatively low, indicating greater accuracy and consistency of the model.

The adequacy of the model was evaluated using the residuals from the least-squares fit as a tool. The quadratic model generated using the response surface result was found to be significant as the p-value is < 0.0001. The standard probability plot was plotted to assess the normality assumptions. It suggests that the assumptions made are agreeable based on the fact that the extra points cluster along the diagonal line. The value of the absorbance generated by performing the experiments, and the corresponding values predicted by the statistical model Figure 3, which relate to the amount of CuO NPs being synthesized, were compared. The results (A) Copper Chloride 50 mM, (B) PH of reaction 5, (C) *Alternanthera sessilis* in 30 mL, and (D) reaction time of 8 hrs indicate that the empirical model is considerably on a par with the experimental one within the given innovative ranges.



Figure 2. (a) 2 D Contour plot (b) 3D surface plot of absorbance versus concentration of copper chloride and volume of *Alternanthera sessilis*.



Figure 3. (a) Normal plot for residuals (b) predicted versus actual plot (c) residual versus predicted plot for the biosynthesis of CuO NPs model.

3.2. Validation of the statistical model.

To establish the significance and to further adequacy of the generated models for maximization of the yield nanoparticles, the experiment was repeated using variable levels suggested by the optimum model, and size was calculated (Table 2). The size of the synthesized CuO NPs was 9.55nm, which was further confirmed by the data generated using the statistical tool. The results emphasized the effectiveness of RSM in experimental design for optimizing the process parameters and in projecting the prominence of both the individual and interactive paraphernalia of the experimental variables in nanoparticle synthesis.

3.3. UV- Vis Spectra studies.

UV–Vis spectra indicated the synthesis of shape and size-controlled nanoparticles. Visual color change from green to greyish black suggests the synthesis of CuO NPs with the use of *Alternanthera sessilis* extract Figure 1 represents the synthesis. Surface Plasmon Resonance (SPR) is an optical property exhibited by noble metal nanoparticles upon interaction with resonant light. The change in color is attributed to the excitation of surface plasma vibrations (SPR) in the presence of metal nanoparticles [18]. The absorbance maxima peak demonstrates the presence of CuO NPs as CuO SPR bands are centered between 335 and 400 nm Figure 4.



Figure 4. UV-Vis spectra of synthesized CuO NPs.

3.4. Size and shape determination of CuO nanoparticles.

HRTEM and SEM are the most powerful analytical tools available, providing information on the morphology of the synthesized CuO NPs.



Figure 5. Electron micrographs of green synthesized CuO NPs. (a) TEM (Scale bar of 20 nm). (b) Magnified view of TEM (scale bar 10 nm). (c) SEM (Scale bar of 2µm). (d) Magnified view of SEM (scale bar 300 nm).

Particles were confirmed to be spherical with a uniform diameter (10-20 nm) using HRTEM Figure 5 a-b and SEM Figure 5 c-d. EDS analysis confirmed the purity of CuO NPs by 57.81% (Cu) and 34.07% (O) weight percent. However, a small amount of chloride was also found in the EDS spectra Figure 6. HRTEM image reveals capping of CuO NPs with a thickness of 20 nm, which could be attributed to the organic compounds of the leaf. Previous studies report the interaction of secondary metabolites with metal and metal oxide nanoparticles [19].



Figure 6. EDS analysis of CuO NPs.

3.5. DLS and Zeta analysis of CuO NPs.

The stability of nanoparticles is essential for medicinal applications. Interactions occurring in nanoparticle solution include the intramolecular interaction among nanoparticles, and intermolecular interactions of nanoparticles with the surrounding fluid eventually results in nanoparticles agglomeration [20]. The high zeta potential value represents nanoparticles stability in an aqueous medium Figure 7. The Zeta potential value of -18 mV indicates excellent colloidal stability.

DLS is a potent technique for monitoring the size of nanoparticles and estimating their colloidal stability [21]. DLS is commonly employed to estimate the hydrodynamic diameter of CuO NPs. The hydrodynamic diameter of CuO NPs was determined to be 80.2 nm using DLS Figure 8.



