






Efficient Synthesis of Two Chloramphenicol Derivatives as Antibacterial Agents

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Abstract: Several protocols have been used to prepare some chloramphenicol analogs using different reagents, which can be expensive and difficult to handle. The aim of this investigation was to synthesize six chloramphenicol analogs (compounds **2** to **7**) using some chemical strategies to evaluate their biological activity against *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Escherichia coli*, and *Klebsiella pneumoniae* with the minimum inhibitory concentration method. The results indicate that protocols used to synthesize chloramphenicol derivatives do not require special conditions such as different pH and higher temperatures to give a good yielding. Besides, only compounds **4** and **6** decreased the bacterial growth of either Gram-positive or Gram-negative bacteria. However, the biological activity of compound **4** was higher compared with both chloramphenicol and compound **6**.

Keywords: synthesis; chloramphenicol; derivatives; Gram-positive; Gram-negative.

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1. Introduction

Infectious diseases are one of the leading causes of death worldwide [1-3]; it is important to mention that several drugs have been used to treat infectious diseases, such as penicillins [4], cephalosporins [5], aminoglycosides [6, 7], fluoroquinones [8], sulphas [9]. However, some of these drugs can produce bacterial resistance [10-12]; in this way, some studies showed that chloramphenicol could induce bacterial resistance in both Gram-positive and Gram-negative bacteria [13-17]. In the search for a therapeutic alternative to reduce bacterial resistance produced by some bacteria, several antibacterial agents have been developed. For example, a series of derivatives of fluorine-chloramphenicol with antibacterial activity on Gram-negative bacteria were synthesized [18]. Other data showed that some aminoacyl-chloramphenicol analogs could cause decreased bacterial growth of *Escherichia coli* [19]. In addition, a report showed the synthesis of different chloramphenicol-amine derivatives with antibacterial activity on *Escherichia coli* [20]. Besides, some carbonyl-chloramphenicol derivatives were prepared as antibacterial agents against *Staphylococcus*

aureus [21]. Recently, a 3-acylchloramphenicol derivative was synthesized from dichloroacetyl and chloramphenicol as a potential antimicrobial agent [22]. It is important to mention that all these methods require special conditions such as different pH and high temperatures. Analyzing these data, the objective of this research was to synthesize some derivatives of chloramphenicol. In addition, the biological activity exerted by chloramphenicol derivatives on *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Escherichia coli*, and *Klebsiella pneumoniae* was evaluated using the minimum inhibitory concentration method.

2. Material and Methods

2.1. General methods.

All reagents were acquired from Sigma-Aldrich company. NMR spectra were recorded on a Varian VXR300/5 FT apparatus (300 MHz/ CDCl_3) using tetramethylsilane as an internal standard. Electron Ionization mass spectrometry was recorded on a Finnigan PolarisQ ion trap mass spectrometer. The melting point was recorded on an electrothermal-900 model apparatus. The infrared spectrum was recorded on a thermo-scientific iSOFT/IR device. Elemental analysis was determined using a PerkinElmer apparatus (Ser. II CHNS / 02400).

2.2. 4-[(2,2-dichloroacetyl)amino]-3-(4-nitrophenyl)-2,6-dioxabicyclo[5.3.1]undeca-1(10),7(11),8-triene-9-carboxylic acid (2).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), 3,5-dinitrobenzoic acid (135 mg, 0.64 mmol) and potassium carbonate anhydrous (90 mg, 0.65 mmol) in dimethyl sulfoxide (5 mL) was stirring for 72 h at 90 °C. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:hexane (4:1) system; yielding 76 % of product; m.p. 102-104 °C; IR (V_{\max} , cm^{-1}) 1712, 1632, 1540 and 1242: ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 3.32-5.04 (m, 5H), 6.02, (m, 1H), 6.22 (m, 1H), 6.60 (m, 1H), 6.66 (broad, 2H), 6.68 (m, 1H), 7.90-7.96 (m, 4H) ppm. ^{13}C NMR (300 Hz, CDCl_3) δ_{C} : 40.82, 64.42, 78.54, 80.44, 102.10, 111.66, 112.52, 122.94, 125.50, 128.14, 145.30, 146.40, 154.36, 157.56, 161.60, 168.10 ppm. EI-MS m/z: 440.01. Anal. Calcd. for $\text{C}_{18}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}_7$. C, 49.00; H, 3.20; Cl, 16.07; N, 6.35; O, 25.38. Found: C, 49.00; H, 3.18.

