

Pharmacological Aspects of a Bioactive Compound Arbutin: A Comprehensive Review

Medha Bhalla¹ , Roopal Mittal² , Manish Kumar³ , Ajay Singh Kushwah^{1,*} 

¹ Department of Pharmacology, Amar Shaheed Baba Ajit Singh Jujhar Singh Memorial College of Pharmacy, Bela, 140111, Ropar, Punjab, India; medhabhalla1997@gmail.com (M.B.);

² Ph.D. Research Scholar, IK Gujral Punjab Technical University, Jalandhar (Punjab) 144601, India; and Assistant Professor, R.K.S.D. College of Pharmacy, Kaithal (Haryana) 136027, India; roopmittal17@gmail.com (R.M.);

³ Chitkara College of Pharmacy, Chitkara University, Punjab, India; mkpharmacology@gmail.com (M.K.);

* Correspondence: kushwah_ph05@yahoo.co.in (A.S.K.);

Scopus Author ID: 56728206700

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Abstract: Over the past few decades, researchers emphasized herbal medicines as an alternative therapeutic approach due to the high success rate of developing a new drug from a natural template. This review aims to discuss the recent pharmacological activities of Arbutin, natural hydroquinone found abundantly in nature. Google Scholar, Pubmed, ScienceDirect, and Scopus databases were explored to retrieve the last ten-year articles related to pharmacological activities of Arbutin. This drug is extensively used to prevent hyperpigmentation disorders like Melasma, Ephelide, Freckles, and Solar Lentigines. It is hydrolyzed in the gut into hydroquinone and glucose. Although its antioxidant and anti-inflammatory properties are responsible for various pharmacological effects, Arbutin is also used traditionally for hypoglycemic, wound healing, and renal activities. Arbutin modulates several pathways such as insulin-like growth factor-1 receptor (IGF-1R), 5' adenosine monophosphate-activated protein kinase (AMPK), and major excitatory pathways and mechanisms (e.g., Glucose-transporter-4 expression). It can attenuate matrix metalloproteinases, α -amylase, α -glucosidase, tyrosinase, and transcription factors. Taken together, these findings make Arbutin a fascinating natural molecule, which might be explored for health care, therapeutic properties, and drug template. Preclinical data indicated the therapeutic significance of Arbutin. However, clinical efficacy and safety need rigorous investigations.

Keywords: Arbutin; herbal; anticancer; pro-inflammatory; antioxidant; hydroquinoline.

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1. Introduction

Pharmaceutical businesses are the backbone of economic development all over the world. It is projected that worldwide prescription and medication expenditure in the United States in 2006 was \$643 billion, accounting for half of the global pharmaceutical market [1]. Mother Nature has gifted a plethora of therapeutic alternatives, contributing ~ 50% of all medications in the modern world. In contrast to natural medicines, conventional drugs used in the existing practice have certain limitations, higher chances of adverse events, less success rate, and large capital investment in the drug development process. Furthermore, many conventional medicines have low patient compliance, unpleasant responses, and now show lesser efficacy, particularly against the infectious course of the disease, primarily due to a mismatch between the evolution of pathogenesis and targets of a conventional drug.

On the other hand, plant-derived products interact more positively with the human body target several pathways (or molecular mechanisms), as a result, exert numerous beneficial effects in the promotion of health care. In recent decades, there has been a steady but substantial growth in the consumption of dietary supplements such as vitamins, herbal nutrients, and therapeutic foods. These dietary supplementary and herbal products are free of major side effects and have a chemical composition that can simultaneously target different etiopathogenic factors. Between 1990 and 1997, over 15 million individuals in the United States reported using herbal supplements in addition to prescription medicines[2]. Likewise, there was a more than 50% rise in the usage of dietary supplements from 1997 to 2002[3].

Arbutin is a natural substance that can be found in the following families, i.e., *Ericaceae* (bearberry, strawberry tree, huckleberry, and heather), *Saxifragaceae*, *Asteraceae*, *Rosaceae*, *Lamiaceae*, and *Apiaceae*. It is abundantly present in meals and can also be consumed in herbal dietary supplements, over-the-counter medications, or through dermal contact from skin-lightening treatments. It is a hydroquinone glycoside chemically. There are two Arbutin isoforms as depicted in Figure 1, one is α -Arbutin (4-hydroxyphenyl-glucopyranoside)[4], and the other is β -Arbutin (4-hydroxyphenyl-glucopyranoside).

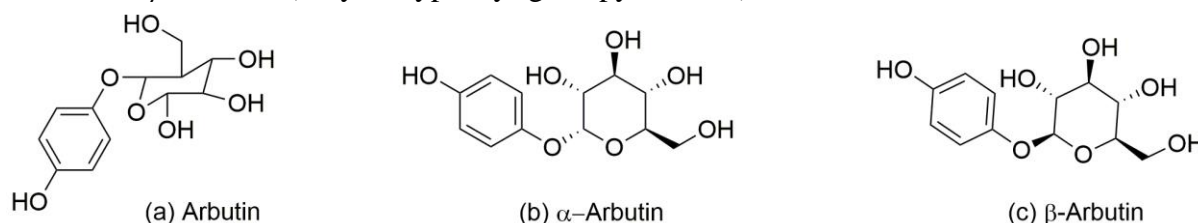


Figure 1. Structure of Arbutin and its isoforms as (a) structure of Arbutin; (b) structure of α -Arbutin; (c) structure of β -Arbutin.

According to the research database Arbutin and its derivatives have been the topic of at least 120 scientific articles reported in the last ten years. The National Institute of Environmental Health Sciences (NIEHS) nominated Arbutin for *in-vitro* and *in-vivo* metabolism and disposal and genotoxicity investigations[5]. Plant extracts are now being replaced by Arbutin and its derivatives that have been chemically and biotechnologically produced.

1.1. Synthesis of Arbutin.

Arbutin can be prepared in the laboratory by Helferich glycosylation reaction of penta-*O*-acetyl- β -D-glucopyranoside and 4-hydroxyphenylacetate, using the catalyst ($\text{BF}_3 \cdot \text{Et}_2\text{O}$) has given expected peracetylated-Arbutin. After that, saponification with potassium carbonate in aqueous methanol or by aminolysis with methanolic ammonia is performed. Figure 2 shows the reactants and catalysts involved in the formation of Arbutin on a laboratory scale. Arbutin has the chemical formula of $\text{C}_{12}\text{H}_{16}\text{O}_7$, and its melting and boiling points are 199.5°C and 561.6°C , respectively. Being a polar compound in nature, Arbutin is freely soluble in water, ethyl ether, alcohol but insoluble in benzene, chloroform, and carbon disulfide. Its maximum solubility in water is 5g/100 ml of water. *Candida antactica* is the biocatalyst reported to be used in the transesterification synthesis of Arbutin palmitate [6].

