

# Exploring the Role of Melatonin in Meditation on Cardiovascular Health

Uma Priya Mohan <sup>1</sup> , Selvaraj Kunjiappan <sup>2</sup> , Tirupathi Pichiah P.B. <sup>3</sup> , Ewa Babkiewicz <sup>4</sup> , Piotr Maszczyk <sup>4</sup> , Sankarganesh Arunachalam <sup>1,\*</sup> 

<sup>1</sup> Center for Cardiovascular and Adverse Drug Reactions, Department of Biotechnology, School of Bio and Chemical Engineering, Kalasalingam Academy of Research and Education, Krishnankoil, Virudhunagar Dt., Tamilnadu, India - 626126

<sup>2</sup> Department of Biotechnology, School of Bio and Chemical Engineering, Kalasalingam Academy of Research and Education, Krishnankoil, Virudhunagar Dt., Tamilnadu, India -626126

<sup>3</sup> Department of Animal Science, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India - 620024

<sup>4</sup> Department of Hydrobiology, Faculty of Biology, University of Warsaw at Biology & Chemistry Research Center, 02-189-Warsaw, Poland

\* Correspondence: [sankarganesh@gmail.com](mailto:sankarganesh@gmail.com) (S.A);

Scopus Author ID 35209520500

Received: 22.11.2021; Accepted: 22.12.2021; Published: 2.02.2022

**Abstract:** Cardiovascular diseases are the leading cause of disease burden globally. Sleep and cardiovascular connection represent a two-way lane. Recently, many reports have suggested that meditation practices have a beneficial effect on cardiovascular diseases. But the exact mechanism was not known. Several reports suggest that meditation induces the secretion of melatonin. The rhythm of melatonin follows a sleep pattern. Thus, the present hypothesis correlates the plausible mechanisms involved in meditation with enhancing cardiac health through melatonin synthesis. An altered modern lifestyle decreases the level of melatonin which disrupts the circadian rhythm, and subsequently, there is a high incidence of cardiovascular diseases. The disrupted cardiac energy metabolism is distorted due to altered circadian rhythm with elevated ROS. The increased level of ROS activates the inflammatory mediators' cytokines and damages the DNA, resulting in altered cardiac physiology. Melatonin regulates the circadian rhythm and acts as a silent regulator for the cardiac energy balance. Melatonin is the central player of circadian rhythm, and it protects cardiomyocytes by acting as an antioxidant, anti-inflammatory mediator, and repairing DNA damage. Meditation induces melatonin and improves cardiac health through the aforementioned mechanisms.

**Keywords:** cardiovascular diseases; meditation; melatonin; ROS; DNA repair; circadian rhythm; Glucose; lipid metabolism.

© 2022 by the authors. This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

### 1.1. Cardiovascular disease and meditation.

Cardiovascular diseases (CVD) remain one of the most common causes of mortality globally among non-communicable diseases [1–3]. Compared to the high-income countries, CVD mortality in low- and middle-income countries is extremely high due to limited access to effective preventive and management programs [4–9]. In these settings, inexpensive intervention and lifestyle changes can prevent CVD development and improve outcomes.

From the wake of this century, yoga and meditation have attracted more and more attention as effective interventions to improve health. Meditation has beneficial effects on various organs, including the heart [10]. A practice of 12-week mindfulness meditation training for heart patients under usual care displayed significant improvement in blood pressure, heart rate, respiratory rate, NT-pro BNP level, and the results of 6 minutes walk test [11,12]. Meditation practices improve the rehabilitation process in patients with cardiovascular diseases [13,14]. American Heart Association advocates meditation as an adjunct therapy to enhance cardiac health [15]. The present article aims at comprehending the possible mechanisms through which meditation protects cardiac functions.

### *1.2. Meditation and melatonin.*

Meditation is defined as a condition of contemplation, concentration, and reflection [16–18]. The practice has a thousand-year history of improving spiritual and emotional well-being by achieving physical relaxation, inner calmness, and psychological balance by the participator, examining their thoughts and feelings. Mantra meditation, concentrating on a particular subject/point (such as heartfulness meditation, *kundalini* meditation), and mindfulness meditation are just examples of meditation types being practiced nowadays [19–21]. The main goal of meditation is to create awareness of a given moment and forget the past bad moments in a non-judgmental fashion [22,23]. Yoga is a set of techniques, including postures, breath control, and meditation originated in India. Tai chi and qigong, traditional Chinese martial arts and medicine, also include meditation practices [24,25].

In the past two decades, a growing number of research reports suggested that meditation practices cure stress, pain, and anxiety-related conditions through psychological intervention [26–29]. Meditation is helpful as an antidote to both physiological and mental stress. Psychological stress is experienced as encountering obstacles to fulfilling an individual's requirements and aspirations and a perceived threat. The body responds to stress with increased heart rate, blood pressure, breath rate, sweating, weakened immunity, and blood clotting. In a meditative state, the human body switches into a state of restful awareness. The body responds with decreased heart rate, normalization of blood pressure, quiet breathing, reduced stress hormone production, strengthened immune system, reduced sweating, and improved blood flow. These observations suggest that meditation has a dramatic long-term structural effect on the body [30–33]. Recent findings demonstrate that meditation increases melatonin levels [10,34–36]. Meditation raises melatonin levels by delaying the hormone's hepatic metabolism or increasing its production in the pineal gland [37]. Melatonin receptors have been found all through the cardiovascular system, including in various vascular tissues [38,39]. Platelet aggregation, nocturnal hypertension, and serum catecholamine levels have all been found to be reduced by exogenous melatonin [40]. Thus, the influence of melatonin on cardiovascular risk factors via meditation is discussed in this review, as well as current advances in our understanding of meditation and melatonin's effects on cardiovascular illnesses.

## **2. Melatonin and Circadian Rhythm**

Meditation practices synchronize the hypothalamo-pituitary-adrenal (HPA) axis and normalize the levels of cortisol and catecholamine [41,42]. Moreover, meditation increase the level of dehydrosterone [43] and anterior hypophyseal hormones such as growth hormone, thyrotropin-releasing hormone (TRH), prolactin, and melatonin [44].

Melatonin has a crucial function in the physiological control of sleep, both in blind and sighted individuals [45]. The rhythm of melatonin follows a sleep pattern [46]. Melatonin exerts hypnotic properties by inhibiting the suprachiasmatic nucleus [47] and promoting peripheral vasodilation to hypothermic reaction. Melatonin is typically used to treat sleep cycle disruptions attributable to jetlag and insomnia [48]. Melatonin is not just an antioxidant and immunomodulatory agent [49]; it also acts as oncostatin and improves well-being [45]. Aging decreases the release of melatonin and thus influences sleep quality in the elderly. It was shown that meditation practices increase melatonin, serotonin, and noradrenaline levels. Meditation practices increase melatonin by restricting its hepatic metabolism or increasing pineal gland synthesis. Considering the melatonin function in sleep management, meditation activities may improve sleep quality by increased release of melatonin [50].

Sleep has been correlated with lowered heart rate, blood pressure, breath rate and rhythm, oxygen intake, fear or excitations, and reduced basal metabolic level [50]. In cardiovascular conditions, decreased level of circulating melatonin has been observed. On the other hand, the increased melatonin level protects the heart from other cardiovascular diseases in several mechanisms [51]. Until now, the mechanism of melatonin-mediated protection of cardiac function has not been elucidated. This review focuses on deriving the underlying mechanism of melatonin's protective effect through meditation.

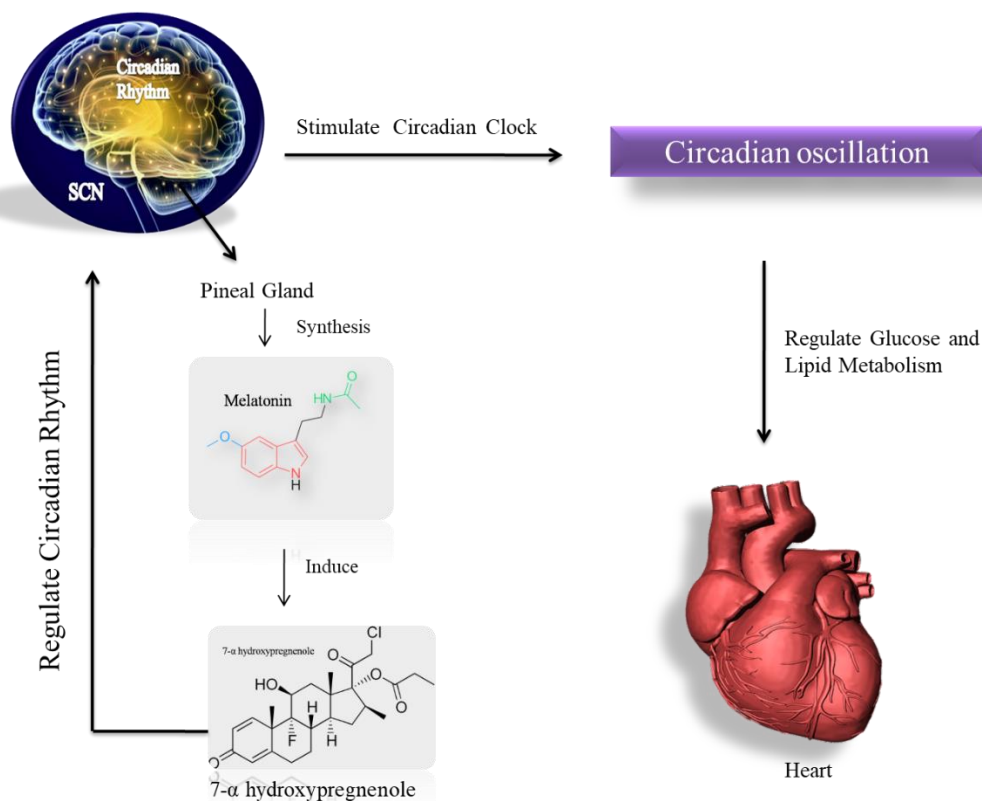
### *2.1. Role of circadian rhythm in cardiac metabolism correlated with melatonin.*

The cardiovascular system exhibits circadian rhythmicity with their acrophases in different parts of the 24-hour cycle [52,53]. The regular synchronization of circadian rhythm signals a biological system's functioning in a coordinated manner [52,54]. The circadian clock's timing is regulated by a signal obtained from the suprachiasmatic nucleus (SCN) through the retina [55,56]. The circadian rhythm is regulated by the clock gene [57,58]. Any disturbance in the clock gene renders impairment in the cardiovascular system [59]. A change in blood pressure synchrony with the circadian rhythm was observed in hypertensive rat strains [59,60].

In humans, circadian rhythm regulates carbohydrate metabolism, essential for glucose homeostasis and energy balance. The disparity between glucose and insulin in tissues and blood cells causes various disorders, including metabolic syndrome, obesity, type 2 diabetes, and cardiovascular diseases [61]. Reaven reported that insulin resistance and elevated circulating postprandial TAG concentration cause cardiovascular diseases [62]. For example, the individuals working in shifts at late hours who consume their food late night have relative glucose and lipid intolerance [63]. Among the workers working night shifts, elevated levels of circulating TAG are observed irrespective of their energy and nutrient intake because of the disrupted circadian rhythm. The increased level of TAG promotes cardiovascular disease [64]. Comparatively, night-time workers have an elevated chance of having cardiovascular diseases than day-time workers [65].

Melatonin is secreted when the eyes do not receive light, and they accelerate the production of  $7\alpha$ -hydroxypregnenolone. Understandably, the regulation of  $7\alpha$ -hydroxypregnenolone synthesis is central to animal circadian rhythms [55,56]. Circadian change in spontaneous locomotor activity concerning the  $7\alpha$ -hydroxypregnenolone synthesis and melatonin secretion were evaluated [55,56]. A strong association between circadian rhythm and metabolism has been well established. [66–68]. The circadian rhythm regulates glucose homeostasis and energy balance through insulin [61].

