Equilibrium and Kinetic Study on the Biosorption of Trypan Blue from Aqueous Solutions using Avocado Seed Powder

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Abstract: This research aims to investigate the removal of trypan blue dye from aqueous solutions by employing ground avocado seeds powder, a low-cost biowaste adsorbent (biosorbent), under various experimental conditions. The effect of contact time, initial dye concentration, and adsorbent dose on dye removal were studied. The experimental kinetic data were fitted to pseudo-first-order and pseudo-second-order kinetic models. Results imply that adsorption of trypan blue on the avocado seed adsorbent nicely followed the pseudo-second-order kinetic model. Equilibrium isotherms were analyzed by Langmuir and Freundlich isotherms, where Langmuir isotherm described the isotherm data with a high-correlation coefficient (R²=0.9948) closer to unity, and maximum adsorption capacity was found to be 19.3 mg g⁻¹. The present study results substantiate that ground avocado seeds are a promising adsorbent for the removal of the dye trypan blue from industrial wastewater.

Keywords: dye removal; biosorbent; wastewater treatment; pollution; kinetic models.

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1. Introduction

Water is a vital and essential compound for life on earth [1-7]. The major sources of water pollution include wastewater discharge from industries and agricultural activities. The presence of contaminants, including heavy metals, dyes, and microorganisms, even in very small amounts, still proves to be very dangerous to not only human health but to aquatic systems and the environment alike [8-14].

Several remediation approaches have been adopted to remove such pollutants from wastewater. Those proposed techniques include precipitation, incineration, flocculation, coagulation, ion exchange, reverse osmosis, membrane filtration, electrochemistry, photoelectrochemistry, among many others. Some downsides of said methods are the low removal efficiency, high production and maintenance costs, and the generation of potentially toxic by-products and introducing this to the environment [15-22].
Adsorption by solid adsorbents is considered an effective method for the treatment and removal of several contaminants in wastewater treatment. Adsorption has advantages over the other methods because of simple design, ease of operation, low cost, and high performance. Nowadays, adsorption processes are widely used to treat industrial wastewater from both organic and inorganic pollutants. Materials widely available such as agricultural and industrial wastes can be used as low-cost adsorbents. Additionally, activated carbon can be produced from these waste materials upon some chemical and thermal treatment and be utilized as an adsorbent for water and wastewater treatment. [23-28] Thus, this study aims to examine the efficacy of avocado seeds, a common biowaste, in adsorbing dyes present in industrial wastewater [29-31].

Biosorption is a physicochemical process by which certain biomass passively concentrates and binds contaminants onto its cellular structure. In the past, the term biosorption was used to describe metal removal by microorganisms. The terms biosorption and bioaccumulation are used interchangeably in the literature; however, a distinction has to be made. Bioaccumulation is based on the use of living cells, while biosorption is based on dead biomass use [32-36]. The biosorption of toxic metals, dyes, and other pollutants is affected by several factors, such as the solution pH, temperature, contaminant concentration, time, and ionic strength. Among all these factors, pH is considered one of the most if not the most important key factor of the biosorption process [37-44]. The pH affects the solution chemistry of metals and the functional groups’ activity present in the biosorbent. As the pH decreases, the competition among proton and metal ions in solution for functional groups present on the biosorbent surface increases, reducing the biosorption capacity. Nevertheless, at high pH values, the metal uptake is affected by the insoluble metal compound's formation. [45-48] Before going in further, some terminology regarding adsorption processes will need to be addressed. Adsorption is when atoms, ions, or molecules from a gas, liquid, or dissolved solid adhere to a surface. The adsorbent is a substance that is usually porous with a high surface area that can adsorb substances onto its surface with the help of intermolecular forces, while the adsorbate is a substance that gets adsorbed on a surface of another substance, which is the adsorbent in this case. The amount of dye adsorbed, \( q_e \), is given by equation 1, where \( C_0 \) is the initial dye concentration, \( C_e \) is the final dye concentration, \( V \) is the solution volume, and \( W \) is the added weight adsorbent [47-50].

Accordingly, when a solution containing pollutants (absorbable solute) comes into contact with a solid with a highly porous surface structure (adsorbent), intermolecular forces of attraction cause some of the pollutants in the solution to get deposited at the solid surface, and that’s the main principle behind using adsorption for wastewater treatment [51-54].