Figure 7. Zeta potential measurement of green synthesized CuO NPs.

Figure 8. Particle size of CuO NPs measured by Dynamic light scattering.

3.6. FTIR and XRD analysis of biosynthesized CuO NPs.

FTIR spectra revealed the presence of biomolecules in *Alternanthera sessilis* extract potential responsible for capping, reduction, and stabilization of CuO NPs. FTIR spectra of *Alternanthera sessilis* (AS) extract and dried CuO NPs are depicted in Figure 9A-B. Previous

literature suggests the role of tannins, carbohydrates, proteins, and ascorbic acid from the *Alternanthera sessilis* extract in synthesis and capping of Ag NPs [22]. These secondary metabolites are known to possess antioxidant, anti-inflammatory potentials [23]. FTIR spectra of AS reveals the presence of broad -OH stretching vibration at 2945.30 cm⁻¹ and cellulose at 3313.71 cm⁻¹. Other bands at 2833.43 cm⁻¹, 1658.78 cm⁻¹, 1413.82 cm⁻¹, 1112.93 cm⁻¹, 1024.20 cm⁻¹ are attributed to the stretching vibration of alkane, stretching vibration of aromatic compounds, ketone, carboxylic acid [24], and C-N stretching vibrations of aromatic nitro compounds [25]. , The presence of similar bands in the FTIR spectra of CuO NPs, suggests the involvement of *Alternanthera sessilis* extract in the synthesis and reduction of nanoparticles. FTIR spectra of CuO NPs indicates the presence of bands at 3439.08 cm⁻¹, 3331.07 cm⁻¹, 1622.13 cm⁻¹, 1396.46 cm⁻¹, 1078.21 cm⁻¹, 802.39 cm⁻¹ and 580.57 cm⁻¹ corresponding to O-H bond stretching vibration, -NH2 stretching vibration of amino acid salts, C=N stretching vibration of alcohols, CH3-metal groups and CS2 stretching vibration. Results were similar to previous literature [26].

The X-ray diffraction pattern of the synthesized CuO NPs is depicted in Figure 10. XRD peaks were indexed using the standard peak values available in JCPDS files (JCPDS card no.: 89-5895) [24,27]. The position and relative intensities of all eleven diffraction peaks were well consistent with the standard XRD pattern of CuO. Some of the characteristics peaks are observed at 20 angles of 32.82°, 35.83°, 39.04°, 49.21°, 53.49°, 58.71°, 62.05°, 66.47°, 68.47°, 72.62°, 75.16° and are assigned respectively to the (110), (111), (200), (202), (020), (202), (113), (022), (113), (311), and (004) diffraction planes. The XRD pattern revealed the formation of CuO NPs with a monoclinic structure. The average size of the synthesized nanoparticles was observed to be 17.28 nm.



Figure 9. FT-IR spectrum of (A) Alternanthera sessilis (B) CuO NPs.



Figure 10. X-ray diffraction pattern of CuO NPs.

3.7. Antioxidant activity.

The DPPH radical scavenging assay of CuO NPs at different concentrations is shown in Figure 11a. The results revealed dose-dependent % inhibition of 27.27 % at 100 µg/mL, 39.81% at 200 µg/mL, 61.89% at 300 µg/mL, 74.97% at 400 µg/mL, 84.63% at 500 µg/mL respectively. CuO NPs demonstrated significant radical scavenging activity when compared to ascorbic acid. Phytochemicals present in the Alternanthera sessilis might have contributed to the high antioxidant potential of CuO NPs [28]. The ABTS radical scavenging assay of the synthesized CuO NPs is shown in Figure 11b. At 500 µg/mL concentration, the inhibition of CuO NPs was found to be 71.02%, and that of the standard was 66.46%. The result shows dose-dependent % inhibition of 16.24 % at 100 µg/mL, 40.02% at 200 µg/mL, 54.30 % at 300 μ g/mL, 62.03 % at 400 μ g/mL, 71.02 % at 500 μ g/mL. The NO radical scavenging assay of the synthesized CuO NPs is sown in Figure 11c. The results demonstrate % inhibition of 25.55 % at 100 µg/mL, 45.16 % at 200 µg/mL, 58.44 % at 300 µg/mL, 75.48 % at 400 µg/mL, 82.68 % at 500 µg/mL. The results suggested that the synthesized CuO NPs have moderate antioxidant activity against ABTS and NO radical scavenging assays. The hydroxyl group of plant phenolic compounds show scavenging potential and, thus, can be a contributor to the antioxidant activity of CuO NPs [29].