Melatonin can influence the circadian rhythm in the cardiovascular system by following pathways such as i) central oscillator in the SCN; ii) the sympathetic output to the heart and vessels; iii) interactions with other hormonal systems involved in cardiovascular system regulation [57]. Thus, melatonin protects the cardiovascular system through its influence on circadian rhythm. Continuous practices of meditation may influence melatonin production. Overall, it is apparent that melatonin protects the heart by regulating circadian rhythm (Figure 1).



**Figure 1.** Role of melatonin in circadian rhythm. Regulation of circadian rhythm via melatonin synthesis correlated with glucose and lipid metabolism.

### 3. Role of Melatonin in Cardiac Energetic Metabolism

The energy needs demand of a healthy heart is primarily met by fatty acids (80%) and the rest from glucose (20%). At rest, cardiomyocytes use 15-20% of maximal oxidative capacity [69]. Any metabolic disturbance triggers cardiac dysfunction. Altered cardiac energy metabolism impairs ATP synthesis [70], gradually increasing cardiac failure risk [71]. On the other hand, dyslipidemia is a significant risk factor for cardiovascular diseases [72]. Melatonin decreases low-density lipoprotein (LDL) level and body weight in high-fat diet-induced non-alcoholic fatty liver disease mice [72].

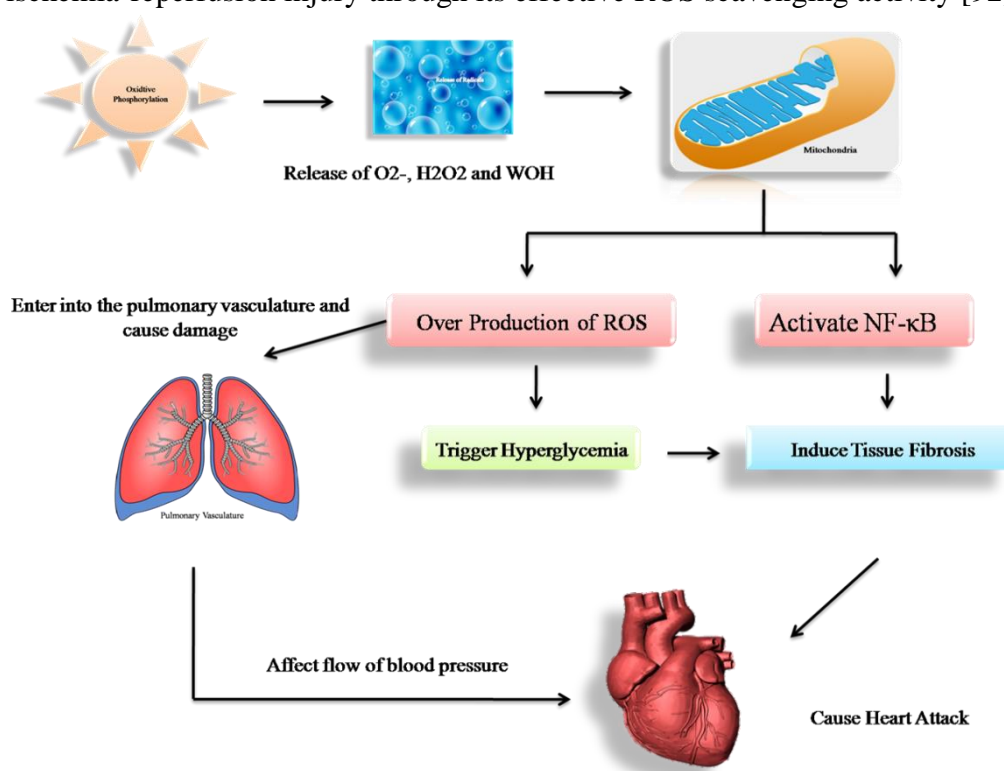
Studies have reported that melatonin influences carbohydrate metabolism [73,74] in pinealectomized animals. It upregulates the expression levels of enzymes involved in lipolysis,  $\beta$ -oxidation, and mitochondrial biogenesis-related genes [74]. Thus, melatonin maintains an adequate energy balance by regulating the energy flow and expenditure [15].

Glucose and lipid metabolism are linked together in several ways. There is a crosstalk between melatonin and insulin signaling [75]. Melatonin promotes glucose transport glycogen synthesis, inhibits lipolysis, regulates body weight and glucose metabolism through the phosphorylation of IRS-1 under insulin [56,76]. Membrane-bound melatonin receptors, MT1 and MT2, reduce the intracellular cAMP level and control cAMP-dependent phosphotyrosine

phosphatase activity, thereby accelerating insulin phosphorylation [76]. This process implies strong evidence regarding the protective effect of melatonin by balancing glucose and lipid metabolism. The alteration in the carbohydrate metabolism induced by glucose intolerance and insulin resistance may be reversed by melatonin [77]. It is one of the functions to maintain the energy balance in the heart.

#### 4. Role of Melatonin in Oxidative Stress

In diabetic complications of microvascular and cardiovascular systems, oxidative stress plays a vital role in promoting the disease. Oxidative phosphorylation plays a crucial role in oxidative stress through electron leakage in the form of  $O_2^-$  [78],  $H_2O_2$ , and  $HO$  [79] and triggers apoptosis [80]. Also, the overproduction of mitochondrial ROS triggers hyperglycemia and causes tissue damage [80] by activating nuclear factor kappa B (NF- $\kappa$ B) [81], which further leads to tissue fibrosis, induced by inflammation [82]. Also, excessive ROS disrupts mitochondrial function by activating the release of inflammatory cytokines, growth factors, elastases, and vasoconstrictors [83–85]. These factors activate other ROS sources such as NADPH oxidases, cyclooxygenases, and lipoxygenases, which further increase ROS production in the pulmonary vasculature causing pulmonary vascular endothelial damage [86–88], leading to reduced blood pressure in the heart. A decade ago, Ianas *et al.* first reported melatonin's free radical scavenging activity [89]. It also stimulates the antioxidants to directly neutralize the free radicals, reactive oxygen, and nitrogen species [90]. The hormone can also reduce nocturnal blood pressure [91]. Paul and Simko reported that melatonin guards the heart against ischemia-reperfusion injury through its effective ROS scavenging activity [92].



**Figure 2.** Mechanism of oxidative stress in cardiac function.

Mitochondrial nitric oxide synthase (NOS) plays a vital role in the formation of reactive nitrogen species (RNS), which induces cellular damage and causes cardiovascular risk [93]. The lack of nitric oxide scavenging could adversely affect [94]. The function of mitochondria complex I and IV are influenced by melatonin [95,96]. Melatonin intensifies the activity of



endogenous antioxidative enzymes [97] and neutralizes the nitric oxide, hydrogen peroxide, singlet oxygen, peroxy nitrite anion, and hypochlorous acid [90] or directly scavenges NO and ONOO or inhibits the synthesis of NOS [98–100]. Melatonin also stimulates antioxidant enzymes such as superoxide dismutase, glutathione peroxidase, and glutathione reductase [90].

The metabolites of melatonin, such as N1-acetyl-N2- formyl-5-methoxy kynuramine and N-acetyl-5-methoxykynuramine, also have a potent antioxidant activity [99,101,102]. Due to scavenging activity on OH, a highly toxic ROS, counteract lipid peroxidation [103,104] (Figure 2). Thus, directly and indirectly, melatonin decreases oxidative stress by antioxidant enzymes and reduces cardiovascular diseases. Overall, it is clear that melatonin and the regulation of circadian rhythm promote cardiac health by reducing oxidative stress.

#### *4.1 The role of melatonin in anti-inflammatory activity correlated with oxidative stress in the cardiovascular system.*

In oxidative stress, the H<sub>2</sub>O<sub>2</sub> production in human chondrocytes induces the release of IL-1b, IL-8, CXCR-4, TXNIP, STS, and IFI-6-16 [105]. Nian *et al.* reported that the cytokines such as TNF $\alpha$  and interleukin-6 (IL-6) are involved in myocardial ischemic injury and regulate the myocyte survival or apoptosis of the cellular inflammatory response [106]. In cardiac failure, IL-1 signaling plays a negative role by repressing the contractility of the heart, stimulating myocardial hypertrophy, and prompting apoptosis of cardiomyocytes [107].

Some physiological and psychological stresses trigger inflammation leading to chronic inflammatory diseases [108,109]. The overexpression of CXCR4 in cardiomyocytes might stimulate the inflammatory cells, increase TNF- $\alpha$  production, and induce cell death/apoptosis [110]. The activation of CXCR4 interacts with beta-adrenergic receptors and stimulates downstream signaling. CXCR4 plays a critical role in neuro-humoral regulation of the heart and the progression of heart failure [111]. Chronic inflammation may lead to apoptosis and myocardial remodeling [112]. Overall, myocardial damage is further intensified by inflammation (Figure 3).

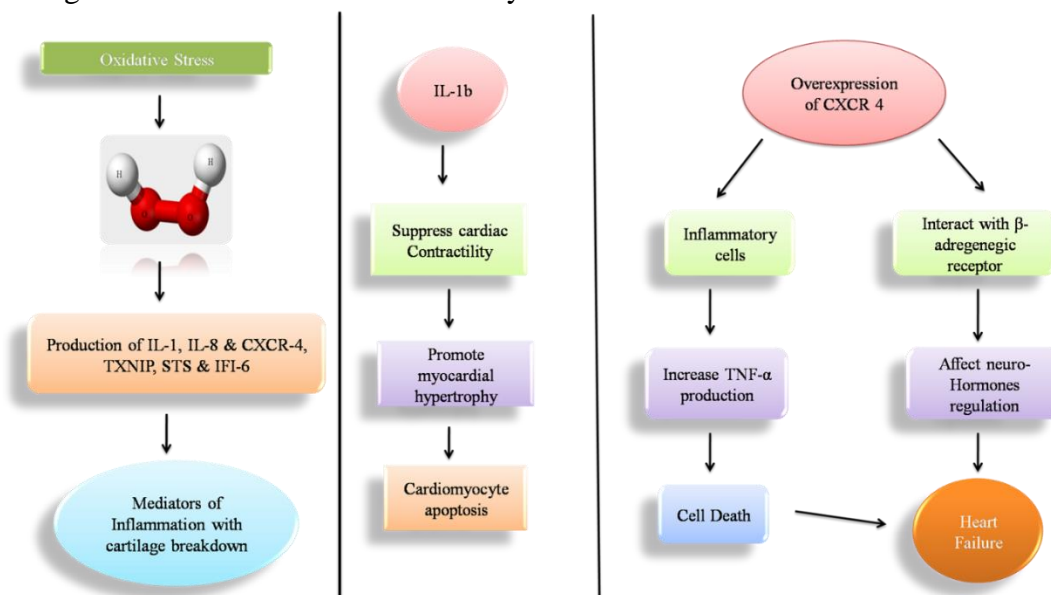
The expression of specific cytokines (iNOS, COX-2, TNF- $\alpha$ , and IL-6) and pro-inflammatory mediators is coordinated by NF- $\kappa$ B [113]. Though in some instances, NF- $\kappa$ B acts as a cardioprotective mediator, in acute hypoxia and reperfusion injury, chronic activation of NF- $\kappa$ B promotes heart failure through its downstream signals. NF- $\kappa$ B signaling triggers chronic inflammation and pro-inflammatory cytokines release (iNOS, COX-2, TNF- $\alpha$ , and IL-6). It may lead to stress response in the endoplasmic reticulum resulting in cardiomyocyte death [114]. The association between elevated circulating chemokine levels and cardiac dysfunction has been well established [115–117].