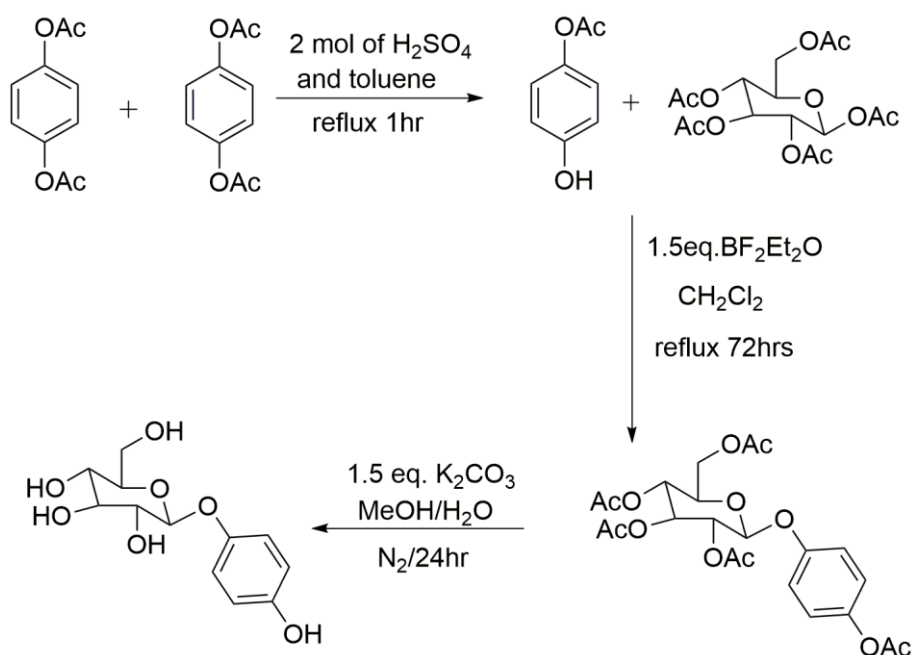


Figure 2. Synthesis of Arbutin in the laboratory by Helferich reaction.

1.2. Pharmacokinetics and pharmacodynamics profile.

Arbutin is absorbed from the gastrointestinal tract (GIT) after getting converted to its bioactive hydroquinone form and inactive glucose by stomach acids. Clinical trials on males and females were performed in which the study groups were asked to take high quantity Arbutin meal (i.e., coffee, tea, bread, Bosc pears, wheat germ). The results showed that there were high levels of plasma hydroquinone after consumption of an Arbutin-rich diet. Urine excretion levels of hydroquinone were also increased after Arbutin-rich meal intake[7]. Preclinical evaluations in female *Wistar* rats showed that Arbutin was excreted unchanged after 16 and 30 hours of administration of an aqueous solution of Arbutin. Oral treatment of Arbutin (500mg/kg) in female rats overloaded with fluid resulted in a four-fold increase in voided urine during the second hour of dosing and a total rise of 61% on the first day. In the urine samples, hydroquinone was not discovered in a free state. Arbutin uptake was enhanced in the jejunum and ileum of rats with a biliary fistula and those given a restricted diet for five days. Arbutin absorption was reduced in both the ileum and the jejunum after administering sodium taurocholate to both groups of mice. As a result, Arbutin had been formulated by various researchers in various formulations such as nanosomes, microspheres, colloidal suspension to increase its bioavailability [8]. Overall, in the present review, information on the recent pharmacological activities of the herbal drug Arbutin research is covered to give a better perspective for potential applications in future research.

2. Pharmacological activities.

2.1. Depigmentation agent.

A copper-containing enzyme, tyrosinase is widely distributed in nature. Tyrosine helps produce melanin, which results in pigmentation of the skin, thus protecting skin from ultraviolet rays-induced damage[9]. However, melanin not only causes hyperpigmentation related disorders like melasma, ephelides, freckles, solar lentigines but also causes browning of fruits, vegetables, fungi, and crustaceans, which anguishes their quality as well as

organoleptic properties, thus reducing their commercial value and the solution to prevent melanin lies in tyrosinase inhibitors[10-12]. Chemically, Melanins are heterogeneous polyphenol polymers of different colors ranging from yellow to brown, consisting of moieties linked by strong carbon-carbon bonds[13]. Compounds that bind to the active site of tyrosinase and inhibit melanin synthesis have been evolved as agents to lighten skin and ameliorate hyperpigmented lesions. Arbutin mainly works by inhibiting tyrosinase, but it does not affect the expression of m-RNA, and for this reason, tyrosine does not get converted to l-DOPA and ultimately melanin[14-16].

In vitro studies have been conducted which have demonstrated that Arbutin oxidizes in the presence of tyrosinase, making the action of Arbutin sustainable in the present. [17-19]. Additionally, Arbutin inhibited UV-induced nuclear factor-kappa-B activation in human keratinocytes [20]. The arbusome, i.e., Arbutin niosomes, have been developed for the hiperpigmenting skin; over and above, the arbusomes did not cause irritancy and cytotoxicity [21].

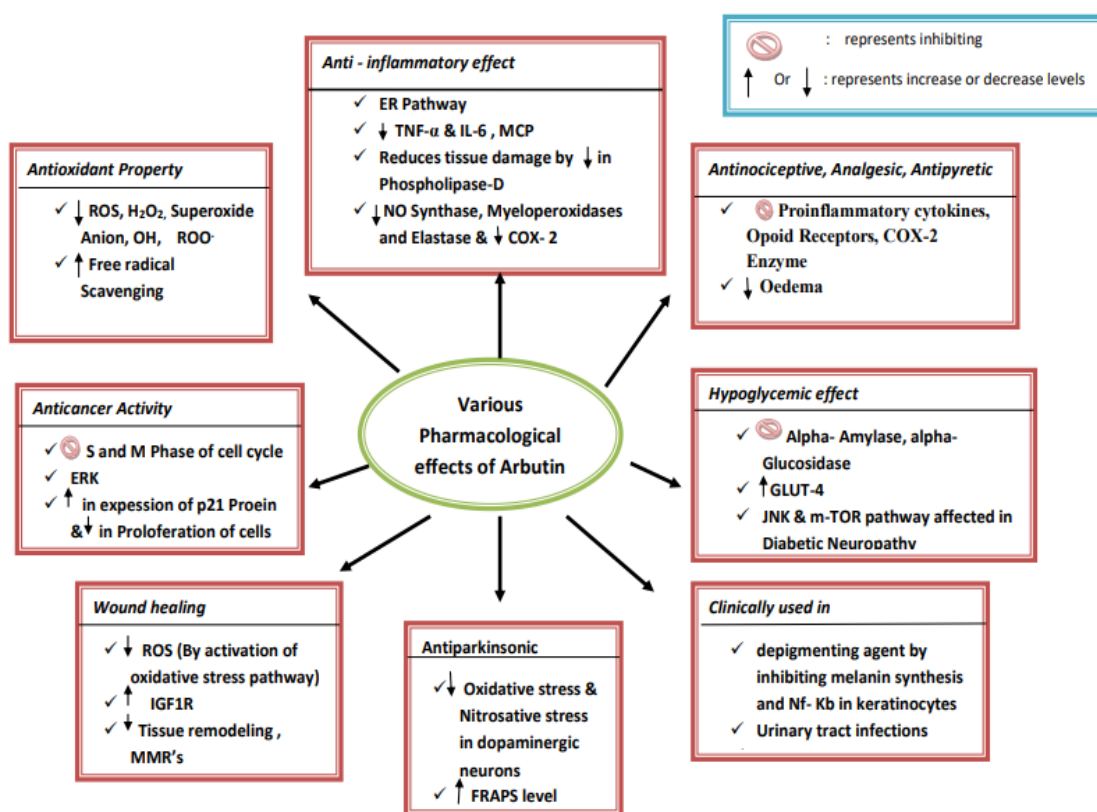


Figure 3. Pharmacological activities of Arbutin.