2.3. 2,2-dichloro-N-[8-fluoro-3-(4-nitrophenyl)-2,6-dioxabicyclo[5.3.1]undeca-1(10),7(11),8-trien-4-yl]acetamide (3).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), 3,5-dinitrofluoro benzene (120 mg, 0.64 mmol) and potassium carbonate anhydrous (90 mg, 0.65 mmol) in dimethyl sulfoxide (5 mL) was stirring for 72 h at 90 °C. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:hexane:water (3:1:1) system; yielding 51% of product; m.p. 92-94 °C; IR (V_{\max} , cm^{-1}) 1632, 1542 and 1242: ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 3.72-5.60 (m, 5H), 6.22, (m, 1H), 6.50-6.96 (m, 2H), 7.60 (broad, 1H), 7.90-7.96 (m, 4H) ppm. ^{13}C NMR (300 Hz, CDCl_3) δ_{C} : 42.90, 64.42, 80.34, 83.12, 100.60, 110.56, 116.90, 122.82, 125.50, 144.64, 146.57, 149.12, 150.86, 151.30, 161.60 ppm. EI-MS m/z: 414.01. Anal. Calcd. for $\text{C}_{17}\text{H}_{13}\text{Cl}_2\text{N}_2\text{O}_5$. C, 49.18; H, 3.16; Cl, 17.08; F, 4.58; N, 6.75; O, 19.27. Found: C, 49.15; H, 3.14.

2.4. 2,2-dichloro-N-[2-(4-nitrophenyl)-6,9-dioxo-1,5-dioxonan-3-yl]acetamide (4).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), succinic acid (100 mg, 0.84 mmol), N,N'-dicyclohexylcarbodiimide (160 mg, 0.77 mmol) in ethanol (5 mL) was stirring for 72 h at room temperature. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:hexane:benzene (3:1:1) system; yielding 65% of product; m.p. 118-120 °C; IR (V_{\max} , cm^{-1}) 1750, 1630 and 1542: ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 2.22-6.22 (m, 7H), 6.26 (m, 1H), 7.74-7.80 (m, 4H), 7.96 (broad, 1H) ppm. ^{13}C NMR (300 Hz, CDCl_3) δ_{C} : 29.70, 30.44, 49.16, 64.40, 69.50, 79.73, 122.85, 126.18, 144.77, 146.90, 161.80, 170.12, 171.22 ppm. EI-MS m/z: 404.01. Anal. Calcd. for $\text{C}_{15}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}_7$. C, 44.46; H, 3.48; Cl, 17.50; N, 6.91; O, 27.64. Found: C, 44.43; H, 3.45.

2.5. 4-{4-[2-(2,2-Dichloro-acetylamino)-1,3-dihydroxy-propyl]-phenoxy}-benzoic acid (5).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), 4-hydroxibenzoic acid (85 mg, 0.61 mmol), and potassium carbonate anhydrous (90 mg, 0.65 mmol) in dimethyl sulfoxide (5 mL) was stirring for 72 h at 90 °C. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:hexane (4:1) system; yielding 74% of product; m.p. 78-80 °C; IR (V_{\max} , cm^{-1}) 3400, 1712, 1682 and 1240 : ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 4.06-5.02 (m, 4H), 6.20 (m, 1H), 6.35 (broad, 4H), 6.66-8.10 (m, 8H) ppm. ^{13}C NMR (300 Hz, CDCl_3) δ_{C} : 57.70, 63.04, 64.44, 72.32, 114.82, 117.22, 124.20, 128.12, 133.86, 141.20, 154.92, 158.30, 163.22, 168.40 ppm. EI-MS m/z: 413.04. Anal. Calcd. for $\text{C}_{18}\text{H}_{17}\text{Cl}_2\text{NO}_6$. C, 52.19; H, 4.14; Cl, 17.12; N, 3.38; O, 23.17. Found: C, 52.16; H, 4.10.