Stress modulates the expression of immune response genes through the central nervous system (CNS) via the effects of hormones and neurotransmitters on the gene transcription control pathway [118]. In psychological stress, melatonin suppresses norepinephrine and epinephrine expression levels in rodents [119–122]. In addition, reduced melatonin level leads to elevated oxidative stress and the release of inflammatory mediators. Oxidative stress-mediated cytotoxicity and up-regulation of inflammatory mediators were observed in cases of decreased melatonin levels [112]. The anti-inflammatory effect of melatonin is mediated through counteracting the inflammatory process by free radical scavenging and activation of the endogenous antioxidant defense machinery [123–128].

Melatonin down-regulates the expression of SIRT1, which elicits anti-inflammatory activity [129]. Melatonin blocks hydrogen peroxide-induced phosphorylation of PI3K/Akt,

p38, ERK, JNK, and MAPK and the activation of NF- $\kappa$ B, which is reversed by sirtinol and SIRT1 siRNA. Thus, NF- $\kappa$ B does not directly induce or promote cardiac failure by eliciting the downstream signals. Melatonin decreases the expression of iNOS and COX-2 and also the production of NO and PGE2 [129].

Melatonin has been shown to modulate the immune system by regulating cytokines [130]. Melatonin promotes immune-stimulatory effects on several immune parameters, such as antibody-dependent cellular cytotoxicity [131,132]. Melatonin reduces the synthesis of TNF- $\alpha$ , IL-1 $\beta$ , IL-8 release, and CXCR-4, TXNIP, STS, and IFI-6-16 [129]. Thus, they do not increase the neuro-humoral regulation in the heart and the progression of heart failure. It was reported that melatonin acts as an antioxidant and exerts numerous anti-inflammatory functions [133–137]. In consequence, melatonin plays a critical role in heart neuro-humoral regulation and the progression of heart failure. Melatonin blocks the production of H<sub>2</sub>O<sub>2</sub> and exerts its anti-inflammatory activity in maintaining the metabolic activity of the cardiovascular system [138,139]. The aforementioned process implies that melatonin acts as a cardioprotective agent by reducing oxidative stress and inflammatory mediators such as NF- $\kappa$ B.



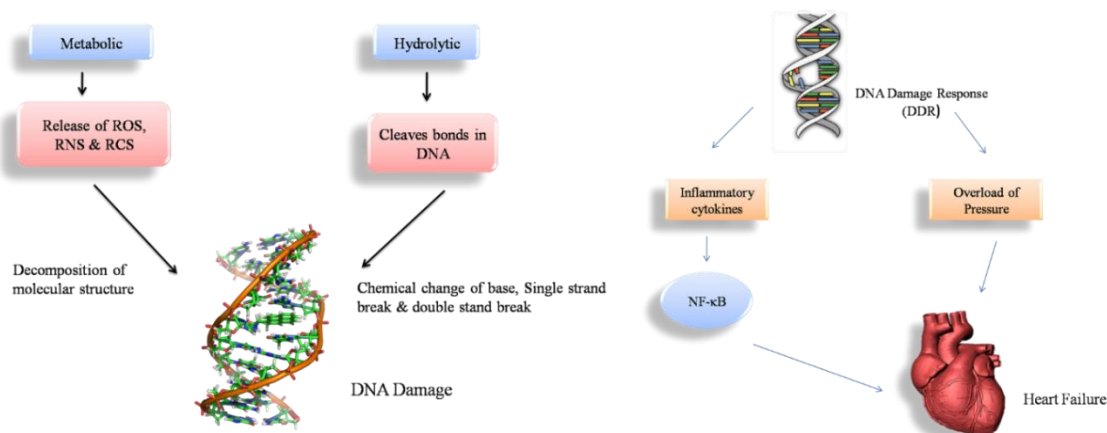
**Figure 3.** Mechanism of the inflammatory mediator in cardiac function. Overexpression of IL-1b and CXCR4 affect cardiac function leads to cell apoptosis. IL-1b – Interleukin and CXCR4 – Chemokine Receptor 4, TNF $\alpha$ – Tumour Necrosis Factor  $\alpha$

## 5. Role of Melatonin in DNA Damage Repair

Single-stranded DNA damage-induced response is also involved in heart failure. In general, DNA damage can occur due to metabolic and hydrolytic processes. In the metabolic process, the release of reactive oxygen, nitrogen and carbonyl species, lipid peroxidation products, and alkylating agents cause DNA damage [140,141]. Reports show that ROS causes DNA damage at a frequency of at least 10,000 times per cell per day in humans [142,143].

In the hydrolytic process, the modification of the molecular structure of DNA causes disturbance in the function of the heart as the unpaired DNA single-strand break (SSB) activates DNA Damage Response (DDR) and also the inflammatory cytokine expression through NF- $\kappa$ B signaling [140]. The activation of DDR causes the pathogenesis of heart failure triggered by pressure overload [140] (Figure 4). Minamino *et al.* observed that DDR activation in cardiomyocytes occurs in patients with end-stage heart failure. They also reported that excess pressure causes heart failure in mice [144].

Melatonin protects DNA from damage and oxidation by balancing oxidant-antioxidant balance, inhibiting neutrophil infiltration, and reducing the 8-OHdG level [145]. The hormone plays a pivotal role in maintaining cardiac metabolism. Melatonin stimulates antioxidant enzymes such as superoxide dismutase (SOD), glutathione peroxidase (GSHPx) [146–148], and glutathione reductase (GR) to neutralize or directly scavenge the free radicals [149,150]. Thus, by reducing the DNA damage induced by oxidative stress, melatonin prevents pathological conditions in the heart. Melatonin neutralizes the free radicals causing DNA damage and gives protection through inactivating the DNA damaging agents [151,152]. Melatonin potentiates the DNA repair capacity against strand breaks caused by DNA damaging agents [153,154].



**Figure 4.** Mechanism of the DNA repair in cardiac function.

## 6. Hypothesis

Meditation has well been documented to improve cardiac health. Cardiomyocytes derive 80% energy from lipids such as fatty acid and 20% from other sources such as glucose, ketone bodies, etc. Glucose and lipid homeostasis is maintained by circadian rhythm in the heart. Melatonin synthesized from the pineal gland regulates the circadian rhythm. In addition to maintaining circadian rhythm, melatonin also regulates glucose and lipid metabolism.

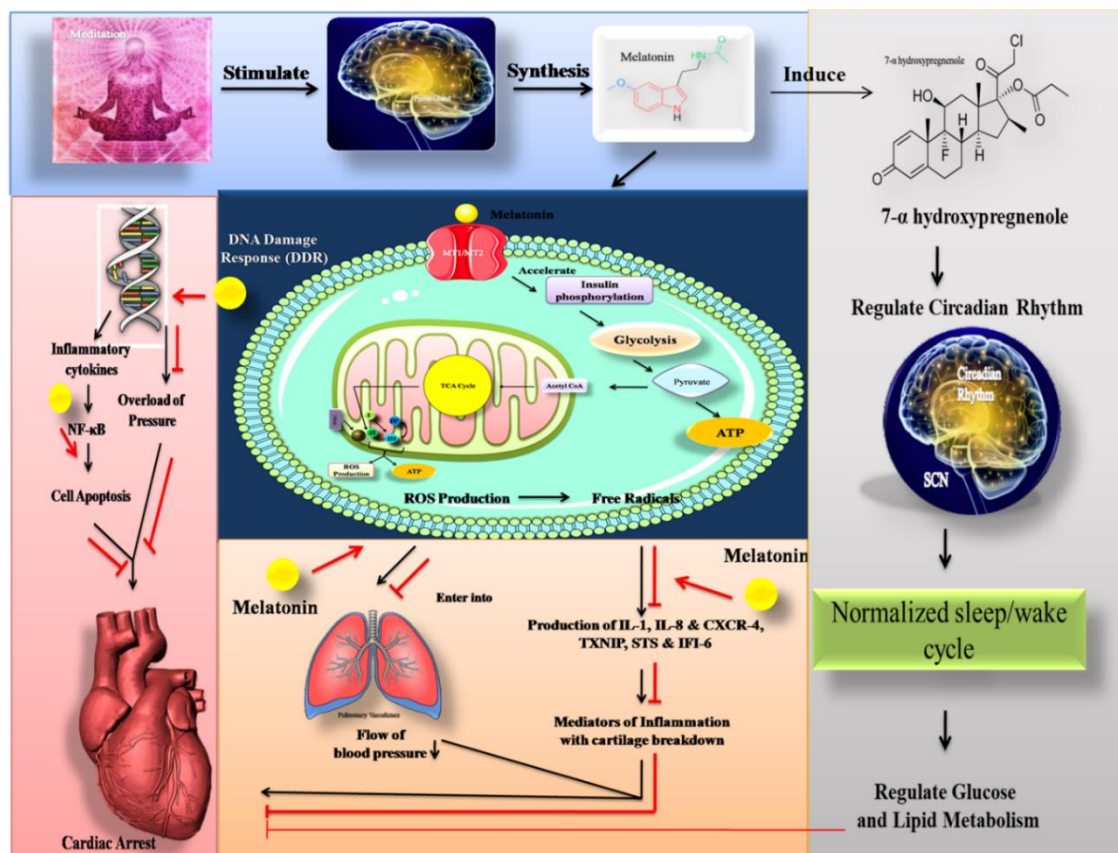
Melatonin receptors are G protein-coupled receptors with two subtypes, MT<sub>1</sub> and MT<sub>2</sub>, regulating glucose metabolism through phosphorylating insulin as crosstalk with insulin and accelerating insulin phosphorylation glycolysis for glucose homeostasis cytoplasm. CoQ neutralizes the free radicals formed during oxidative phosphorylation. Interestingly, the expression of CoQ is regulated by PINK1, whose expresses is dependent on melatonin.

Elevated ROS caused by O<sub>2</sub><sup>-</sup>, H<sub>2</sub>O<sub>2</sub>, and WHO upon entering the pulmonary artery causes pulmonary endothelial damage and reduces the blood flow and blood pressure, ultimately resulting in cardiac dysfunction. On the other hand, the increased production of ROS also causes DNA damage. The DNA damage causes pressure overload and congestive heart failure. Free radicals also trigger the IL-1, IL-8, CXCR-4, TXNIP, STS, and IFI-6, which mediates cartilage breakdown. They also activate the NFκB, which acts as an anti-inflammatory agent, but prolonged exposure causes cell apoptosis leading to a decreased heart size. Melatonin also acts as DNA repairing agent repairing the DNA damage in cardiomyocytes and preventing subsequent inflammatory reactions.

Collectively, melatonin, in addition to being the regulator of circadian rhythm, acts as an antioxidant, anti-inflammatory, DNA repair agent. Further, melatonin also regulates glucose and lipid metabolisms. A number of reports suggest that the melatonin level is increased during



meditation. Independently, meditation has been shown to promote cardiac health. Therefore, we hypothesize that meditation-mediated improvement in cardiac health is through the action of melatonin. (Figure 5).