2.2. Treatment of urinary tract infections (UTIs).

UTIs are bacterial infections that mostly affect otherwise healthy women and are not related to underlying anatomic or functional abnormalities or persistent co-morbidities. Lower urinary tract infections (UTIs) are characterized by frequent urination (polylakiuria), discomfort at menstruation with just a little volume of urine discharged (dysuria), urgency, and blood in the urine (hematuria). Patients suffering from upper UTIs may have fever and flank discomfort [22]. Arbutin is absorbed immediately by glucose transporters and subsequently hydrolyzed in the liver to produce aglycone (also known as hydroquinone) and glucose [23]. Some hydroquinone is absorbed through the gastrointestinal tract. In the liver, hydroquinone is conjugated to glucuronic acid and sulfides, making it water-soluble. The hydroquinone

glucuronide and sulfide are subsequently eliminated in the urine with a pH of 8 and above [24]. The complex spontaneously dissociates, producing free hydroquinone, which has antibacterial action [25]. Avorn *et al.* conducted large, double-blind research in which 153 women with bacteriuria and pyuria were given 300ml of saccharin-sweetened cranberry juice or a daily placebo drink [26]. The individuals who were given cranberry juice had much more sterile urination than placebos. Cranberry did not demonstrate a protective effect against new bacterial colonization in this research, just a conversion from colonized to non-colonized condition. However, Cranberry reduced or eliminated the germs to a clinically significant degree which accounted for an essential step toward preventing frank cystitis. Bailey *et al.* conducted another, smaller research including 12 women who had at least six UTIs in the previous year found that none of the women receiving 400mg of *cranberry* extract daily for 12 weeks developed a UTI. Among them, eight of the women continued to take the extract after the trial concluded that no infection had occurred two years later [27]. Jepson *et al.* also conducted a comprehensive study found that preventive cranberry usage in those with recurrent UTIs dramatically decreased the frequency of such infections after a year [28]. Afshar *et al.* conducted a double-blind, randomized, and controlled comparative effectiveness trial was conducted. Women between the ages of 18 and 75 were suspected of having UTIs. At least two symptoms, dysuria, urgency, frequency, or lower abdominal pain, were assessed for general practice and enrolled in the clinical trial. Participants were given a daily dosage of Arbutin 105mg for successive five days (intervention) or the usual medication, fosfomycin 3g, once a day (control). Antibiotic treatment was administered to women who had worsening or persistent symptoms. The number of all antibiotic courses from day 0–28, independent of medical reason, and the symptom load, defined as a weighted sum of the daily total symptom ratings from day 0–7, are two co-primary outcomes. The results were considered good if the superiority of first treatment with Arbutin was established compared to the co-primary outcome number of antibiotic courses and non-inferiority of initial therapy with Arbutin compared to the co-primary outcome symptom burden [29].

2.3. Antioxidant effect.

Cardiovascular diseases (CVDs) are one of the leading causes of mortality globally, with more people dying from CVDs than any other cause each year. The primary cause of CVD, including cardiomyopathies, congestive heart failure, and atherosclerosis, is oxidative stress caused by free radicals [30]. One of the primary free radicals responsible for cardiotoxicity has been identified as reactive oxygen species (ROS). Hydrogen peroxide (H_2O_2), superoxide anion (O_2^-), reactive hydroxyl radicals (OH), and peroxy radicals are examples of frequent ROS [31]. Sivasangari *et al.*, in their study, addressed that Arbutin has been shown in trials to have a prophylactic cardiopreventive effect. The creation of reactive oxygen species was considerably enhanced when isoproterenol was delivered to H9c2 cells, but the formation of isoproterenol-induced reactive oxygen stress was dramatically decreased when Arbutin was already provided to the H9c2 cells. Arbutin pretreatment significantly enhanced isoproterenol-induced alterations in mitochondrial membrane potential in H9c2 cells and avoided apoptosis in the cell line [32].

The Bang *et al.* investigated if Arbutin may be hydrolyzed by skin microflora to generate its active form, hydroquinone. *Staphylococcus epidermidis* and *Staphylococcus aureus* were employed to assess the hydrolytic capability of Arbutin. It was hydrolyzed by both strains, with activity ranging from 0.16 to 4.51nmol/min/mg. Hydrolyzed hydroquinone

outperformed Arbutin in terms of 1,1-diphenyl-2-picrylhydrazyl radical scavenging activity and tyrosinase inhibition. These data showed that normal skin microflora might enhance Arbutin's skin lightening impact owing to hydroquinone's antioxidant activity [17, 33].

Oxidative stress is associated with aging and various age-related bone pathological disorders. It alters bone remodeling by triggering apoptosis in osteoblasts and osteoclasts. As a result, various bone disorders, such as osteoporosis, rheumatoid arthritis, arthritis, joint pain, may develop [34]. Increased levels of oxidative stress occur with the implantation of metallic prostheses, which is the most often used therapy to ensure appropriate load-bearing characteristics. Thus, it is beneficial to protect bone cells from oxidative stress, which aggravates inflammation and might result in implant rejection to avoid osteointegration [35]. Cells intrinsically fought the harmful effects of reactive oxygen species (ROS) by a variety of methods, including the up-regulation of free radical scavenger enzymes via the activation of the forkhead box O (FoxO) family of ubiquitous transcription factors [36]. One of the most creative research techniques focused on exploiting plant-derived antioxidant compounds to alleviate the harmful effects of free radicals in order to restore the oxidative equilibrium and cellular balance [37]. In their work, Maria *et al.* addressed that nothing was known about Arbutin antioxidant effects on human osteoblasts; hence, the study investigated the *in-vitro* protective impact of Arbutin on osteoblast-like cells (Saos-2) and periosteum-derived progenitor cells (PDPCs). Surprisingly, Arbutin protected bone cells against oxidative stress, preserving cell viability and differentiation [38].

Further, an antioxidant coating containing Arbutin was electro-synthesized on titanium based on these promising results. A polyacrylate-based method was created for the first time to release the effective concentration of Arbutin *in-situ*. The novel coating was evaluated from a physicochemical and morphological standpoint in order to produce an optimal system that was tested *in-vitro* with cells. Morpho-functional analyses revealed that the Arbutin-loaded covering was highly viable and compatible, promoting the expression of periosteum-derived progenitor cells and differentiation markers even under oxidative stress. These findings corroborated that the coatings in *in-vitro* antioxidant activity demonstrated a strong DPPH radical scavenging action. Taken together, their findings provide exciting possibilities for the future development of natural bioactive coatings for orthopedics titanium implants.

2.4. Anti-inflammatory effect.

Zhang *et al.* addressed in their study that Arbutin possessed the ability to reduce inflammation by lowering levels of tumor necrosis factor- α (TNF- α), interleukin-6, and the estrogen receptor pathway [39]. The research showed that the Arbutin could reduce oxidative stress-induced inflammation and autophagy in rats with lipopolysaccharide-induced cardiac damage via the inhibiting estrogen receptor route. Neutrophils are the most important phagocytic cells in the human body, serving as the first line of defense during inflammation by Pecivova *et al.* [40]. Matsuda *et al.* investigated that the combined impact of Arbutin and prednisolone or dexamethasone showed a synergistic effect on Type-IV allergic reaction-induced immune inflammation in their study. When combined with prednisolone or dexamethasone, the Arbutin significantly reduced swelling compared to prednisolone alone [41]. Lee *et al.* addressed that Arbutin reduced nitric oxide generation and the expression of cyclooxygenase (COX-2) and nitrous oxide synthetase in murine microglial cell lines stimulated by lipopolysaccharides. It was also said that Arbutin's mode of action included the suppression of pro-inflammatory cytokines such as interleukin-1b, Tumor necrotic factor- α ,

and MCP-1, as well as reducing cell adhesion ability and the production of adhesion molecules [42].