2. Materials and Methods

2.1. Materials.

Chemicals were used as received without any modification or further steps of purification. Avocado seeds were the natural biosorbent choice for this study. Trypan Blue (C\(_{34}\)H\(_{24}\)N\(_6\)Na\(_4\)O\(_{14}\)S\(_4\)) was the dye used to test out the adsorbent’s effectiveness, as shown in figure 1. Usually, more popular dyes like methylene blue or crystal violet are used for such studies, but that does not undermine the importance of studying the effectiveness of a natural biosorbent on adsorbing the trypan blue dye. Trypan Blue is an azo acid dye commonly used as a stain to differentiate amongst viable and non-viable cells. This azo dye turns dead cells
blue while leaving viable cells unstained. It’s also used for directly dyeing cotton textiles. However, it’s not as popular in the textile industry as the aforementioned dyes above.

Furthermore, it is a known animal carcinogen and an experimental teratogen. Its introduction to the environment in the form of waste from the textile industry can prove to be a serious threat to the environment and animals and humans alike. The dye is readily soluble in water and was found to have a UV maximum absorption range of 584-617 nm.

![Figure 1. Structure of the trypan blue dye.](https://doi.org/10.33263/BRIAC113.1104211053)

2.2. Preparation of the adsorbent and dye stock solutions.

In the typical synthetic procedure, Avocados seeds were extracted from locally bought avocados. They were then thoroughly washed with distilled water, dried at 80 °C in an oven for 24h, and then cut and ground to a fine powder. During the thermal treatment, a loss of biomass was produced. The percentage of loss was determined using equation 10 shown below, where, \( m_i \) and \( m_f \) (g) are the initial and final biomass, respectively.

The Trypan Blue Dye purchased from Serva chemicals of 99.9% purity was available in powder form, so a stock solution was made by dissolving a certain amount of the dye into distilled water to obtain a stock solution with a final concentration of \( 1 \times 10^{-3} \) M using equation 11, where \( wt \) is the weight of the dye needed in grams, \( C \) is the final desired concentration in M, \( V \) is the volume of the solution in ml, \( Mr \) is the molecular weight of the dye, and divide by a thousand to convert milliliter to liter. Accordingly, the stock solution was diluted even further to obtain six trypan blue solutions in various concentrations, namely, \( 1 \times 10^{-5} \) M, \( 2 \times 10^{-5} \) M, \( 4 \times 10^{-5} \) M, \( 6 \times 10^{-5} \) M, and \( 8 \times 10^{-5} \) M.

2.3. The adsorption experiments.

For all the experiments, 250 ml conical flasks were used along with hot plate stirrers and stirring bars of the same size. Stuart Magnetic stirrer with heating analog US152 was used for both the heating and stirring at a fixed stirring rate (scale 3 on the device). Additionally, a fixed volume of 25 ml of the dye solution was used for all the tests. The adsorbent dose and time sometimes varied depending on the test. The effect of time, adsorbent dose, and initial dye concentration on the dye's percentage removal were explored thoroughly in this study.

After the tests are carried out, a UV reading at \( \lambda_{\text{max}} = 580 \) nm is taken to determine the final absorption value. Hence, the dye's final concentration remaining in the solution is determined using the best fit line equation of the calibration curve. Finally, equation 13 is used to calculate the % adsorption or removal of the dye in the solution [1, 11, 24, 42].

3. Results and Discussion

3.1. Effect of initial dye concentration.

Table 1 discuss the influence of using different concentrations on the absorbance. The effect of initial dye concentration was the first effect studied in this work. A series of five Trypan Blue solutions with \( 1 \times 10^{-5} \) M, \( 2 \times 10^{-5} \) M, \( 4 \times 10^{-5} \) M, \( 6 \times 10^{-5} \) M, and \( 8 \times 10^{-5} \) initial dye concentrations were used in this study. The calibration curve is shown in figure 3.

https://doi.org/10.33263/BRIAC113.1104211053

https://biointerfaceresearch.com/
Table 1. The impact of using different concentrations on absorbance.