**Figure 5.** Schematic diagram depicting the possible mechanism of meditation-mediated improvement in cardiac health through melatonin. Meditation stimulates the pineal gland to synthesize the melatonin and subsequent induction the 7- $\alpha$  hydroxypregnenolone to regulate the circadian rhythm. The circadian rhythm regulates cardiac glucose and lipid homeostasis. In an altered modern lifestyle, circadian rhythm is disrupted, and consequently, there is a high occurrence of cardiovascular diseases. The altered circadian rhythm with elevated ROS disrupt glucose and lipid homeostasis. The increased level of ROS activates the inflammatory mediators' cytokines and damages the DNA, resulting in altered cardiac physiology. Melatonin is the central player of circadian rhythm, and it protects cardiomyocytes by acting as an antioxidant, anti-inflammatory mediator. Melatonin also repairs DNA damage. The red color refers to the inhibition of the mechanism. MT1/MT2 – Melatonin receptor 1 & 2; TCA – Tricarboxylic acid; ROS – Reactive Oxidative Species; IL-1, IL-8 – Interleukin 1 & 8; CXCR – Chemokine receptor; TXNIP - Thioredoxin Interacting Protein; STS – Steroid Sulfatase; IFI-6 – Interferon 6; NF- $\kappa$ B - Nuclear Factor kappa-light-chain enhancer of activated B cells. Red color lines indicate inhibition. The blue color background implies meditation regulates melatonin. The brown color background implies a Circadian rhythm. Dark blue implies – Mitochondrial function (Glucose and lipid metabolism and Antioxidants). Light Orange color implies – Anti-inflammatory. Light pink color Implies – DNA Damage.

## 7. Conclusions

Sleep and cardiovascular connection are two-way lanes. With heart disease, someone might have other health issues, including sleep disorders. Likewise, heart disease signs may worsen by sleep issues, such as obstructive sleep apnea (OSA) and insomnia. It is necessary to sleep a decent night, whether or not your heart is stable. Sleep improves both the heart and energy, thought abilities, and fitness. People will feel more pressure from the core if they are willing to cope with the sleep issues.

Owing to the frequent rising of sleep apnea patients, they have inadequate quality sleep and remain tired all day long. They may have impaired cardiovascular function as well. Sleep dysfunction is 47-83 percent, 35 percent, and 12-53 percent of those with coronary disease, auric fibrillation (heart rhythm disturbances, and stroke). The cardiac condition is exceptionally susceptible. Researchers report that sleep apnea that is not treated is one to five times more likely to die from heart failure. The meditation on sleep is a rare, directed experience that provides all-around sleep relief alone and helps us let go of the day — all that happened and all that was said — to relax the mind while relaxing the body at the same time. Scientifically speaking, meditation helps decrease the heart rate by ignition and relaxing breathing, improving the chances of a quality night's sleep.

Cardiovascular diseases (CVD) are increasing the risk factor worldwide. Nowadays, yoga and meditation pay considerable attention to the defense of CVD. But the scientific mechanism of meditation behind CVD protection was not known. Meditation strategies have been documented to improve some amounts of HPA. Data from the American Journal of Practice were taken from the report. Studies have shown that melatonin impacts ischemia-reperfusion, transient myocardial hypoxia, pulmonary hypertension, elevated blood pressure, valvular cardiac failure, artery disorders, and lipid metabolism.

Although melatonin's function in the sense of heart failure has been studied in a few clinical trials, recent laboratory research results support the possible usage of melatonin in heart failure as preventive and adjunctive curative therapy. Melatonin could be a promising treatment alternative for cardiovascular disorders as cheap and well-accepted medicine. Recently several reports suggested that the continuous practice of meditation increases the production of melatonin, a hormone. Based on these, we derived the plausible meditation mechanism, which protects the heart from CVD. In these, we revealed the connection between meditation and melatonin and the role of melatonin in cardiovascular protection. Melatonin regulates the circadian rhythm, but it can also act as an anti-inflammatory, antioxidant agent and have a role in glucose and lipid homeostasis and DNA repair mechanisms because these are the reason which leads to cause cardiovascular diseases. Melatonin, which produces during the night only due to the modern lifestyle, decreased melatonin production. Recently shreds of evidence from the science community revealed melatonin production during the meditation. Thus, we conclude that melatonin production during meditation protects the heart from CVD. Further, we need to prove our hypothesis in real-time experiments.

## **Funding**

The study was financially supported by the Department of Science and Technology, Government of India under the SATYAM scheme (Project file No. DST/SATYAM/2017/119(G)).

## **Acknowledgments**

The study was financially supported by the Department of Science and Technology, Government of India under the SATYAM scheme (Project file No. DST/SATYAM/2017/119(G)).

## **Conflicts of Interest**

The authors declare no competing financial interest and no conflicts of interest.

## References

1. Naghavi, M.; Abajobir, A.A.; Abbafati, C.; Abbas, K.M.; Abd-Allah, F.; Abera, S.F.; Aboyans, V.; Adetokunboh, O.; Afshin, A.; Agrawal, A. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* **2017**, *390*, 1151–1210, [https://doi.org/10.1016/S0140-6736\(17\)32152-9](https://doi.org/10.1016/S0140-6736(17)32152-9).
2. Wang, H.; Abajobir, A.A.; Abate, K.H.; Abbafati, C.; Abbas, K.M.; Abd-Allah, F.; Abera, S.F.; Abraha, H.N.; Abu-Raddad, L.J.; Abu-Rmeileh, N.M.E. Global, regional, and national under-5 mortality, adult mortality, age-specific mortality, and life expectancy, 1970–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* **2017**, *390*, 1084–1150, [https://doi.org/10.1016/S0140-6736\(17\)31833-0](https://doi.org/10.1016/S0140-6736(17)31833-0).
3. Vos, T.; Abajobir, A.A.; Abate, K.H.; Abbafati, C.; Abbas, K.M.; Abd-Allah, F.; Abdulkader, R.S.; Abdulle, A.M.; Abebo, T.A.; Abera, S.F. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* **2017**, *390*, 1211–1259, [https://doi.org/10.1016/S0140-6736\(17\)32154-2](https://doi.org/10.1016/S0140-6736(17)32154-2).
4. Ralston, J.; Reddy, K.S.; Fuster, V.; Narula, J. Cardiovascular diseases on the global agenda: the United Nations high level meeting, sustainable development goals, and the way forward. *Glob. Heart* **2016**, *11*, 375–379, <https://doi.org/10.1016/j.gheart.2016.10.029>.
5. Joshi, P.; Islam, S.; Pais, P.; Reddy, S.; Dorairaj, P.; Kazmi, K.; Pandey, M.R.; Haque, S.; Mendis, S.; Rangarajan, S. Risk factors for early myocardial infarction in South Asians compared with individuals in other countries. *Jama* **2007**, *297*, 286–294, <https://doi.org/10.1001/jama.297.3.286>.
6. Xavier, D.; Pais, P.; Devereaux, P.J.; Xie, C.; Prabhakaran, D.; Reddy, K.S.; Gupta, R.; Joshi, P.; Kerkar, P.; Thanikachalam, S. Treatment and outcomes of acute coronary syndromes in India (CREATE): a prospective analysis of registry data. *Lancet* **2008**, *371*, 1435–1442, [https://doi.org/10.1016/S0140-6736\(08\)60623-6](https://doi.org/10.1016/S0140-6736(08)60623-6).
7. Yusuf, S.; Rangarajan, S.; Teo, K.; Islam, S.; Li, W.; Liu, L.; Bo, J.; Lou, Q.; Lu, F.; Liu, T. Cardiovascular risk and events in 17 low-, middle-, and high-income countries. *N. Engl. J. Med.* **2014**, *371*, 818–827, <https://doi.org/10.1056/NEJMoa1311890>.
8. Assembly, G. *Resolution adopted by the General Assembly on 31 July 2014; A/RES/69/314*, 19 August, 2015;
9. Clark, H. NCDs: a challenge to sustainable human development. *Lancet (London, England)* **2013**, *381*, 510–511, [https://doi.org/10.1016/s0140-6736\(13\)60058-6](https://doi.org/10.1016/s0140-6736(13)60058-6).
10. Tacón, A.M.; McComb, J.; Caldera, Y.; Randolph, P. Mindfulness meditation, anxiety reduction, and heart disease: a pilot study. *Fam. Community Health* **2003**, *26*, 25–33, <https://doi.org/10.1097/00003727-200301000-00004>.
11. Younge, J.O.; Wery, M.F.; Gotink, R.A.; Utens, E.M.W.J.; Michels, M.; Rizopoulos, D.; van Rossum, E.F.C.; Hunink, M.G.M.; Roos-Hesselink, J.W. Web-based mindfulness intervention in heart disease: a randomized controlled trial. *PLoS One* **2015**, *10*, <https://pubmed.ncbi.nlm.nih.gov/26641099/>.
12. Younge, J.O.; Gotink, R.A.; Baena, C.P.; Roos-Hesselink, J.W.; Hunink, M.G.M. Mind–body practices for patients with cardiac disease: a systematic review and meta-analysis. *Eur. J. Prev. Cardiol.* **2015**, *22*, 1385–1398, <https://doi.org/10.1177/2047487314549927>.
13. Delui, M.H.; Yari, M. Comparison of cardiac rehabilitation programs combined with relaxation and meditation techniques on reduction of depression and anxiety of cardiovascular patients. *Open Cardiovasc. Med. J.* **2013**, *7*, 99, <https://dx.doi.org/10.2174%2F1874192401307010099>.
14. Parswani, M.J.; Sharma, M.P.; Iyengar, S.S. Mindfulness-based stress reduction program in coronary heart disease: A randomized control trial. *Int. J. Yoga* **2013**, *6*, 111, <https://dx.doi.org/10.4103%2F0973-6131.113405>.
15. Levine, G.N.; Lange, R.A.; Bairey-Merz, C.N.; Davidson, R.J.; Jamerson, K.; Mehta, P.K.; Michos, E.D.; Norris, K.; Ray, I.B.; Saban, K.L. Meditation and cardiovascular risk reduction: a scientific statement from the American Heart Association. *J. Am. Heart Assoc.* **2017**, *6*, e002218, <https://doi.org/10.1161/JAHA.117.002218>.
16. Hernández, S.E.; Dorta, R.; Suero, J.; Barros-Loscertales, A.; González-Mora, J.L.; Rubia, K. Larger whole brain grey matter associated with long-term Sahaja Yoga Meditation: A detailed area by area comparison. *PLoS One* **2020**, *15*, e0237552, <https://doi.org/10.1371/journal.pone.0237552>.
17. Bushell, W.; Castle, R.; Williams, M.A.; Brouwer, K.C.; Tanzi, R.E.; Chopra, D.; Mills, P.J. Meditation and yoga practices as potential adjunctive treatment of SARS-CoV-2 infection and COVID-19: a brief overview of key subjects. *J. Altern. Complement. Med.* **2020**, *26*, 547–556, <https://doi.org/10.1089/acm.2020.0177>.
18. Moszeik, E.N.; von Oertzen, T.; Renner, K.-H. Effectiveness of a short Yoga Nidra meditation on stress, sleep, and well-being in a large and diverse sample. *Curr. Psychol.* **2020**, 1–15, <https://link.springer.com/article/10.1007/s12144-020-01042-2>.
19. Farias, M.; Maraldi, E.; Wallenkampf, K.C.; Lucchetti, G. Adverse events in meditation practices and meditation-based therapies: a systematic review. *Acta Psychiatr. Scand.* **2020**, *142*, 374–393,