Figure 11. In vitro antioxidant activity of copper oxide nanoparticles using (A) DPPH assay (B) ABTS assay, and (C) NO assay.

3.8. In vivo toxicity of green synthesized CuO Nanoparticles in zebrafish embryos.

Rate of hatching and viability was considered to be viral parameters to evaluate the toxicity of CuO NPs on Zebrafish embryonic development [30]. The mortality rate of zebrafish embryos was high in CuO NPs treated embryo when compared to the AS-CuO treated one. Compared with the AS-CuO groups, the mortality rate was estimated to be 5 times more in CuO treated groups at a concentration of 120 μ M. The hatching rates of zebrafish embryos after the CuO NPs treatments were given in Figure 12A. As shown in Figure 12B, control groups and the CuO NPs treated embryos started to hatch at 48 hpf. A dose-dependent decrease in the hatching rate decreased significantly was observed between 48 and 72 hpf [31]. As compared to the AS-CuO group, the hatching rate in the highest concentration (150 μ M AS-CuO) group was decreased by about 30%. The highest body malformation rate was found for the highest concentration of CuO NPs treatment (150 μ M) Figure 12C.



Figure 12. Zebrafish embryo viability, hatchability, and embryo development.

ĺ	Factor 1	Factor 2	Factor 3	Factor 4	Response
Run	A: Concentration	B: pH	C: Volume	D: Time	Absorbance
	Μ		ml	hrs	a. u
1	0.05	6	30	7	0.77
16	0.075	6	30	8	0.84
8	0.05	6	40	8	0.85
14	0.05	6	20	8	0.87
12	0.025	6	30	8	0.88
20	0.05	4	30	9	0.89
6	0.05	6	30	9	0.92
9	0.05	4	20	8	0.95
4	0.025	4	30	8	0.96
2	0.025	5	30	9	0.97
3	0.05	4	30	7	0.98
21	0.05	5	20	9	0.98
24	0.05	5	40	7	0.98
25	0.025	5	30	7	0.98
22	0.025	5	20	8	0.99
5	0.05	5	20	7	1
15	0.025	5	40	8	1
27	0.05	4	40	8	1.02
13	0.075	5	30	7	1.03
7	0.075	4	30	8	1.04
29	0.075	5	40	8	1.04
10	0.075	5	20	8	1.05
11	0.075	5	30	9	1.08
28	0.05	5	40	9	1.08
17	0.05	5	30	8	1.22
19	0.05	5	30	8	1.23
18	0.05	5	30	8	1.24
23	0.05	5	30	8	1.24
26	0.05	5	30	8	1.24

Table 1. Experimental matrix and responses	(absorbance) for Box-Behnken design
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Number of runs	Absorbance at 400 nm
1	1.22
2	1.24
3	1.24
Average	1.23
Predicted	1.27

4. Conclusions

In the present study, the biosynthesis of CuO NPs was attempted using greener techniques by optimizing all crucial parameters using statistical designs. The presence of CuO in the synthesized nanoparticle was confirmed using UV-Vis spectrophotometry, FTIR, EDS, and XRD techniques. The morphology and the size of CuO NPs were determined by SEM and TEM studies. In summary, successful green synthesis of stable, well-characterized CuO nanoparticle was carried out, the *in vitro* and *in vivo* toxic impact was investigated with comparison to commercially available CuO NPs. Data from this report suggested a novel method of green synthesis of CuO nanoparticles.

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Conflicts of Interest

The authors declare no conflict of interest.

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