<table>
<thead>
<tr>
<th>Concentration (mol/L) (1x10^{-5})</th>
<th>Absorbance</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.274</td>
</tr>
<tr>
<td>2</td>
<td>0.59</td>
</tr>
<tr>
<td>4</td>
<td>1.165</td>
</tr>
<tr>
<td>6</td>
<td>1.694</td>
</tr>
<tr>
<td>8</td>
<td>1.965</td>
</tr>
</tbody>
</table>

Figure 3. Calibration curve.

Table 2. The effect of initial dye concentration.

<table>
<thead>
<tr>
<th>C_0 (mol/L) (1x10^{-5})</th>
<th>Absorbance</th>
<th>C_e (mol/L)</th>
<th>% Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.061</td>
<td>1.55E-06</td>
<td>84.5 %</td>
</tr>
<tr>
<td>2</td>
<td>0.12</td>
<td>8.37E-07</td>
<td>95.8 %</td>
</tr>
<tr>
<td>4</td>
<td>0.215</td>
<td>4.68E-06</td>
<td>88.3 %</td>
</tr>
<tr>
<td>6</td>
<td>0.293</td>
<td>7.84E-06</td>
<td>86.9 %</td>
</tr>
<tr>
<td>8</td>
<td>0.736</td>
<td>2.57E-05</td>
<td>67.8 %</td>
</tr>
</tbody>
</table>

As discussed before, table 2 shows the effect of initial dye concentration was investigated through a series of five Trypan Blue solutions with 1x10^{-5} M, 2x10^{-5} M, 4x10^{-5} M, 6x10^{-5} M, and 8x10^{-5} initial dye concentrations used in this study.

Figure 4. Effect of initial dye concentration.

It seems that the investigated series of the selected five Trypan Blue solutions with 1x10^{-5} M, 2x10^{-5} M, 4x10^{-5} M, 6x10^{-5} M, and 8x10^{-5} M initial dye concentrations used in this study have different removal percentage as 2x10^{-5} M is the best and efficient concentration in removal percentage, however, increasing the concentration could not enhance the removal efficiency as it keeps decreasing until the minimum removal percentage was recorded for 8x10^{-5} M as shown in figure 4.
3.2. Effect of contact time.

Like temperature, pH, and the biosorbent dose, all other variables were kept constant in this study. The solutions were all stirred for ten minutes, filtered, and then their UV absorptions were measured after, as shown in table 3. It appears that $2 \times 10^{-5}$ M is the optimum initial dye concentration capable of removing 95.8% of the trypan blue dye from the prepared aqueous solution.

Beyond this point, it appears that saturation was achieved, and an increase in the dye concentration would lead the equilibrium to shift back to the left and promote desorption instead of adsorption.


Table 3. The effect of contact time.

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>Absorbance</th>
<th>$C_e$(mol/L)</th>
<th>% Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>0.594</td>
<td>2.13E-05</td>
<td>65.8 %</td>
</tr>
<tr>
<td>20</td>
<td>0.584</td>
<td>1.96E-05</td>
<td>67.3 %</td>
</tr>
<tr>
<td>30</td>
<td>0.428</td>
<td>1.33E-05</td>
<td>77.8 %</td>
</tr>
<tr>
<td>40</td>
<td>0.389</td>
<td>1.17E-05</td>
<td>80.5 %</td>
</tr>
</tbody>
</table>

Figure 5. Effect of contact time.

Once the initial dye concentration was examined, the contact time was the second effect to be studied. Studying the effect of contact time is of great importance because it could lead to greater efficiency if, for instance, 30 minutes was the time need to get optimum results, then leaving it for more time would be ineffective and might lead to desorption. All the variables like temperature, biosorbent dose, and dye concentration were fixed at $6 \times 10^{-5}$ M, and only contact time was varied from 10 to 60 minutes. As seen in figure 5, starting from the 30-minute mark, the plot starts to plateau. Hence, it can be concluded that this is an experimental $q_e$ value, and going beyond this point wouldn’t be productive since the percent removal wouldn’t vary by much at all since the adsorbent is most likely fully saturated at this point.

3.3. Effect of biosorbent dose.

Table 4 reveals that the dose of the biosorbent used is one of the most important variables studied in this work since it determines the capacity of the biosorbent.