2.5. Cardioprotective activity.

Myocardial infarction is an early condition of myocardium necrosis that results from an imbalance in coronary blood supply to any heart region, resulting in cardiac tissue death (myocardial necrosis). The main consequences of myocardial infarction were the peroxidation of membrane lipids, loss of plasma membrane integrity, and hyperlipidaemia [43]. Sivasangari *et al.* demonstrated the cardioprotective effect of Arbutin (25mg/kg, 50mg/kg) on isoproterenol-induced myocardial infarction in their investigation [44]. Isoproterenol-induces myocardial injury by hypoxia, calcium overload, and disruption of the electron transport chain pathway, as well as the coronary hypotension, excessive free radical generation, and energy loss owing to its oxidative metabolism [32]. Furthermore, lipids accumulate in cardiac tissue, increasing the amounts of lipoproteins, low-density lipids (LDL), apoptotic pathway beginning by the lysosome, which was initially referred to as suicide bags, store certain hydrolytic enzymes, and became activated owing to the production of free radicals [45]. The inhibition of membrane peroxidation might result in the prevention of hydrolytic enzyme leakage [46]. Ravichandran *et al.* also concluded in their study that the Arbutin-treated rats had decreased activity of hydrolase enzymes such as β -glucosidase, β -glucuronidase, α - and β -galactosidase, cathepsin-B, as well as cathepsin-D, which caused a reduction in DNA damage, normal levels of collagen, prevented lipid changes, and maintained normal membrane fluidity and functions of the myocardial cells. The cARB is a selective inhibitor of the catalytic activity of cAMP-dependent protein kinase, which might lower HSL (triacylglycerol lipase) activity and therefore the hydrolysis of stored triglycerides. Arbutin also reduced the levels of all atherogenic cholesterol (TC, LDLC, and VLDL-C) while increasing the levels of anti-atherogenic cholesterol (HDL-C). Arbutin preserved the activity of mitochondrial respiratory chain enzymes such as Alpha-ketoglutarate dehydrogenase (α -KGDH), which were thought to be reduced in cardiac disease. Arbutin, as an antioxidant, inhibited the lipid peroxidation of lysosomal membranes, resulting in a reduction of lysosomal enzymes [46].

2.6. Wound healing activity.

Matrix Metalloproteinase proteins (MMPs) are calcium-dependent zinc-containing enzymes found in acute and chronic wounds. MMPs are usually present at low levels in normal individuals. MMPs are rapidly expressed and activated during tissue remodeling by keratinocytes, fibroblasts, endothelial cells, and inflammatory cells such as monocytes, lymphocytes, and macrophages [47]. In their study, Polouliakh *et al.* noticed that supplementing Arbutin on human dermal fibroblasts minimized reactive oxygen species (ROS) generation by inhibiting the oxidative stress pathway, activated the insulin-like growth factor-1 receptor (IGF-1R) pathway, and promoted wound healing by down-regulating Matrix Metalloproteinase proteins (MMPs) [48].

2.7. Pro-apoptotic activity.

Apoptosis is a complicated self cell programmed death that causes cell shrinkage, chromatin condensation, and inter-nucleosomal DNA breakage [49]. Jiang *et al.* conducted preclinical studies to examine the overall impact of Arbutin and its acetylated form on

melanogenesis and the pro-apoptotic implications on B16 murine melanoma cells. The results revealed that both Arbutin and also its acetylated form caused cell apoptosis, reduced cell viability, G-1 cell cycle phase arrest, and mitochondrial disruption in melanoma cells [50].

2.8. Anticancer activity.

The Kamei *et al.* incubated human colon carcinoma-15 (HCT-15) cells with various concentrations of Arbutin (2.5, 12.5 and 50g/ml [9.2, 45.9, 180M]) for four days in a 5 percent CO₂ incubator, and results showed that there was weakly inhibition of the growth of human colon carcinoma HCT-15 cells by blocking the S and M phases of the cell cycle [51]. Some *in-vitro* experiments were conducted to investigate Arbutin's anticancer efficacy, and the results showed that the substance suppressed extracellular regulated kinase (ERK) and raised p21 protein expression. As a consequence, TCCSUP human bladder cell growth was reduced [52]. The Arbutinis also reported possessing radioprotective properties, thus helping protect cancer development [53].

2.9. Anti-diabetic and anti-hyperlipidaemic activity.

Diabetes is a chronic illness caused by insulin insufficiency, reduced organ response to insulin, or both [54] and has a major influence on patient health, quality of life, mortality rates, and the health service [55]. Carbohydrates are the foundation of the human diet. The enzymes called pancreatic amylase and intestinal-glucosidase break down carbohydrates and aid in absorbing glucose and fructose. Preventing carbohydrate absorption after meal intake is one of the treatment techniques for lowering blood glucose in diabetic patients [56]. Yousefi *et al.* studied *Pyrus bioessieriana* Buhse leaf extract (PbBLE). Its phytochemical component Arbutin was shown to have hypoglycemic and hypolipidemic effects *in-vivo* by *in-vitro* enzymatic carbohydrate digestion with α -amylase and α -glucosidase powder [57]. The reduction in glucose absorption from the small intestine appeared to be a successful strategy for overcoming insulin resistance in diabetic individuals. Further, studies by Madar *et al.* demonstrated that the intestinal glucosidase inhibitor acarbose slowed the time course of sugar and starch digestion in male streptozotocin-induced diabetic rats [58]. Fiedman *et al.* demonstrated that male Zucker rats avoided hyperglycemia and increased glucose transporter type-4 (GLUT-4) expression in the muscle, which mediates the bulk of the glucose transport response in insulin-sensitive tissue [59].

It had been revealed that the phenolic glycosides, which were found in a wide array of plants and comparable manmade materials and are produced by glycosidase, have an affinity for the intestinal glucose transport system and decrease the rate of glucose absorption from the gut. Because each glycoside had a different affinity for intestinal glucose transport, glycosides with an adequate inhibitory impact on glucose absorption would be useful for avoiding postprandial hyperglycemia [60]. Taki *et al.* performed *in-vivo* investigations on postprandial glucose reduction in mice revealed that the biologically active form hydroquinone and Arbutin had good effects and may be used as an adjuvant in diabetes treatment [61].

2.10. Diabetic neuropathy.

Diabetic neuropathy (DN) is a common disorder and is defined as signs and symptoms of peripheral nerve dysfunction in a long-suffering with diabetes mellitus (DM), in whom other causes of peripheral nerve dysfunction have been excluded [62]. Lina *et al.* studied Arbutin

potential for diabetic neuropathy were observed in this study. The functions of Arbutin in high glucose (HG)-induced cell apoptosis and autophagy in HK-2 cells were observed. The results revealed that the Arbutin alleviated HG-induced cell apoptosis autophagy and regulated the related protein levels in HK-2 cells by JNK and mTOR pathways [63, 64].

2.11. Anti-nociceptive, analgesic, and antipyretic activity.

Nonsteroidal anti-inflammatory drugs (NSAIDs) are the usual pain relievers, mostly used to treat inflammation, pyrexia, and mild to moderate pain [65]. However, their therapeutic use is restricted by the associated adverse effects, including stomach ulcers, gastrointestinal bleeding, cardiovascular abnormalities, and kidney dysfunction issues [66]. The hydroquinone or benzene-1,4-diol could be shaped by hydrolyzing Arbutin with the enzyme β -glucosidase [17]. It possesses anti-inflammatory and anti-tyrosinase properties and is not harmful to the stomach [42, 67]. Its anti-inflammatory action was mediated via the inhibition of pro-inflammatory cytokines (TNF- α , IL-1, IL-6) or by the release of vasodilator mediators such as nitric oxide (NO) [68]. In this work, Sakhteman *et al.* assessed the anti-nociceptive effect of hydroquinone derivatives in mice using the tail-flick method. It was dogged that these compounds had an anti-nociceptive action via opioid receptor blocking [69]. This study discovered anti-nociceptive, anti-inflammatory, antipyretic properties. At 10mg/kg, 20mg/kg, and 40mg/kg, synthetic Arbutin especially lower tonic visceral chemically-induced nociception. The findings revealed that the released hydroquinone compounds have analgesic, antipyretic, and anti-inflammatory effects and a lower gastric-ulcerogenic potential. This might be attributed to the COX-2 enzyme being preferentially inhibited. Furthermore, Fawad *et al.* conducted *in-vitro* investigations that indicated that the hydroquinone derivatives possessed a high binding affinity to the cyclooxygenase-2 (COX-2) enzymes [70].