- <https://doi.org/10.1111/acps.13225>.
20. Behan, C. The benefits of meditation and mindfulness practices during times of crisis such as COVID-19. *Ir. J. Psychol. Med.* **2020**, *37*, 256–258, <https://www.cambridge.org/core/journals/irish-journal-of-psychological-medicine/article/benefits-of-meditation-and-mindfulness-practices-during-times-of-crisis-such-as-covid19/076BCD69B41BC5A0A1F47E9E78C17F2A>.
21. Iwamoto, S.K.; Alexander, M.; Torres, M.; Irwin, M.R.; Christakis, N.A.; Nishi, A. Mindfulness meditation activates altruism. *Sci. Rep.* **2020**, *10*, 1–7, <https://www.nature.com/articles/s41598-020-62652-1>.
22. Freeman, G. *Meditation*; Glenda Freeman, 2020. Available online: [https://books.google.co.in/books?hl=en&lr=&id=XIQzEAAQBAJ&oi=fnd&pg=PA8&dq=Freeman,+G.+Meditation%3B+Glenda+Freeman,+2020&ots=y1IvoBHahn&sig=fmFTQVsbSQiusysjaovWTBxXf98&redir\\_esc=y#v=onepage&q&f=false](https://books.google.co.in/books?hl=en&lr=&id=XIQzEAAQBAJ&oi=fnd&pg=PA8&dq=Freeman,+G.+Meditation%3B+Glenda+Freeman,+2020&ots=y1IvoBHahn&sig=fmFTQVsbSQiusysjaovWTBxXf98&redir_esc=y#v=onepage&q&f=false).
23. Vivot, R.M.; Pallavicini, C.; Zamberlan, F.; Vigo, D.; Tagliazucchi, E. Meditation increases the entropy of brain oscillatory activity. *Neuroscience* **2020**, *431*, 40–51, <https://doi.org/10.1016/j.neuroscience.2020.01.033>.
24. Basu Ray, I.; Menezes, A.R.; Malur, P.; Hiltbold, A.E.; Reilly, J.P.; Lavie, C.J. Meditation and coronary heart disease: A review of the current clinical evidence. *Ochsner J.* **2014**, *14*, 696–703, <http://www.ochsnerjournal.org/content/14/4/696.abstract>.
25. Barrós-Loscertales, A.; Hernández, S.E.; Xiao, Y.; González-Mora, J.L.; Rubia, K. Resting State Functional Connectivity Associated With Sahaja Yoga Meditation. *Front. Hum. Neurosci.* **2021**, *15*, 65, <https://doi.org/10.3389/fnhum.2021.614882>.
26. Hofmann, S.G.; Sawyer, A.T.; Witt, A.A.; Oh, D. The effect of mindfulness-based therapy on anxiety and depression: A meta-analytic review. *J. Consult. Clin. Psychol.* **2010**, *78*, 169, <https://psycnet.apa.org/doi/10.1037/a0018555>.
27. Fumero, A.; Peñate, W.; Oyanadel, C.; Porter, B. The effectiveness of mindfulness-based interventions on anxiety disorders. a systematic meta-review. *Eur. J. Investig. Heal. Psychol. Educ.* **2020**, *10*, 704–719, <https://doi.org/10.3390/ejihpe10030052>.
28. Białkowska, J.; Juranek, J.; Wojtkiewicz, J. Behavioral Medicine Methods in Treatment of Somatic Conditions. *Biomed Res. Int.* **2020**, 2020, <https://doi.org/10.1155/2020/5076516>.
29. Hudak, J.; Hanley, A.W.; Marchand, W.R.; Nakamura, Y.; Yabko, B.; Garland, E.L. Endogenous theta stimulation during meditation predicts reduced opioid dosing following treatment with Mindfulness-Oriented Recovery Enhancement. *Neuropsychopharmacology* **2021**, *46*, 836–843, <https://www.nature.com/articles/s41386-020-00831-4>.
30. Krygier, J.R.; Heathers, J.A.J.; Shahrestani, S.; Abbott, M.; Gross, J.J.; Kemp, A.H. Mindfulness meditation, well-being, and heart rate variability: A preliminary investigation into the impact of intensive vipassana meditation. *Int. J. Psychophysiol.* **2013**, *89*, 305–313, <https://doi.org/10.1016/j.ijpsycho.2013.06.017>.
31. Pimple, J.; Agrawal, T. Efficacy of practicing positive psychological interventions, yoga, and mindfulness meditation in COVID-19 lockdown. *Int. J. Indian Psychol.* **2020**, *8*, 293–303, <https://doi.org/10.25215/0802.239>.
32. Babbar, S.; Oyarzabal, A.J.; Oyarzabal, E.A. Meditation and mindfulness in pregnancy and postpartum: a review of the evidence. *Clin. Obstet. Gynecol.* **2021**, *64*, 661–682, <https://doi.org/10.1097/grf.0000000000000640>.
33. de Oliveira, S.S.I.; Gonçalves, S.L.M.; de Mello Weig, K.; Magalhães Filho, T.R.; Martinez, O.E.R.; da Cunha Kalil, M.T.A.; Boggiss, G.P.; Mandarino, D.; Tanganeli, J.P.C.; Almada, T.S. Temporomandibular disorders: Guidelines and self-care for patients during COVID-19 pandemic. *Brazilian Dent. Sci.* **2020**, *23*, 8-p, <https://bds.ict.unesp.br/index.php/cob/article/view/2255>.
34. Liou, C.-H.; Hsieh, C.-W.; Hsieh, C.-H.; Chen, D.-Y.; Wang, C.-H.; Chen, J.-H.; Lee, S.-C. Detection of night-time melatonin level in Chinese Original Quiet Sitting. *J. Formos. Med. Assoc.* **2010**, *109*, 694–701, [https://doi.org/10.1016/S0929-6646\(10\)60113-1](https://doi.org/10.1016/S0929-6646(10)60113-1).
35. Solberg, E.E.; Holen, A.; Ekeberg, Ø.; Østerud, B.; Halvorsen, R.; Sandvik, L. The effects of long meditation on plasma melatonin and blood serotonin. *Med. Sci. Monit.* **2004**, *10*, CR96–CR101, <https://www.medscimonit.com/abstract/index/idArt/11604>.
36. Fernandes, C.A.; Nóbrega, Y.K.M.; Tosta, C.E. Pranic meditation affects phagocyte functions and hormonal levels of recent practitioners. *J. Altern. Complement. Med.* **2012**, *18*, 761–768, <https://doi.org/10.1089/acm.2010.0718>.
37. Massion, A.O.; Teas, J.; Hebert, J.R.; Wertheimer, M.D.; Kabat-Zinn, J. Meditation, melatonin and breast/prostate cancer: hypothesis and preliminary data. *Med. Hypotheses* **1995**, *44*, 39–46, [https://doi.org/10.1016/0306-9877\(95\)90299-6](https://doi.org/10.1016/0306-9877(95)90299-6).
38. Zanoloni, A.; Forni, A.; Zanoloni-Muciaccia, W.; Zanussi, C. Effect of pinealectomy on arterial blood pressure and food and water intake in the rat. *J. Endocrinol. Invest.* **1978**, *1*, 125–130, <https://link.springer.com/article/10.1007/BF03350359>.
39. Baker, J.; Kimpinski, K. Role of melatonin in blood pressure regulation: An adjunct anti-hypertensive agent. *Clin. Exp. Pharmacol. Physiol.* **2018**, *45*, 755–766, <https://doi.org/10.1111/1440-1681.12942>.
40. Pandi-Perumal, S.R.; BaHammam, A.S.; Ojike, N.I.; Akinseye, O.A.; Kendzerska, T.; Buttoo, K.;



- Dhandapany, P.S.; Brown, G.M.; Cardinali, D.P. Melatonin and human cardiovascular disease. *J. Cardiovasc. Pharmacol. Ther.* **2017**, *22*, 122–132, <https://doi.org/10.1177%2F1074248416660622>.
41. Jevning, R.; Wilson, A.F.; VanderLaan, E.F. Plasma prolactin and growth hormone during meditation. *Psychosom. Med.* **1978**, *40*, 329–333, <https://psycnet.apa.org/doi/10.1097/00006842-197806000-00005>.
  42. Infante, J.R.; Torres-Avisbal, M.; Pinel, P.; Vallejo, J.A.; Peran, F.; Gonzalez, F.; Contreras, P.; Pacheco, C.; Latre, J.M.; Roldan, A. Catecholamine levels in practitioners of the transcendental meditation technique. *Physiol. Behav.* **2001**, *72*, 141–146, [https://doi.org/10.1016/s0031-9384\(00\)00386-3](https://doi.org/10.1016/s0031-9384(00)00386-3).
  43. Glaser, J.L.; Brind, J.L.; Vogelman, J.H.; Eisner, M.J.; Dillbeck, M.C.; Wallace, R.K.; Chopra, D.; Orentreich, N. Elevated serum dehydroepiandrosterone sulfate levels in practitioners of the Transcendental Meditation (TM) and TM-Sidhi programs. *J. Behav. Med.* **1992**, *15*, 327–341, <https://doi.org/10.1007/bf00844726>.
  44. Tooley, G.A.; Armstrong, S.M.; Norman, T.R.; Sali, A. Acute increases in night-time plasma melatonin levels following a period of meditation. *Biol. Psychol.* **2000**, *53*, 69–78, [https://doi.org/10.1016/s0301-0511\(00\)00035-1](https://doi.org/10.1016/s0301-0511(00)00035-1).
  45. Pandi-Perumal, S.R.; Srinivasan, V.; Maestroni, G.J.M.; Cardinali, D.P.; Poeggeler, B.; Hardeland, R. Melatonin: nature's most versatile biological signal? *FEBS J.* **2006**, *273*, 2813–2838, <https://doi.org/10.1111/j.1742-4658.2006.05322.x>.
  46. Dijk, D.-J.; Cajochen, C. Melatonin and the circadian regulation of sleep initiation, consolidation, structure, and the sleep EEG. *J. Biol. Rhythms* **1997**, *12*, 627–635, <https://doi.org/10.1177/074873049701200618>.
  47. Turek, F.W.; Gillette, M.U. Melatonin, sleep, and circadian rhythms: rationale for development of specific melatonin agonists. *Sleep Med.* **2004**, *5*, 523–532, <https://doi.org/10.1016/j.sleep.2004.07.009>.
  48. Martinez, D.; Lenz, M. do C.S. Circadian rhythm sleep disorders. *Indian J. Med. Res.* **2010**, *131*, 141–149, <https://pubmed.ncbi.nlm.nih.gov/20308739/>.
  49. Maestroni, G.J.M. The immunotherapeutic potential of melatonin. *Expert Opin. Investig. Drugs* **2001**, *10*, 467–476, <https://doi.org/10.1517/13543784.10.3.467>.
  50. Nagendra, R.P.; Maruthai, N.; Kuttu, B.M. Meditation and its regulatory role on sleep. *Front. Neurol.* **2012**, *3*, 54, <https://doi.org/10.3389/fneur.2012.00054>.
  51. Domínguez-Rodríguez, A.; Abreu-González, P.; García, M.J.; Sanchez, J.; Marrero, F.; Armas-Trujillo, D. de Decreased nocturnal melatonin levels during acute myocardial infarction. *J. Pineal Res.* **2002**, *33*, 248–252, <https://doi.org/10.1034/j.1600-079X.2002.02938.x>.
  52. Touitou, Y.; Haus, E. *Biologic rhythms in clinical and laboratory medicine*; Springer Science & Business Media, 2012.
  53. Kanki, M.; Young, M.J. Corticosteroids and circadian rhythms in the cardiovascular system. *Curr. Opin. Pharmacol.* **2021**, *57*, 21–27, <https://doi.org/10.1016/j.coph.2020.10.007>.
  54. Man, A.W.C.; Li, H.; Xia, N. Circadian rhythm: potential therapeutic target for atherosclerosis and thrombosis. *Int. J. Mol. Sci.* **2021**, *22*, 676, <https://doi.org/10.3390/ijms22020676>.
  55. Tsutsui, K.; Haraguchi, S.; Inoue, K.; Miyabara, H.; Suzuki, S.; Ogura, Y.; Koyama, T.; Matsunaga, M.; Vaudry, H. Identification, Biosynthesis, and Function of 7 $\alpha$ -Hydroxypregnenolone, a New Key Neurosteroid Controlling Locomotor Activity, in Nonmammalian Vertebrates. *Ann. N. Y. Acad. Sci.* **2009**, *1163*, 308–315.
  56. Nishida, S. Metabolic effects of melatonin on oxidative stress and diabetes mellitus. *Endocrine* **2005**, *27*, 131–135, <https://link.springer.com/article/10.1385/ENDO:27:2:131>.
  57. Zeman, M.; Herichova, I. Melatonin and clock genes expression in the cardiovascular system. *Front. Biosci. - Sch.* **2013**, *5 S*, 743–753, <https://doi.org/10.2741/s404>.
  58. Ruan, W.; Yuan, X.; Eltzschig, H.K. Circadian rhythm as a therapeutic target. *Nat. Rev. Drug Discov.* **2021**, *20*, 287–307, <https://doi.org/10.1038/s41573-020-00109-w>.
  59. Klöting, I.; Berg, S.; Kovács, P.; Voigt, B.; Vogt, L.; Schmidt, S. Diabetes and Hypertension in Rodent Models a. *Ann. N. Y. Acad. Sci.* **1997**, *827*, 64–84.
  60. Chirico, N.; Van Laake, L.W.; Sluijter, J.P.G.; van Mil, A.; Dierickx, P. Cardiac circadian rhythms in time and space: The future is in 4D. *Curr. Opin. Pharmacol.* **2021**, *57*, 49–59, <https://doi.org/10.1016/j.coph.2020.11.006>.
  61. Drăgoi, C.M.; Letiția, A.; Drăgoi, C.M.; Arsene, A.L.; Dinu-pîrvu, C.E.; Dinu-pîrvu, C.E.; Dumitrescu, I.B.; Dumitrescu, B.; Popa, D.E.; Udeanu, I.; et al. World ' s largest Science , Technology & Medicine Open Access book publisher Melatonin : A Melatonin : Silent Regulator of the Glucose Homeostasis Homeostasis. *Carbohydrate* **2017**, 99–113, <https://doi.org/10.5772/66625>.
  62. Reaven, G.M. Role of insulin resistance in human disease. *Diabetes* **1988**, *37*, 1595–1607, <https://doi.org/10.2337/diab.37.12.1595>.
  63. Romon, M.; Nuttens, M.-C.; Fievet, C.; Pot, P.; Bard, J.M.; Furon, D.; Fruchart, J.C. Increased triglyceride levels in shift workers. *Am. J. Med.* **1992**, *93*, 259–262, [https://doi.org/10.1016/0002-9343\(92\)90230-9](https://doi.org/10.1016/0002-9343(92)90230-9).
  64. Griffin, B.A.; Freeman, D.J.; Tait, G.W.; Thomson, J.; Caslake, M.J.; Packard, C.J.; Shepherd, J. Role of plasma triglyceride in the regulation of plasma low density lipoprotein (LDL) subfractions: relative contribution of small, dense LDL to coronary heart disease risk. *Atherosclerosis* **1994**, *106*, 241–253, [https://doi.org/10.1016/0021-9150\(94\)90129-5](https://doi.org/10.1016/0021-9150(94)90129-5).