Table 4. The effect of biosorbent dose.

<table>
<thead>
<tr>
<th>Mass (g)</th>
<th>Absorbance</th>
<th>$C_e$(mol/L)</th>
<th>% Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.05</td>
<td>1.321</td>
<td>4.94E-05</td>
<td>17.6 %</td>
</tr>
<tr>
<td>0.1</td>
<td>0.994</td>
<td>3.62E-05</td>
<td>39.7 %</td>
</tr>
<tr>
<td>0.2</td>
<td>0.693</td>
<td>2.40E-05</td>
<td>59.9 %</td>
</tr>
<tr>
<td>0.3</td>
<td>0.334</td>
<td>9.49E-06</td>
<td>84.2 %</td>
</tr>
<tr>
<td>0.4</td>
<td>0.55</td>
<td>1.82E-05</td>
<td>69.6 %</td>
</tr>
</tbody>
</table>
Figure 6. Effect of biosorbent dose.

The amount of avocado seed powder (biosorbent) was changed from 0.05 to 0.4 grams in 25 ml of dye solution of the same concentration of $6 \times 10^{-5}$ M. All the other variables such as the contact time, pH, and temperature were kept constant. Once again, the solutions were all stirred for ten minutes, filtered, and then their UV absorptions were measured after. Figure 6 shows that as the dose increases from 0.05 to 0.3 grams, the removal percentage increased from 17.6% to 84.2%, but then there’s a dip in the percent removal with the test run with 0.4 grams of the biosorbent, and this might be due to the saturation effect of the dye-binding sites leading to aggregation, for instance. From the data shown in the table, it appears that 0.3 g is the optimum biosorbent dose.

3.4. Adsorption isotherms.

In order to use the equations of the adsorption isotherms, the concentrations need to be in mg/L. The equation below was used to change the concentrations from molarity to mg/L, and the results are shown below in table 5. To quantitatively compare the two models’ accuracy, the correlation coefficients were calculated where the model having a higher correlation coefficient would fit the data best.

$Concentration\ (\frac{mg}{L}) = Concentration\ (M) \times Molecular\ Weight\ of\ the\ Dye \times 100$

<table>
<thead>
<tr>
<th>$C_0\ (M)\ (1 \times 10^{-5})$</th>
<th>$C_0\ (mg/L)$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>9.608</td>
</tr>
<tr>
<td>4</td>
<td>38.432</td>
</tr>
<tr>
<td>6</td>
<td>57.648</td>
</tr>
</tbody>
</table>

Figure 7. Langmuir isotherm.
The higher correlation coefficient ($R^2=0.9948$), as shown in figure 7 indicates that the Langmuir isotherm model describes trypan blue's adsorption onto the avocado seed powder best. Moreover, the separation factor, $R_L$, is 0.464, lying in the middle of zero and one, which is matching the typical adsorption process according to the Langmuir model. However, it also appears from figure 8 that the Freundlich model still can be considered valid and can be used to describe our adsorption of interest since the value of the non-linearity constant, $n$, is 1.16 and hence greater than one indicating a favorable adsorption process according to the Freundlich model as shown in table 6.

Table 6. Langmuir and Freundlich isotherm constants for the adsorption of trypan blue dye using avocado seed powder.

<table>
<thead>
<tr>
<th></th>
<th>$Q_m$</th>
<th>$K_L$</th>
<th>$R^2$</th>
<th>$R_L$</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Langmuir Adsorption Isotherm</strong></td>
<td>19.3</td>
<td>0.12</td>
<td>0.9948</td>
<td>0.464</td>
</tr>
<tr>
<td><strong>Freundlich Adsorption Isotherm</strong></td>
<td>2.03</td>
<td>1.16</td>
<td>0.9876</td>
<td></td>
</tr>
</tbody>
</table>

3.5. Adsorption kinetics.

The calculations were carried out for the 57.648 mg/l dye solution to investigate the kinetic model for this adsorption model. Other conditions like oscillation speed, temperature, and biosorbent dose were kept constant. Table 7 shows the results obtained from figures 9 and 10. It can be seen that the correlation coefficient ($R^2=0.9916$) for the pseudo-second-order kinetic model was higher than that of the pseudo-first-order kinetic model ($R^2=0.8398$).