2.12. Nootropic, anti-depressant, anxiolytic activity.

According to the world health organization data (2017), depression is the most prevalent disease in the world, affecting over 300 million people, and bipolar disorder is the second most common. Depression affected around 6.3 percent of the Ukrainian population [71].

Starchenko *et al.* investigated the anti-anxiety, nootropic, anti-depressant, anti-inflammatory, and antibacterial effects of phyto-extract *Calluna vulgaris leaf* Hull.), a flowering plant in the Ericaceae family. The results demonstrated that the extract had anti-inflammatory action since there was no significant swelling. Blood plasma examination revealed that the experimental group had fewer leucocytes, monocytes, neutrophils, and phagocytes. Furthermore, the extract had an antimicrobial action since it inhibited the development of *Pseudomonas aeruginosa*. Furthermore, the anti-depressant was also tested using a forced swim test, with positive results [72].

2.13. Anti-parkinsonian activity.

The gradual loss of dopaminergic neurons in the substantia nigra area of the brain causes motor impairment [73]. Dadgar *et al.* set out to explore the effects of Arbutin administration on behavioral impairment oxidative and nitrosative stress in a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced experimental animals model of Parkinson's disease [74]. It had been proposed earlier that oxidative stress and mitochondrial

dysfunction play key roles in developing Parkinson's disease. MPTP poisoning was most likely caused by reactive nitrogen species such as peroxynitrite, which may harm macromolecules by oxidizing DNA, peroxidizing lipids, and forming protein carbonyls [75]. To generate Parkinson's disease in mice, the doses of MPTP (20mg/kg; i.p.) were administered at 2-hour intervals. Arbutin (50mg/kg, i.p.) was administered one week before MPTP injections and was maintained consecutively for seven days after the lesion. The forepaw stride length, locomotion test, as well as hanging wire test was used to assess behavioral impairments. The Arbutin therapy enhanced motor abilities in an MPTP-induced PD model as compared to the control group by enhancing FRAPS levels in PD mouse brains and protecting dopaminergic neurons from oxidation or injury [76].

3. Conclusions

Great efforts have been made on Arbutin and similar hydroquinoline derivatives, and the findings have revealed that these phytoconstituents have outstanding therapeutic behavior. Many innovative tests are being conducted to investigate the drug's various therapeutic activities. We hope that this comprehensive review article will serve as a resource for identifying new prospects. Its antioxidant property is one of the keenly observed characteristics that inhibit the production of free radicals and scavenges the generated free radicals. Additionally, tests have been conducted on Abrutin to enhance its penetration for maximizing skin lightening effects since research studies pose beneficial effects of this chemical in contrast to other products that are much harmful and sensitive to skin. Many studies have contributed to the tenfold potential of alpha-Arbutin to its natural derivative.

Furthermore, Arbutin's other pharmacological actions can open a key to many discoveries. Its pro-apoptotic action is also a new activity that can be utilized to avoid cell damage and be used prophylactically in various therapies. Therefore, Arbutin would be a better candidate for further research. It could be tested for its cytotoxic properties, using computational tools to understand their mechanism of action, structural conformation, and structural activity relationship. Even though numerous actions of this moiety are known, there are many gaps in this medication's therapeutic potential that need to be investigated.

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Conflict of interest

The authors declare that they had no conflicts of interest.

References

1. Wu, M.; Janssen, S. Dosed without prescription: A framework for preventing pharmaceutical contamination of our nation's drinking water. *Environ. Sci. Technol.* **2011**, *45*, 366-367, <https://doi.org/10.1021/es104032a>.
2. Schulte, J. M.; Nolt, B. J.; Williams, R. L.; Spinks, C. L.; Hellsten, J. J. Violence and threats of violence experienced by public health field-workers. *JAMA* **1998**, *280*, 439-442, <https://doi.org/10.1001/jama.280.5.439>.
3. Tindle, H. A.; Davis, R. B.; Phillips, R. S.; Eisenberg, D. M. Trends in use of complementary and alternative medicine by US adults: 1997-2002. *Altern. Ther. Health Med.* **2005**, *11*, 42, [https://pubmed.ncbi.nlm.nih.gov/15712765/#:~:text=Overall%20CAM%20use%20for%20the,and%20yoga%20\(3.7%25%20vs..](https://pubmed.ncbi.nlm.nih.gov/15712765/#:~:text=Overall%20CAM%20use%20for%20the,and%20yoga%20(3.7%25%20vs..)
4. He, J.; Wei, M.; Zhao, P.; Xu, T.; Liu, C. Chitosan-coated magnetic nanoparticles used as substrate immobilization carrier for α -arbutin biosynthesis process. *Colloids Interface Sci. Commun.* **2021**, *42*, 100391, <https://doi.org/10.1016/j.colcom.2021.100391>.
5. Gallo, F. R.; Multari, G.; Pagliuca, G.; Panusa, A.; Palazzino, G.; Giambenedetti, M.; Petitto, V.; Nicoletti, M. Bearberry identification by a multidisciplinary study on commercial raw materials. *Nat. Prod. Res.* **2013**, *27*, 735-742, <https://doi.org/10.1080/14786419.2012.696253>.
6. Liu, K. J. Synthesis of lipophilic arbutin ester by enzymatic transesterification in high pressure carbon dioxide. *Enzyme Microb. Technol.* **2021**, *148*, 109818, <https://doi.org/10.1016/j.enzmictec.2021.109818>.
7. Hiramoto, K.; Kida, T.; Kikugawa, K. Increased urinary hydrogen peroxide levels caused by coffee drinking. *Biol. Pharm. Bull.* **2002**, *25*, 1467-1471, <https://doi.org/10.1248/bpb.25.1467>.
8. Bostanudin, M. F.; Salam, A.; Mahmood, A.; Arafat, M.; Kaharudin, A. N.; Sahudin, S.; Mat Lazim, A.; Azfaralariff, A. Formulation and In-Vitro Characterisation of Cross-Linked Amphiphilic Guar Gum Nanocarriers for Percutaneous Delivery of Arbutin. *J. Pharm. Sci.* **2021**, *110*, 3907-3918, <https://doi.org/10.1016/j.xphs.2021.08.014>.
9. Sánchez-Ferrer, Á.; Rodríguez-López, J. N.; García-Cánovas, F.; García-Carmona, F. Tyrosinase: a comprehensive review of its mechanism. *Biochimica et Biophysica Acta (BBA)-Protein Structure and Molecular Enzymology* **1995**, *1247*, 1-11, [https://doi.org/10.1016/0167-4838\(94\)00204-T](https://doi.org/10.1016/0167-4838(94)00204-T).
10. Couteau, C.; Coiffard, L. Overview of skin whitening agents: Drugs and cosmetic products. *Cosmetics* **2016**, *3*, 27, <https://doi.org/10.3390/cosmetics3030027>.
11. Ortiz-Ruiz, C. V.; Berna, J.; Rodriguez-Lopez, J. N.; Tomas, V.; Garcia-Canovas, F. Tyrosinase-catalyzed hydroxylation of 4-hexylresorcinol, an antibrowning and depigmenting agent: a kinetic study. *J. Agric. Food Chem.* **2015**, *63*, 7032-7040, <https://doi.org/10.1021/acs.jafc.5b02523>.
12. Pillaiyar, T.; Manickam, M.; Jung, S.-H. Inhibitors of melanogenesis: a patent review (2009–2014). *Expert Opin. Ther. Pat.* **2015**, *25*, 775-788, <https://doi.org/10.1517/13543776.2015.1039985>.
13. Rodriguez-Lopez, J. N.; Tudela, J.; Varon, R.; Garcia-Carmona, F.; Garcia-Canovas, F. Analysis of a kinetic model for melanin biosynthesis pathway. *J. Biol. Chem.* **1992**, *267*, 3801-3810, [https://doi.org/10.1016/S0021-9258\(19\)50597-X](https://doi.org/10.1016/S0021-9258(19)50597-X).
14. Hu, Z.-M.; Zhou, Q.; Lei, T.-C.; Ding, S.-F.; Xu, S.-Z. Effects of hydroquinone and its glucoside derivatives on melanogenesis and antioxidation: Biosafety as skin whitening agents. *J. Dermatol. Sci.* **2009**, *55*, 179-184, <https://doi.org/10.1016/j.jdermsci.2009.06.003>.
15. Lim, Y.-J.; Lee, E. H.; Kang, T. H.; Ha, S. K.; Oh, M. S.; Kim, S. M.; Yoon, T.-J.; Kang, C.; Park, J.-H.; Kim, S. Y. Inhibitory effects of arbutin on melanin biosynthesis of α -melanocyte stimulating hormone-induced hyperpigmentation in cultured brownish guinea pig skin tissues. *Arch. Pharm. Res.* **2009**, *32*, 367-373, <https://doi.org/10.1007/s12272-009-1309-8>.
16. Maeda, K.; Fukuda, M. Arbutin: mechanism of its depigmenting action in human melanocyte culture. *J. Pharmacol. Exp. Ther.* **1996**, *276*, 765-769, <https://jpet.aspetjournals.org/content/276/2/765.abstract>.
17. Bang, S. H.; Han, S. J.; Kim, D. H. Hydrolysis of arbutin to hydroquinone by human skin bacteria and its effect on antioxidant activity. *J. Cosmet. Dermatol.* **2008**, *7*, 189-193, <https://doi.org/10.1111/j.1473-2165.2008.00387.x>.
18. Hori, I.; Nihei, K. I.; Kubo, I. Structural criteria for depigmenting mechanism of arbutin. *Phytother. Res.* **2004**, *18*, 475-479, <https://doi.org/10.1002/ptr.1456>.
19. Rudeekulthamrong, P.; Kaulpiboon, J. Optimization of amylomaltase for the synthesis of α -arbutin derivatives as tyrosinase inhibitors. *Carbohydr. Res.* **2020**, *494*, 108078, <https://doi.org/10.1016/j.carres.2020.108078>.
20. Ahn, K. S.; Moon, K.-Y.; Lee, J.; Kim, Y. S. Downregulation of NF- κ B activation in human keratinocytes by melanogenic inhibitors. *J. Dermatol. Sci.* **2003**, *31*, 193-201, [https://doi.org/10.1016/S0923-1811\(03\)00039-2](https://doi.org/10.1016/S0923-1811(03)00039-2).
21. Radmard, A.; Saeedi, M.; Morteza-Semnani, K.; Hashemi, S. M. H.; Nokhodchi, A. An eco-friendly and green formulation in lipid nanotechnology for delivery of a hydrophilic agent to the skin in the treatment and management of hyperpigmentation complaints: Arbutin niosome (Arbusome). *Colloids Surf. B Biointerfaces* **2021**, *201*, 111616, <https://doi.org/10.1016/j.colsurf.2021.111616>.