65. Knutsson, A. Shift work and coronary heart disease. *Scand. J. Soc. Med. Suppl.* **1989**, 1–36, <https://pubmed.ncbi.nlm.nih.gov/2683043/>.
66. Green, C.B.; Takahashi, J.S.; Bass, J. The meter of metabolism. *Cell* **2008**, *134*, 728–742, <https://doi.org/10.1016/j.cell.2008.08.022>.
67. Eckel-Mahan, K.; Sassone-Corsi, P. Metabolism and the circadian clock converge. *Physiol. Rev.* **2013**, *93*, 107–135, <https://doi.org/10.1152/physrev.00016.2012>.
68. Bass, J.; Turek, F.W. Sleepless in America: a pathway to obesity and the metabolic syndrome? *Arch. Intern. Med.* **2005**, *165*, 15–16, <https://doi.org/10.1001/archinte.165.1.15>.
69. Stanley, W.C.; Recchia, F.A.; Lopaschuk, G.D. Myocardial substrate metabolism in the normal and failing heart. *Physiol. Rev.* **2005**, *85*, 1093–1129, <https://doi.org/10.1152/physrev.00006.2004>.
70. Doenst, T.; Nguyen, T.D.; Abel, E.D. Cardiac metabolism in heart failure: implications beyond ATP production. *Circ. Res.* **2013**, *113*, 709–724, <https://doi.org/10.1161/CIRCRESAHA.113.300376>.
71. Heusch, G.; Libby, P.; Gersh, B.; Yellon, D.; Böhm, M.; Lopaschuk, G.; Opie, L. Cardiovascular remodelling in coronary artery disease and heart failure. *Lancet* **2014**, *383*, 1933–1943, [https://doi.org/10.1016/S0140-6736\(14\)60107-0](https://doi.org/10.1016/S0140-6736(14)60107-0).
72. Sniderman, A.D.; Pencina, M.; Thanassoulis, G. Limitations in the conventional assessment of the incremental value of predictors of cardiovascular risk. *Curr. Opin. Lipidol.* **2015**, *26*, 210–214, <https://doi.org/10.1097/mol.0000000000000181>.
73. Lima, F.B.; Machado, U.F.; Bartol, I.; Seraphim, P.M.; Sumida, D.H.; Moraes, S.M.F.; Hell, N.S.; Okamoto, M.M.; Saad, M.J.A.; Carvalho, C.R.O. Pinealectomy causes glucose intolerance and decreases adipose cell responsiveness to insulin in rats. *Am. J. Physiol. Metab.* **1998**, *275*, E934–E941, <https://doi.org/10.1152/ajpendo.1998.275.6.e934>.
74. Bähr, I.; Mühlbauer, E.; Schucht, H.; Peschke, E. Melatonin stimulates glucagon secretion in vitro and in vivo. *J. Pineal Res.* **2011**, *50*, 336–344, <https://doi.org/10.1111/j.1600-079X.2010.00848.x>.
75. Ha, E.; Yim, S.; Chung, J.; Yoon, K.; Kang, I.; Cho, Y.H.; Baik, H.H. Melatonin stimulates glucose transport via insulin receptor substrate-1/phosphatidylinositol 3-kinase pathway in C2C12 murine skeletal muscle cells. *J. Pineal Res.* **2006**, *41*, 67–72, <https://doi.org/10.1111/j.1600-079x.2006.00334.x>.
76. Nishida, S.; Sato, R.; Murai, I.; Nakagawa, S. Effect of pinealectomy on plasma levels of insulin and leptin and on hepatic lipids in type 2 diabetic rats. *J. Pineal Res.* **2003**, *35*, 251–256, <https://doi.org/10.1034/j.1600-079x.2003.00083.x>.
77. Sartori, C.; Dessen, P.; Mathieu, C.; Monney, A.; Bloch, J.; Nicod, P.; Scherrer, U.; Duplain, H. Melatonin improves glucose homeostasis and endothelial vascular function in high-fat diet-fed insulin-resistant mice. *Endocrinology* **2009**, *150*, 5311–5317, <https://doi.org/10.1210/en.2009-0425>.
78. Livrea, M.A.; Tesoriere, L.; D'Arpa, D.; Morreale, M. Reaction of melatonin with lipoperoxyl radicals in phospholipid bilayers. *Free Radic. Biol. Med.* **1997**, *23*, 706–711, [https://doi.org/10.1016/s0891-5849\(97\)00018-x](https://doi.org/10.1016/s0891-5849(97)00018-x).
79. Adams Jr, J.D.; Klaidman, L.K.; Leung, A.C. MPP<sup>+</sup> and MPDP<sup>+</sup> induced oxygen radical formation with mitochondrial enzymes. *Free Radic. Biol. Med.* **1993**, *15*, 181–186, [https://doi.org/10.1016/0891-5849\(93\)90057-2](https://doi.org/10.1016/0891-5849(93)90057-2).
80. Olanow, C.W.; Tatton, W.G. Etiology and pathogenesis of Parkinson's disease. *Annu. Rev. Neurosci.* **1999**, *22*, 123–144, <https://doi.org/10.1146/annurev.neuro.22.1.123>.
81. Vincent, A.M.; Edwards, J.L.; Sadidi, M.; Feldman, E.L. The antioxidant response as a drug target in diabetic neuropathy. *Curr. Drug Targets* **2008**, *9*, 94–100, <https://doi.org/10.2174/138945008783431754>.
82. Traaseth, N.; Elfering, S.; Solien, J.; Haynes, V.; Giulivi, C. Role of calcium signaling in the activation of mitochondrial nitric oxide synthase and citric acid cycle. *Biochim. Biophys. Acta - Bioenerg.* **2004**, *1658*, 64–71, <https://doi.org/10.1016/j.bbabbio.2004.04.015>.
83. Sanchez, O.; Marcos, E.; Perros, F.; Fadel, E.; Tu, L.; Humbert, M.; Darteville, P.; Simonneau, G.; Adnot, S.; Eddahibi, S. Role of endothelium-derived CC chemokine ligand 2 in idiopathic pulmonary arterial hypertension. *Am. J. Respir. Crit. Care Med.* **2007**, *176*, 1041–1047, <https://doi.org/10.1164/rccm.200610-1559oc>.
84. Guignabert, C.; Tu, L.; Le Hir, M.; Ricard, N.; Sattler, C.; Seferian, A.; Huertas, A.; Humbert, M.; Montani, D. Pathogenesis of pulmonary arterial hypertension: lessons from cancer. *Eur. Respir. Rev.* **2013**, *22*, 543–551, <https://doi.org/10.1183/09059180.00007513>.
85. Keeley, F.W.; Rabinovitch, M. Increased Pulmonary Artery Elastolytic Activity in Adult Rats with Monocrotaline-induced Progressive Hypertensive Pulmonary Vascular Disease Compared with Infant Rats with Nonprogressive Disease? *Am Rev Respir Dis* **1992**, *146*, 213–223, <https://doi.org/10.1164/ajrccm/146.1.213>.
86. Ahmed, L.A.; Al Arqam, Z.O.; Zaki, H.F.; Agha, A.M. Role of oxidative stress, inflammation, nitric oxide and transforming growth factor-beta in the protective effect of diosgenin in monocrotaline-induced pulmonary hypertension in rats. *Eur. J. Pharmacol.* **2014**, *740*, 379–387, <https://doi.org/10.1016/j.ejphar.2014.07.026>.
87. Majzunova, M.; Dovinova, I.; Barancik, M.; Chan, J.Y.H. Redox signaling in pathophysiology of hypertension. *J. Biomed. Sci.* **2013**, *20*, 69, <https://link.springer.com/article/10.1186/1423-0127-20-69>.