Table 7. Results and calculations for the first-order model.

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>$q_t$</th>
<th>$q_e$</th>
<th>$\log q_e - q_t$</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>3.734</td>
<td>0.573</td>
</tr>
<tr>
<td>10</td>
<td>3.09</td>
<td>3.74</td>
<td>-0.191</td>
</tr>
<tr>
<td>20</td>
<td>3.23</td>
<td>3.74</td>
<td>-0.296</td>
</tr>
</tbody>
</table>
Furthermore, the experimental $q_e$ value of 3.74 was closer to the $q_e$ calculated from the pseudo-second-order kinetic plot ($q_e=4.12$) instead of the one calculated from the pseudo-first-order kinetic plot ($q_e=2.91$) as shown in Table 8.

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>$q_e$</th>
<th>$q_t$</th>
<th>$t/q_t$</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>3.74</td>
<td>0</td>
<td>$\infty$</td>
</tr>
<tr>
<td>10</td>
<td>3.74</td>
<td>3.09</td>
<td>3.23</td>
</tr>
<tr>
<td>20</td>
<td>3.74</td>
<td>3.23</td>
<td>6.18</td>
</tr>
<tr>
<td>30</td>
<td>3.74</td>
<td>3.74</td>
<td>8.02</td>
</tr>
<tr>
<td>40</td>
<td>3.74</td>
<td>3.86</td>
<td>10.3</td>
</tr>
</tbody>
</table>

The data extracted from the plots shown in Table 9 indicate that the trypan blue dye’s adsorption from aqueous solution perfectly follows the pseudo-second-order kinetic model. Since the data support that the mechanism is typically a pseudo-second-order adsorption mechanism, the trypan blue sorption process’s overall rate appears to be controlled by chemical processes as opposed to physical ones.

<table>
<thead>
<tr>
<th>Concentration (mg/L)</th>
<th>$q_e$ experimental</th>
<th>$q_e$ calculated</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>57.648</td>
<td>3.74</td>
<td>2.91</td>
<td>0.839</td>
</tr>
</tbody>
</table>

Table 9. Pseudo first and second order ‘Models’ parameters for the adsorption of trypan blue by avocado seed powder.
4. Conclusions

This study demonstrated that avocado seed powder is an ideal and optimum biosorbent that can be used to remove trypan blue dye from polluted wastewater. The experimental results revealed that biosorption depends mainly on the contact time, initial dye concentration, and adsorbent dosage. However, it also might have been dependent on other variables like temperature and pH. The latter two variables weren't included in this study since ties didn’t permit, but for future work studying their effects on the adsorption efficiency of the adsorbent would prove very insightful.

The isotherm data plotted indicate that the Langmuir isotherm model fitted the trypan blue dye's removal from aqueous solution better with a correlation coefficient of 0.9948 as opposed to 0.9876, the one obtained from plotting the Freundlich isotherm. Additionally, RL's separation factor was lying between zero and one at a value of 0.464, indicating a favorable adsorption process according to the Langmuir model. Since the Langmuir isotherm model fitted the data better, therefore, the system exhibits monolayer biosorption on a homogenous surface.

The detailed kinetic studies confirmed that the sorption rate is a pseudo-second-order model with a higher correlation coefficient of 0.9916. The \( q_e \) calculated from the pseudo-second-order kinetic plot (\( q_e = 4.12 \)) was closer to the experimental \( q_e \) value of 3.74 g/mg than the calculated \( q_e \) value from the pseudo-first-order kinetic model plot.

Additionally, since the pseudo-second-order adsorption mechanism was predominant, the trypan blue sorption process's overall rate appears to be controlled by chemical processes as opposed to physical ones.

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Conflicts of Interest

The authors declare no conflict of interest.

References

orbed Water.

ization.

he continuous synthesis of Pd supported on Fe3O4 nanoparticles: A highly effective and

d Suzuki cross
coupling reaction.

Igarashi, H.; Uchida, H.; Watan

Catalysts

Mandali, P.K.; Chand, D.K. Palladium nanoparticles catalyze

154,
catalyst for carbon(sp2) carbon(sp2) cross

coupling reactions.