22. Grabe, M.; Bjerklund-Johansen, T.; Botto, H.; Çek, M.; Naber, K.; Tenke, P.; Wagenlehner, F. Guidelines on urological infections. *Eur. Urol.* **2015**, *182*, 237-257, https://uroweb.org/wp-content/uploads/18-Urological-Infections_LR.pdf.
23. Alvarado, F. The relationship between Na⁺ and the active transport of arbutin in the small intestine. *Biochim. Biophys. Acta Bioenerg.* **1965**, *109*, 478-494, [https://doi.org/10.1016/0926-6585\(65\)90173-1](https://doi.org/10.1016/0926-6585(65)90173-1).
24. Frohne, D. The urinary disinfectant effect of extract from leaves uva ursi. *Planta Med.* **1970**, *18*, 1-25, <https://doi.org/10.1055/s-0028-1099743>.
25. Bone, K.; Simon Mills, M.; Fnimh, M. *Principles and practice of phytotherapy: modern herbal medicine*. Elsevier Health Sciences: **2012**, <https://www.elsevier.com/books/principles-and-practice-of-phytotherapy/9780443069925>.
26. Avorn, J.; Monane, M.; Gurwitz, J. H.; Glynn, R. J.; Choodnovskiy, I.; Lipsitz, L. A. Reduction of bacteriuria and pyuria after ingestion of cranberry juice. *JAMA* **1994**, *271*, 751-754, <https://doi.org/10.1001/jama.1994.03510340041031>.
27. Bailey, D. T.; Dalton, C.; Daugherty, F. J.; Tempesta, M. S. Can a concentrated cranberry extract prevent recurrent urinary tract infections in women? A pilot study. *Phytomedicine* **2007**, *14*, 237-241, <https://doi.org/10.1016/j.phymed.2007.01.004>.
28. Jepson, R. G.; Williams, G.; Craig, J. C. Cranberries for preventing urinary tract infections. *Cochrane Database Syst. Rev.* **2012**, <https://doi.org/10.1002/14651858.CD001321.pub5>.
29. Afshar, K.; Fleischmann, N.; Schmiemann, G.; Bleidorn, J.; Hummers-Pradier, E.; Friede, T.; Wegscheider, K.; Moore, M.; Gágyor, I. Reducing antibiotic use for uncomplicated urinary tract infection in general practice by treatment with uva-ursi (REGATTA)—a double-blind, randomized, controlled comparative effectiveness trial. *BMC Complement Altern. Med.* **2018**, *18*, 1-8, <https://doi.org/10.1186/s12906-018-2266-x>.
30. Giordano, F. J. Oxygen, oxidative stress, hypoxia, and heart failure. *J. Clin. Investig.* **2005**, *115*, 500-508, <https://doi.org/10.1172/JCI24408>.
31. Pryor, W.; Houk, K.; Foote, C.; Fukuto, J.; Ignarro, L. Free radical biology and medicine: it's a gas, man! *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2006**, <https://doi.org/10.1152/ajpregu.00614.2005>.
32. Sivasangari, S.; Asaikumar, L.; Vennila, L. Arbutin prevents alterations in mitochondrial and lysosomal enzymes in isoproterenol-induced myocardial infarction: An in vivo study. *Hum. Exp. Toxicol.* **2021**, *40*, 100-112, <https://doi.org/10.1177/0960327120945790>.
33. Tan, J.; Yadav, M. K.; Devi, S.; Kumar, M. Neuroprotective effects of arbutin against oxygen and glucose deprivation-induced oxidative stress and neuroinflammation in rat cortical neurons. *Acta Pharm.* **2022**, *72*, 123-134, <https://doi.org/10.2478/acph-2022-0002>.
34. Domazetovic, V.; Marcucci, G.; Iantomasi, T.; Brandi, M. L.; Vincenzini, M. T. Oxidative stress in bone remodeling: role of antioxidants. *Clin. Cases Miner. Bone Metab.* **2017**, *14*, 209, <https://doi.org/10.11138/cmbm/2017.14.1.209>.
35. Żukowski, P.; Maciejczyk, M.; Waszkiel, D. Sources of free radicals and oxidative stress in the oral cavity. *Arch. Oral Biol.* **2018**, *92*, 8-17, <https://doi.org/10.1016/j.archoralbio.2018.04.018>.
36. Storz, P. Forkhead homeobox type O transcription factors in the responses to oxidative stress. *Antioxid. Redox Signal.* **2011**, *14*, 593-605, <https://doi.org/10.1089/ars.2010.3405>.
37. Kasote, D. M.; Katyare, S. S.; Hegde, M. V.; Bae, H. Significance of antioxidant potential of plants and its relevance to therapeutic applications. *Int. J. Biol. Sci.* **2015**, *11*, 982, <https://doi.org/10.7150/ijbs.12096>.
38. Bonifacio, M. A.; Cerqueni, G.; Cometa, S.; Licini, C.; Sabbatini, L.; Mattioli-Belmonte, M.; De Giglio, E. Insights into Arbutin effects on bone cells: Towards the development of antioxidant titanium implants. *Antioxidants* **2020**, *9*, 579, <https://doi.org/10.3390/antiox9070579>.
39. Zhang, B.; Zeng, M.; Li, B.; Wang, Y.; Kan, Y.; Wang, S.; Meng, Y.; Gao, J.; Feng, W.; Zheng, X. Inhibition of oxidative stress and autophagy by arbutin in lipopolysaccharide-induced myocardial injury. *Pharmacogn. Mag.* **2019**, *15*, 507, <http://www.phcog.com/text.asp?2019/15/63/507/258393>.
40. Pečivová, J.; Radomír Nosál, K. S.; Mačičková, T. Arbutin and decrease of potentially toxic substances generated in human blood neutrophils. *Interdiscip. Toxicol.* **2014**, *7*, 195, <https://doi.org/10.2478/intox-2014-0028>.
41. Matsuda, H.; Nakata, H.; Tanaka, T.; Kubo, M. Pharmacological study on *Arctostaphylos uva-ursi* (L.) Spreng. II. Combined effects of arbutin and prednisolone or dexamethazone on immuno-inflammation. *Yakugaku Zasshi: J. Pharm. Soc. Jpn.* **1990**, *110*, 68-76, https://doi.org/10.1248/yakushi1947.110.1_68.
42. Lee, H.-J.; Kim, K.-W. Anti-inflammatory effects of arbutin in lipopolysaccharide-stimulated BV2 microglial cells. *J. Inflamm. Res.* **2012**, *61*, 817-825, <https://doi.org/10.1007/s00011-012-0474-2>.
43. Gürgün, C.; Ildizli, M.; Yavuzgil, O.; Sin, A.; Apaydin, A.; Çınar, C.; Kültürsay, H. The effects of short term statin treatment on left ventricular function and inflammatory markers in patients with chronic heart failure. *Int. J. Cardiol.* **2008**, *123*, 102-107, <https://doi.org/10.1016/j.ijcard.2006.11.152>.
44. Sivasangari, S.; Asaikumar, L.; Vennila, L.; Vijayakumar, N. Preventive Effect of Arbutin on Isoproterenol-Induced Oxidative Stress, Mitochondrial Damage and Apoptosis in H9c2 Cells. *Int. J. Nutr. Pharmacol. Neurol. Dis.* **2019**, *9*, 97, https://doi.org/10.4103/ijnpnd.ijnpnd_24_19.