88. Bowers, R.; Cool, C.; Murphy, R.C.; Tudor, R.M.; Hopken, M.W.; Flores, S.C.; Voelkel, N.F. Oxidative stress in severe pulmonary hypertension. *Am. J. Respir. Crit. Care Med.* **2004**, *169*, 764–769, <https://doi.org/10.1164/rccm.200301-147OC>.
89. Ianas, O.; Olinescu, R.; Badescu, I. Melatonin involvement in oxidative processes. *Rev. Roum. Med. Endocrinol.* **1991**, *29*, 147, <https://pubmed.ncbi.nlm.nih.gov/1821072/>.
90. Reiter, R.J.; Tan, D.X.; Osuna, C.; Gitto, E. Actions of melatonin in the reduction of oxidative stress: A review. *J. Biomed. Sci.* **2000**, *7*, 444–458, <https://doi.org/10.1007/BF02253360>.
91. Cagnacci, A.; Cannoletta, M.; Renzi, A.; Baldassari, F.; Arangino, S.; Volpe, A. Prolonged melatonin administration decreases nocturnal blood pressure in women. *Am. J. Hypertens.* **2005**, *18*, 1614–1618, <https://doi.org/10.1016/j.amjhyper.2005.05.008>.
92. Paulis, L.; Šimko, F. Blood pressure modulation and cardiovascular protection by melatonin: Potential mechanisms behind. *Physiol. Res.* **2007**, *56*, 671–684, [https://www.biomed.cas.cz/physiolres/pdf/56/56\\_671.pdf](https://www.biomed.cas.cz/physiolres/pdf/56/56_671.pdf).
93. Giulivi, C. Characterization and function of mitochondrial nitric-oxide synthase. *Free Radic. Biol. Med.* **2003**, *34*, 397–408, [https://doi.org/10.1016/S0891-5849\(02\)01298-4](https://doi.org/10.1016/S0891-5849(02)01298-4).
94. Okatani, Y.; Wakatsuki, A.; Watanabe, K.; Taniguchi, K.; Fukaya, T. Weak vasoconstrictor activity of melatonin in human umbilical artery: relation to nitric oxide-scavenging action. *Eur. J. Pharmacol.* **2001**, *417*, 125–129, [https://doi.org/10.1016/S0014-2999\(01\)00802-0](https://doi.org/10.1016/S0014-2999(01)00802-0).
95. Absi, E.; Ayala, A.; Machado, A.; Parrado, J. Protective effect of melatonin against the 1-methyl-4-phenylpyridinium-induced inhibition of complex I of the mitochondrial respiratory chain. *J. Pineal Res.* **2000**, *29*, 40–47, <https://doi.org/10.1034/j.1600-079x.2000.290106.x>.
96. Martin, M.; Macias, M.; Escames, G.; Reiter, R.J.; Agapito, M.T.; Ortiz, G.G.; Acuña-Castroviejo, D. Melatonin-induced increased activity of the respiratory chain complexes I and IV can prevent mitochondrial damage induced by ruthenium red in vivo. *J. Pineal Res.* **2000**, *28*, 242–248, <https://doi.org/10.1034/j.1600-079x.2000.280407.x>.
97. Tsutsui, K.; Matsunaga, M.; Miyabara, H.; Ukena, K. Neurosteroid biosynthesis in the quail brain: a review. *J. Exp. Zool. Part A Comp. Exp. Biol.* **2006**, *305*, 733–742, <https://doi.org/10.1002/jez.a.302>.
98. Reiter, R.J.; Tan, D.; Manchester, L.C.; Qi, W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen species. *Cell Biochem. Biophys.* **2001**, *34*, 237–256, <https://doi.org/10.1385/cbb:34:2:237>.
99. Allegra, M.; Reiter, R.J.; Tan, D.; Gentile, C.; Tesoriere, L.; Livrea, M.A. The chemistry of melatonin's interaction with reactive species. *J. Pineal Res.* **2003**, *34*, 1–10, <https://doi.org/10.1034/j.1600-079X.2003.02112.x>.
100. Tan, D.-X.; Manchester, L.C.; Reiter, R.J.; Qi, W.-B.; Karbownik, M.; Calvo, J.R. Significance of melatonin in antioxidative defense system: reactions and products. *Neurosignals* **2000**, *9*, 137–159, <https://doi.org/10.1159/000014635>.
101. Tan, D.-X.; Manchester, L.C.; Reiter, R.J.; Plummer, B.F.; Limson, J.; Weintraub, S.T.; Qi, W. Melatonin directly scavenges hydrogen peroxide: a potentially new metabolic pathway of melatonin biotransformation. *Free Radic. Biol. Med.* **2000**, *29*, 1177–1185, [https://doi.org/10.1016/S0891-5849\(00\)00435-4](https://doi.org/10.1016/S0891-5849(00)00435-4).
102. Tan, D.-X.; Manchester, L.C.; Burkhardt, S.; Sainz, R.M.; Mayo, J.C.; Kohen, R.; Shohami, E.; Huo, Y.-S.; HARDELAND, R.; Reiter, R.J. N 1-acetyl-N 2-formyl-5-methoxykynuramine, a biogenic amine and melatonin metabolite, functions as a potent antioxidant. *FASEB J.* **2001**, *15*, 2294–2296, <https://doi.org/10.1096/fj.01-0309fje>.
103. Reiter, R.J.; Guerrero, J.M.; García, J.J.; Acuña-Castroviejo, D. Reactive oxygen intermediates, molecular damage, and aging: relation to melatonin. *Ann. N. Y. Acad. Sci.* **1998**, *854*, 410–424, <https://doi.org/10.1111/j.1749-6632.1998.tb09920.x>.
104. Reiter, R.J.; Melchiorri, D.; Sewerynek, E.; Peggeler, B.; Barlow-Walden, L.; Chuang, J.; Ortiz, G.G.; Acuña-Castroviejo, D. A review of the evidence supporting melatonin's role as an antioxidant. *J. Pineal Res.* **1995**, *18*, 1–11, <https://doi.org/10.1111/j.1600-079x.1995.tb00133.x>.
105. Lim, H.D.; Kim, Y.S.; Ko, S.H.; Yoon, I.J.; Cho, S.G.; Chun, Y.H.; Choi, B.J.; Kim, E.C. Cytoprotective and anti-inflammatory effects of melatonin in hydrogen peroxide-stimulated CHON-001 human chondrocyte cell line and rabbit model of osteoarthritis via the SIRT1 pathway. *J. Pineal Res.* **2012**, *53*, 225–237, <https://doi.org/10.1111/j.1600-079X.2012.00991.x>.
106. Nian, M.; Lee, P.; Khaper, N.; Liu, P. Inflammatory cytokines and postmyocardial infarction remodeling. *Circ. Res.* **2004**, *94*, 1543–1553, <https://doi.org/10.1161/01.RES.0000130526.20854.f4>.
107. Tassell, B.W. Van; Toldo, S.; Mezzaroma, E.; Abbate, A. Targeting Interleukin-1 in Heart Disease Inflammation: From Tissue Repair to Mechanism of Disease Central Role of Interleukin-1 in the Sterile Inflammatory Response. **2014**, *128*, 804–828, <https://doi.org/10.1161/CIRCULATIONAHA.113.003199>.
108. Pace, T.W.W.; Negi, L.T.; Adame, D.D.; Cole, S.P.; Sivilli, T.I.; Brown, T.D.; Issa, M.J.; Raison, C.L. Effect of compassion meditation on neuroendocrine, innate immune and behavioral responses to psychosocial stress. *Psychoneuroendocrinology* **2009**, *34*, 87–98, <https://doi.org/10.1016/j.psyneuen.2008.08.011>.
109. Rohleder, N. Stimulation of systemic low-grade inflammation by psychosocial stress. *Psychosom. Med.* **2014**, *76*, 181–189, <https://doi.org/10.1097/PSY.0000000000000049>.

110. Chen, J.; Chemaly, E.; Liang, L.; Kho, C.; Lee, A.; Park, J.; Altman, P.; Schecter, A.D.; Hajjar, R.J.; Tarzami, S.T. Effects of CXCR4 gene transfer on cardiac function after ischemia-reperfusion injury. *Am. J. Pathol.* **2010**, *176*, 1705–1715, <https://doi.org/10.2353/ajpath.2010.090451>.
111. LaRocca, T.J.; Schwarzkopf, M.; Altman, P.; Zhang, S.; Gupta, A.; Gomes, I.; Alvin, Z.; Champion, H.C.; Haddad, G.; Hajjar, R.J.  $\beta$ 2-Adrenergic receptor signaling in the cardiac myocyte is modulated by interactions with CXCR4. *J. Cardiovasc. Pharmacol.* **2010**, *56*, 548, <https://dx.doi.org/10.1097%2FFJC.0b013e3181f713fe>.
112. Frangogiannis, N.G.; Entman, M.L. Targeting the chemokines in myocardial inflammation. *Circulation* **2004**, *110*, 1341–1342, <https://doi.org/10.1161/01.CIR.0000141560.18364.63>.
113. Xie, Q.W.; Kashiwabara, Y.; Nathan, C. Role of transcription factor NF-kappa B/Rel in induction of nitric oxide synthase. *J. Biol. Chem.* **1994**, *269*, 4705–4708, [https://doi.org/10.1016/S0021-9258\(17\)37600-7](https://doi.org/10.1016/S0021-9258(17)37600-7).
114. Shan, W.; Nicol, C.J.; Ito, S.; Bility, M.T.; Kennett, M.J.; Ward, J.M.; Gonzalez, F.J.; Peters, J.M. Peroxisome proliferator-activated receptor- $\beta/\delta$  protects against chemically induced liver toxicity in mice. *Hepatology* **2008**, *47*, 225–235, <https://doi.org/10.1002/hep.21925>.
115. Shioi, T.; Matsumori, A.; Kihara, Y.; Inoko, M.; Ono, K.; Iwanaga, Y.; Yamada, T.; Iwasaki, A.; Matsushima, K.; Sasayama, S. Increased expression of interleukin-1 $\beta$  and monocyte chemotactic and activating factor/monocyte chemoattractant protein-1 in the hypertrophied and failing heart with pressure overload. *Circ. Res.* **1997**, *81*, 664–671, <https://doi.org/10.1161/01.RES.81.5.664>.
116. Seino, Y.; Ikeda, U.; Sekiguchi, H.; Morita, M.; Konishi, K.; Kasahara, T.; Shimada, K. Expression of leukocyte chemotactic cytokines in myocardial tissue. *Cytokine* **1995**, *7*, 301–304, <https://doi.org/10.1006/cyto.1995.0037>.
117. Behr, T.M.; Wang, X.; Aiyar, N.; Coatney, R.W.; Li, X.; Koster, P.; Angermann, C.E.; Ohlstein, E.; Feuerstein, G.Z.; Winaver, J. Monocyte chemoattractant protein-1 is upregulated in rats with volume-overload congestive heart failure. *Circulation* **2000**, *102*, 1315–1322, <https://doi.org/10.1161/01.cir.102.11.1315>.
118. Irwin, M.R.; Cole, S.W. Reciprocal regulation of the neural and innate immune systems. *Nat. Rev. Immunol.* **2011**, *11*, 625–632, <https://doi.org/10.1038/nri3042>.
119. Girouard, H.; Chulak, C.; LeJossec, M.; Lamontagne, D.; de Champlain, J. Chronic antioxidant treatment improves sympathetic functions and  $\beta$ -adrenergic pathway in the spontaneously hypertensive rats. *J. Hypertens.* **2003**, *21*, 179–188, <https://doi.org/10.1097/00004872-200301000-00028>.
120. K.-Laflamme, A.; Wu, L.; Foucart, S.; Champlain, J. de Impaired basal sympathetic tone and  $\alpha$ 1-adrenergic responsiveness in association with the hypotensive effect of melatonin in spontaneously hypertensive rats. *Am. J. Hypertens.* **1998**, *11*, 219–229, [https://doi.org/10.1016/S0895-7061\(97\)00401-9](https://doi.org/10.1016/S0895-7061(97)00401-9).
121. Viswanathan, M.; Hissa, R.; George, J.C. Suppression of sympathetic nervous system by short photoperiod and melatonin in the Syrian hamster. *Life Sci.* **1986**, *38*, 73–79, [https://doi.org/10.1016/0024-3205\(86\)90277-8](https://doi.org/10.1016/0024-3205(86)90277-8).
122. Wang, M.; Yokotani, K.; Nakamura, K.; Murakami, Y.; Okada, S.; Osumi, Y. Melatonin inhibits the central sympatho-adrenomedullary outflow in rats. *Jpn. J. Pharmacol.* **1999**, *81*, 29–33, <https://doi.org/10.1254/jjp.81.29>.
123. Bravo, R.; Matito, S.; Cubero, J.; Paredes, S.D.; Franco, L.; Rivero, M.; Rodríguez, A.B.; Barriga, C. Tryptophan-enriched cereal intake improves nocturnal sleep, melatonin, serotonin, and total antioxidant capacity levels and mood in elderly humans. *Age (Omaha)*. **2013**, *35*, 1277–1285, <https://doi.org/10.1007/s11357-012-9419-5>.
124. Espino, J.; Pariente, J.A.; Rodríguez, A.B. Oxidative stress and immunosenescence: therapeutic effects of melatonin. *Oxid. Med. Cell. Longev.* **2012**, *2012*, <https://doi.org/10.1155/2012/670294>.
125. Ghosh, A.K.; Naaz, S.; Bhattacharjee, B.; Ghosal, N.; Chattopadhyay, A.; Roy, S.; Reiter, R.J.; Bandyopadhyay, D. Mechanism of melatonin protection against copper-ascorbate-induced oxidative damage in vitro through isothermal titration calorimetry. *Life Sci.* **2017**, *180*, 123–136, <https://doi.org/10.1016/j.lfs.2017.05.022>.
126. Reiter, R.J.; Mayo, J.C.; Tan, D.; Sainz, R.M.; Alatorre-Jimenez, M.; Qin, L. Melatonin as an antioxidant: under promises but over delivers. *J. Pineal Res.* **2016**, *61*, 253–278, <https://doi.org/10.1111/jpi.12360>.
127. Tan, D.-X.; Manchester, L.C.; Esteban-Zubero, E.; Zhou, Z.; Reiter, R.J. Melatonin as a potent and inducible endogenous antioxidant: synthesis and metabolism. *Molecules* **2015**, *20*, 18886–18906, <https://doi.org/10.3390/molecules201018886>.
128. Zhou, J.; Zhang, S.; Zhao, X.; Wei, T. Melatonin impairs NADPH oxidase assembly and decreases superoxide anion production in microglia exposed to amyloid- $\beta$ 1–42. *J. Pineal Res.* **2008**, *45*, 157–165, <https://doi.org/10.1111/j.1600-079x.2008.00570.x>.
129. Lim, H.; Kim, Y.; Ko, S.; Yoon, I.; Cho, S.; Chun, Y.; Choi, B.; Kim, E. Cytoprotective and anti-inflammatory effects of melatonin in hydrogen peroxide-stimulated CHON-001 human chondrocyte cell line and rabbit model of osteoarthritis via the SIRT1 pathway. *J. Pineal Res.* **2012**, *53*, 225–237, <https://doi.org/10.1111/j.1600-079x.2012.00991.x>.
130. Kruk, J.; Aboul-Enein, B.H.; Duchnik, E. Exercise-induced oxidative stress and melatonin supplementation: current evidence. *J. Physiol. Sci.* **2021**, *71*, 1–19, <https://link.springer.com/article/10.1186/s12576-021->