45. Mohanty, I.; Arya, D. S.; Dinda, A.; Talwar, K. K.; Joshi, S.; Gupta, S. K. Mechanisms of cardioprotective effect of *Withania somnifera* in experimentally induced myocardial infarction. *Basic Clin. Pharmacol. Toxicol.* **2004**, *94*, 184-190, <https://doi.org/10.1111/j.1742-7843.2004.pto940405.x>.
46. Ravichandran, L.; Puvanakrishnan, R.; Joseph, K. T. Influence of isoproterenol-induced myocardial infarction on certain glycohydrolases and cathepsins in rats. *Biochem. Med. Metabol. Biol.* **1991**, *45*, 6-15, [https://doi.org/10.1016/0885-4505\(91\)90003-4](https://doi.org/10.1016/0885-4505(91)90003-4).
47. Yan, C.; Boyd, D. D. Regulation of matrix metalloproteinase gene expression. *J. Cell. Physiol.* **2007**, *211*, 19-26, <https://doi.org/10.1002/jcp.20948>.
48. Polouliakh, N.; Ludwig, V.; Meguro, A.; Kawagoe, T.; Heeb, O.; Mizuki, N. Alpha-Arbutin promotes wound healing by lowering ROS and Upregulating insulin/IGF-1 pathway in human dermal fibroblast. *Front. Physiol.* **2020**, *11*, <https://doi.org/10.3389/fphys.2020.586843>.
49. Reed, J. C. Mechanisms of apoptosis avoidance in cancer. *Curr. Opin. Oncol.* **1999**, *11*, 68, <https://doi.org/10.1097/00001622-199901000-00014>.
50. Jiang, L.; Wang, D.; Zhang, Y.; Li, J.; Wu, Z.; Wang, Z. Investigation of the pro-apoptotic effects of arbutin and its acetylated derivative on murine melanoma cells. *Int. J. Mol. Med.* **2018**, *41*, 1048-1054, <https://doi.org/10.3892/ijmm.2017.3256>.
51. Kamei, H.; Koide, T.; Kojima, T.; Hashimoto, Y.; Hasegawa, M. Inhibition of cell growth in culture by quinones. *Cancer Biother. Radiopharm.* **1998**, *13*, 185-188, <https://doi.org/10.1089/cbr.1998.13.185>.
52. Li, H.; Jeong, Y.-M.; Kim, S. Y.; Kim, M.-K.; Kim, D.-S. Arbutin inhibits TCCSUP human bladder cancer cell proliferation via up-regulation of p21. *Die Pharmazie* **2011**, *66*, 306-309, <https://doi.org/10.1691/ph.2011.0785>.
53. Benković, V.; Marčina, N.; Horvat Knežević, A.; Šikić, D.; Rajevac, V.; Milić, M.; Kopjar, N. Potential radioprotective properties of arbutin against ionising radiation on human leukocytes in vitro. *Mutat. Res. Genet. Toxicol. Environ. Mutagen* **2021**, *872*, 503413, <https://doi.org/10.1016/j.mrgentox.2021.503413>.
54. Bhandari, M. R.; Jong-Anurakkun, N.; Hong, G.; Kawabata, J. α -Glucosidase and α -amylase inhibitory activities of Nepalese medicinal herb Pakhanbhed (*Bergenia ciliata*, Haw.). *Food Chem.* **2008**, *106*, 247-252, <https://doi.org/10.1016/j.foodchem.2007.05.077>.
55. Kwon, Y.-I.; Apostolidis, E.; Shetty, K. In vitro studies of eggplant (*Solanum melongena*) phenolics as inhibitors of key enzymes relevant for type 2 diabetes and hypertension. *Bioresour. Technol.* **2008**, *99*, 2981-2988, <https://doi.org/10.1016/j.biortech.2007.06.035>.
56. Kim, Y.-M.; Wang, M.-H.; Rhee, H.-I. A novel α -glucosidase inhibitor from pine bark. *Carbohydr. Res.* **2004**, *339*, 715-717, <https://doi.org/10.1016/j.carres.2003.11.005>.
57. Yousefi, F.; Mahjoub, S.; Pouramir, M.; Khadir, F. Hypoglycemic activity of *Pyrus brossieriana* Buhse leaf extract and arbutin: Inhibitory effects on alpha amylase and alpha glucosidase. *Casp. J. Intern. Med.* **2013**, *4*, 763, <https://pubmed.ncbi.nlm.nih.gov/24294470/>.
58. Madar, Z. The effect of acarbose and miglitol (BAY-M-1099) on postprandial glucose levels following ingestion of various sources of starch by nondiabetic and streptozotocin-induced diabetic rats. *J. Nutr.* **1989**, *119*, 2023-2029, <https://doi.org/10.1093/jn/119.12.2023>.
59. Friedman, J. E.; de Vente, J. E.; Peterson, R. G.; Dohm, G. L. Altered expression of muscle glucose transporter GLUT-4 in diabetic fatty Zucker rats (ZDF/Drt-fa). *Am. J. Physiol. Endocrinol. Metab.* **1991**, *261*, E782-E788, <https://doi.org/10.1152/ajpendo.1991.261.6.E782>.
60. Welsch, C. A.; Lachance, P. A.; Wasserman, B. P. Dietary phenolic compounds: inhibition of Na⁺-dependent D-glucose uptake in rat intestinal brush border membrane vesicles. *J. Nutr.* **1989**, *119*, 1698-1704, <https://doi.org/10.1093/jn/119.11.1698>.
61. Takii, H.; Matsumoto, K.; Kometani, T.; Okada, S.; Fushiki, T. Lowering Effect of Phenolic Glycosides on the Rise in Postprandial Glucose in Mice. *Biosci. Biotechnol. Biochem.* **1997**, *61*, 1531-1535, <https://doi.org/10.1271/bbb.61.1531>.
62. Sadikot, S. M.; Nigam, A.; Das, S.; Bajaj, S.; Zargar, A. H.; Prasannakumar, K. M.; Sosale, A.; Munichoodappa, C.; Seshiah, V.; Singh, S. K.; Jamal, A.; Sai, K.; Sadasivrao, Y.; Murthy, S. S.; Hazra, D. K.; Jain, S.; Mukherjee, S.; Bandyopadhyay, S.; Sinha, N. K.; Mishra, R.; Dora, M.; Jena, B.; Patra, P.; Goenka. The burden of diabetes and impaired glucose tolerance in India using the WHO 1999 criteria: prevalence of diabetes in India study (PODIS). *Diabetes Res. Clin. Pract.* **2004**, *66*, 301-307, <https://doi.org/10.1016/j.diabres.2004.04.008>.
63. Lv, L.; Zhang, J.; Tian, F.; Li, X.; Li, D.; Yu, X. Arbutin protects HK-2 cells against high glucose-induced apoptosis and autophagy by up-regulating microRNA-27a. *Artif. Cells Nanomed. Biotechnol.* **2019**, *47*, 2940-2947, <https://doi.org/10.1016/j.diabres.2004.04.008>.
64. Kumar, M.; Kumar, A.; Sindhu, R. K.; Kushwah, A. S. Arbutin attenuates monosodium L-glutamate induced neurotoxicity and cognitive dysfunction in rats. *Neurochem. Int.* **2021**, *151*, 105217, <https://doi.org/10.1016/j.neuint.2021.105217>.
65. Chattopadhyay, M.; Velazquez, C. A.; Pruski, A.; Nia, K. V.; Abdellatif, K. R.; Keefer, L. K.; Kashfi, K. Comparison between 3-Nitrooxyphenyl acetylsalicylate (NO-ASA) and O2-(acetylsalicyloxymethyl)-1-(pyrrolidin-1-yl) diazen-1-ium-1, 2-diolate (NONO-ASA) as safe anti-inflammatory, analgesic, antipyretic, antioxidant prodrugs. *J. Pharmacol. Exp. Ther.* **2010**, *335*, 443-450, <https://doi.org/10.1124/jpet.110.171017>.

66. Wallace, J. L.; Keenan, C. M.; Granger, D. N. Gastric ulceration induced by nonsteroidal anti-inflammatory drugs is a neutrophil-dependent process. *Am. J. Physiol. Gastrointest. Liver Physiol.* **1990**, *259*, G462-G467, <https://doi.org/10.1152/ajpgi.1990.259.3.G462>.
67. Taha, M. M. E.; Salga, M. S.; Ali, H. M.; Abdulla, M. A.; Abdelwahab, S. I.; Hadi, A. H. A. Gastroprotective activities of *Turnera diffusa* Willd. ex Schult. revisited: Role of arbutin. *J. Ethnopharmacol.* **2012**, *141*, 273-281, <https://doi.org/10.1016/j.jep.2012.02.030>.
68. Cho, J. Y. Suppressive effect of hydroquinone, a benzene metabolite, on in vitro inflammatory responses mediated by macrophages, monocytes, and lymphocytes. *Mediators Inflamm.* **2008**, *2008*, <https://doi.org/10.1155/2008/298010>.
69. Sakhteman, A.; Sharifzadeh, M.; Moradi, A.; Nadri, H.; Tabrizian, K.; Amanlou, M.; Asadipour, A.; Divsalar, K.; Shafiee, A.; Foroumadi, A. Anti-nociceptive activity of some 1, 4-substituted piperidine derivatives using tail flick method in mice. *Afr. J. Pharmacy Pharmacol.* **2011**, *5*, 352-357, <https://doi.org/10.5897/AJPP10.389>.
70. Fawad, K.; Islam, N. U.; Subhan, F.; Shahid, M.; Ali, G.; Rahman, F.-U.; Mahmood, W.; Ahmad, N. Novel hydroquinone derivatives alleviate algnesia, inflammation and pyrexia in the absence of gastric ulcerogenicity. *Trop. J. Pharm. Res.* **2018**, *17*, 53-63, <https://doi.org/10.4314/tjpr.v17i1.9>.
71. Organization, W. H. *Depression and other common mental disorders: global health estimates*; World Health Organization: 2017, <https://apps.who.int/iris/bitstream/handle/10665/254610/W?sequence=1>.
72. Starchenko, G.; Hrytsyk, A.; Raal, A.; Koshovyi, O. Phytochemical profile and pharmacological activities of water and hydroethanolic dry extracts of *Calluna vulgaris* (L.) Hull. herb. *Plants (Basel)* **2020**, *9*, 751, <https://doi.org/10.3390/plants9060751>.
73. Dawson, G.; Rogers, S.; Munson, J.; Smith, M.; Winter, J.; Greenson, J.; Donaldson, A.; Gerdtts, J. Randomized, controlled trial of an intervention for toddlers with autism: The early start denver model. *Pediatrics* **2009**, *125*, e17-23, <https://doi.org/10.1542/peds.2009-0958>.
74. Dadgar, M.; Pouramir, M.; Dastan, Z.; Ghasemi-Kasman, M.; Ashrafpour, M.; Moghadamnia, A. A.; Khafri, S.; Pourghasem, M. Arbutin attenuates behavioral impairment and oxidative stress in an animal model of Parkinson's disease. *Avicenna J. Phytomed.* **2018**, *8*, 533-542, <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6235658/>.
75. Przedborski, S.; Chen, Q.; Vila, M.; Giasson, B. I.; Djaldatti, R.; Vukosavic, S.; Souza, J. M.; Jackson-Lewis, V.; Lee, V. M.-Y.; Ischiropoulos, H. Oxidative post-translational modifications of α -synuclein in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) mouse model of Parkinson's disease. *J. Neurochem.* **2001**, *76*, 637-640, <https://doi.org/10.1046/j.1471-4159.2001.00174.x>.
76. Przedborski, S.; Tieu, K.; Perier, C.; Vila, M. MPTP as a mitochondrial neurotoxic model of Parkinson's disease. *J. Bioenerg. Biomembr.* **2004**, *36*, 375-379, <https://doi.org/10.1023/B:JOB.0000041771.66775.d5>.