- 00812-2.
131. Giordano, M.; Palermo, M.S. Melatonin-induced enhancement of antibody-dependent cellular cytotoxicity. *J. Pineal Res.* **1991**, *10*, 117–121, <https://doi.org/10.1111/j.1600-079X.1991.tb00827.x>.
  132. Guerra, J.; Devesa, J. Melatonin Exerts Anti-Inflammatory, Antioxidant, and Neuromodulatory Effects That Could Potentially Be Useful in the Treatment of Vertigo. *Int. J. Otolaryngol.* **2021**, *2021*, <https://doi.org/10.1155/2021/6641055>.
  133. Mauriz, J.L.; Collado, P.S.; Veneroso, C.; Reiter, R.J.; González-Gallego, J. A review of the molecular aspects of melatonin's anti-inflammatory actions: recent insights and new perspectives. *J. Pineal Res.* **2013**, *54*, 1–14, <https://doi.org/10.1111/j.1600-079X.2012.01014.x>.
  134. Dong, Y.; Fan, C.; Hu, W.; Jiang, S.; Ma, Z.; Yan, X.; Deng, C.; Di, S.; Xin, Z.; Wu, G. Melatonin attenuated early brain injury induced by subarachnoid hemorrhage via regulating NLRP3 inflammasome and apoptosis signaling. *J. Pineal Res.* **2016**, *60*, 253–262, <https://doi.org/10.1111/jpi.12300>.
  135. Galano, A.; Tan, D.X.; Reiter, R.J. Melatonin as a natural ally against oxidative stress: A physicochemical examination. *J. Pineal Res.* **2011**, *51*, 1–16, <https://doi.org/10.1111/j.1600-079X.2011.00916.x>.
  136. Agil, A.; Reiter, R.J.; Jiménez-Aranda, A.; Ibán-Arias, R.; Navarro-Alarcón, M.; Marchal, J.A.; Adem, A.; Fernández-Vázquez, G. Melatonin ameliorates low-grade inflammation and oxidative stress in young Zucker diabetic fatty rats. *J. Pineal Res.* **2013**, *54*, 381–388, <https://doi.org/10.1111/jpi.12012>.
  137. Bonnefont-Rousselot, D.; Collin, F.; Jore, D.; Gardès-Albert, M. Reaction mechanism of melatonin oxidation by reactive oxygen species in vitro. *J. Pineal Res.* **2011**, *50*, 328–335, <https://doi.org/10.1111/j.1600-079X.2010.00847.x>.
  138. Tu, Y.; Song, E.; Wang, Z.; Ji, N.; Zhu, L.; Wang, K.; Sun, H.; Zhang, Y.; Zhu, Q.; Liu, X. Melatonin attenuates oxidative stress and inflammation of Müller cells in diabetic retinopathy via activating the Sirt1 pathway. *Biomed. Pharmacother.* **2021**, *137*, 111274, <https://doi.org/10.1016/j.biopha.2021.111274>.
  139. Wang, B.; Zuo, X.; Peng, L.; Wang, X.; Zeng, H.; Zhong, J.; Li, S.; Xiao, Y.; Wang, L.; Ouyang, H. Melatonin ameliorates oxidative stress-mediated injuries through induction of HO-1 and restores autophagic flux in dry eye. *Exp. Eye Res.* **2021**, *205*, 108491, <https://doi.org/10.1016/j.exer.2021.108491>.
  140. Higo, T.; Naito, A.T.; Sumida, T.; Shibamoto, M.; Okada, K.; Nomura, S.; Nakagawa, A.; Yamaguchi, T.; Sakai, T.; Hashimoto, A.; et al. DNA single-strand break-induced DNA damage response causes heart failure. *Nat. Commun.* **2017**, *8*, 1–13, <https://doi.org/10.1038/ncomms15104>.
  141. Shih, Y.-H.; Chiu, K.-C.; Wang, T.-H.; Lan, W.-C.; Tsai, B.-H.; Wu, L.-J.; Hsia, S.-M.; Shieh, T.-M. Effects of melatonin to arecoline-induced reactive oxygen species production and DNA damage in oral squamous cell carcinoma. *J. Formos. Med. Assoc.* **2021**, *120*, 668–678, <https://doi.org/10.1016/j.jfma.2020.07.037>.
  142. Rao, K.S. Genomic damage and its repair in young and aging brain. *Mol. Neurobiol.* **1993**, *7*, 23–48, <https://doi.org/10.1007/bf02780607>.
  143. Ames, B.N.; Shigenaga, M.K.; Hagen, T.M. Oxidants, antioxidants, and the degenerative diseases of aging. *Proc. Natl. Acad. Sci.* **1993**, *90*, 7915–7922, <https://doi.org/10.1073/pnas.90.17.7915>.
  144. Sano, M.; Minamino, T.; Toko, H.; Miyauchi, H.; Orimo, M.; Qin, Y.; Akazawa, H.; Tateno, K.; Kayama, Y.; Harada, M. p53-induced inhibition of Hif-1 causes cardiac dysfunction during pressure overload. *Nature* **2007**, *446*, 444–448, <https://www.nature.com/articles/nature05602>.
  145. Aydemir, S.; Akgun, S.G.; Beceren, A.; Yuksel, M.; Kumas, M.; Erdogan, N.; Sardas, S.; Omurtag, G.Z. Melatonin ameliorates oxidative DNA damage and protects against formaldehyde-induced oxidative stress in rats. *Int. J. Clin. Exp. Med.* **2017**, *10*, 6250–6261, <https://acikerisim.medipol.edu.tr/xmlui/handle/20.500.12511/2751>.
  146. Singh, N.P.; Danner, D.B.; Tice, R.R.; Brant, L.; Schneider, E.L. DNA damage and repair with age in individual human lymphocytes. *Mutat. Res.* **1990**, *237*, 123–130, [https://doi.org/10.1016/0921-8734\(90\)90018-m](https://doi.org/10.1016/0921-8734(90)90018-m).
  147. Reiter, R.J.; Herman, T.S.; Meltz, M.L. Melatonin reduces gamma radiation-induced primary DNA damage in human blood lymphocytes. *Mutat. Res. Mol. Mech. Mutagen.* **1998**, *397*, 203–208, [https://doi.org/10.1016/S0027-5107\(97\)00211-X](https://doi.org/10.1016/S0027-5107(97)00211-X).
  148. Vijayalaxmi; Reiter, R.J.; Sewerynek, E.; Poeggeler, B.; Leal, B.Z.; Meltz, M.L. Marked reduction of radiation-induced micronuclei in human blood lymphocytes pretreated with melatonin. *Radiat. Res.* **1995**, *143*, 102–106, <https://doi.org/10.2307/3578932>.
  149. Osseni, R.A.; Rat, P.; Bogdan, A.; Warnet, J.-M.; Touitou, Y. Evidence of prooxidant and antioxidant action of melatonin on human liver cell line HepG2. *Life Sci.* **2000**, *68*, 387–399, [https://doi.org/10.1016/S0024-3205\(00\)00955-3](https://doi.org/10.1016/S0024-3205(00)00955-3).
  150. Clapp-Lilly, K.L.; Smith, M.A.; Perry, G.; Harris, P.L.; Zhu, X.; Duffy, L.K. Melatonin acts as antioxidant and pro-oxidant in an organotypic slice culture model of Alzheimer's disease. *Neuroreport* **2001**, *12*, 1277–1280, <https://doi.org/10.1097/00001756-200105080-00044>.
  151. Hatch, E.M.; Kulukian, A.; Holland, A.J.; Cleveland, D.W.; Stearns, T. Cep152 interacts with Plk4 and is required for centriole duplication. *J. Cell Biol.* **2010**, *191*, 721–729, <https://doi.org/10.1083/jcb.201006049>.
  152. Wang, S.; Wei, M.; Zhu, W. Melatonin increases doxorubicin-induced apoptosis via oxidative DNA damage in oral squamous cell carcinoma. *DNA Repair (Amst.)* **2021**, *103154*, <https://doi.org/10.1016/j.dnarep.2021.103154>.

153. Liu, R.; Fu, A.; Hoffman, A.E.; Zheng, T.; Zhu, Y. Melatonin enhances DNA repair capacity possibly by affecting genes involved in DNA damage responsive pathways. *BMC Cell Biol.* **2013**, *14*, 1, <https://doi.org/10.1186/1471-2121-14-1>.
154. Zhao, F.; Whiting, S.; Lambourne, S.; Aitken, R.J.; Sun, Y. Melatonin alleviates heat stress-induced oxidative stress and apoptosis in human spermatozoa. *Free Radic. Biol. Med.* **2021**, *164*, 410–416, <https://doi.org/10.1016/j.freeradbiomed.2021.01.014>.