2.6. (2E)-N-[2-hydroxy-1-(hydroxymethyl)-2-(4-nitrophenyl)ethyl]-2-(phenylhydrazono)-acetamide (6).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), phenylhydrazine hydrochloride (100 mg, 0.69 mmol), copper(II) chloride anhydrous (100 mg, 0.74 mmol) in methanol (5 mL) was stirring for 72 h at room temperature. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:water (4:1) system; yielding 74% of product; m.p. 156-158 °C; IR (V_{\max} , cm^{-1}) 3400. 3320, 1630 and 1540: ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 3.62 (m, 2H), 4.00-5.00 (m, 4H), 6.00 (broad, 5H), 6.44 (m, 1H), 6.80-7.26 (m, 5H), 7.60-7.80 (m, 4H) ppm. ^{13}C NMR (300 Hz, CDCl_3) δ_{C} : 59.90, 63.02, 72.32, 115.60, 121.10, 122.54, 126.94, 128.22, 128.44, 146.68, 149.12, 150.30, 158.60 ppm. EI-MS m/z: 358.12. Anal. Calcd. for $\text{C}_{17}\text{H}_{18}\text{N}_4\text{O}_5$. C, 56.98; H, 5.06; N, 15.63; O, 22.32. Found: C, 56.95; H, 5.04.

2.7. (2E)-2-hydrazinylidene-N-[2-hydroxy-1-(hydroxymethyl)-2-(4-nitrophenyl)ethyl]acetamide (7).

In a round bottom flask (10 mL), chloramphenicol (200 mg, 0.62 mmol), hydrazine hydrochloride (30 μL , 0.95 mmol), copper(II) chloride anhydrous (100 mg, 0.74 mmol) in methanol (5 mL) was stirring for 72 h at room temperature. Then the solvent is evaporated on a rotary evaporator and the product is separated using the methanol:water (4:1) system; yielding 78% of product; m.p. 158-160 °C; IR (V_{\max} , cm^{-1}) 3400, 3380, 3320, 1542: ^1H NMR (300 MHz, CDCl_3 -d) δ_{H} : 4.00-5.00 (m, 4H), 5.18 (broad, 5H), 6.00 (m, 1H), 7.60-7.80 (m, 4H) ppm. ^{13}C

NMR (300 Hz, CDCl₃) δ_C : 59.90, 63.02, 72.32, 122.54, 126.94, 129.70, 146.72, 150.36, 158.24 ppm. EI-MS m/z: 282.09. Anal. Calcd. for C₁₁H₁₄N₄O₅. C, 46.81; H, 5.00; N, 19.85; O, 28.34. Found: C, 46.78; H, 5.00.

2.8. Biological evaluation.

2.8.1. Bacteria strains.

Staphylococcus aureus (ATCC 33591, MRSA), *Streptococcus pneumoniae* (ATCC 6303,) *Escherichia coli* (ATCC 14035), and *Klebsiella pneumoniae* (ATCC 4352) were acquired from the strain bank from Laboratory of Pharmacochemistry, Faculty of Chemical-Biological Sciences of the Autonomous University of Campeche.

2.9. Drugs.

Compounds **2** to **7** were evaluated, and chloramphenicol was used as a control. It is important to mention that the compounds used in this study were dissolved in methanol (stock solution), and all dilutions were carried out with distilled water.

2.10. Antimicrobial activity.

The biological activity produced by compounds **2** to **7** on *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Escherichia coli*, and *Klebsiella pneumoniae* was carried out using the minimum inhibitory concentration (MIC) [23]. In this way, the bacteria were incubated in specific mediums; Staphylococcus 110 agar (*Staphylococcus aureus*), chocolate agar (*Staphylococcus aureus*), and Mc-Conkey (*Escherichia coli* and *Klebsiella pneumoniae*) for 24 h at 37 °C to determine the bacterial growth. This stage was carried out using 12 tubes containing 2 mg/2 ml of culture medium (soybean trypticase); From the first tube, a 1 mg/ml aliquot of chloramphenicol (control) and either of compounds **2** to **7** (1 mg/mL) was added, and the resulting solution was shaken; then, in the next 11 tubes, different dilutions of any of compounds **2** to **7** at a dose of 0.5 to 0.0004 mg/mL were added with constant stirring (Table 1). Then, each tube was inoculated with 0.1 ml of the bacterial suspension, whose concentration corresponded to the McFarland scale (9×10^8 cells/mL). Following, all the tubes were incubated at 37 °C for 24 h. Finally, a loop was taken from each of them, inoculated into the appropriate cultures for different bacterial organisms, and incubated for 24 h at 37 °C.

2.11. Theoretical analysis.

The interaction of chloramphenicol and compounds **2** to **7** with either *Staphylococcus aureus* or *Escherichia coli* was evaluated using both 5fpo and 3zbi proteins surface as theoretical models [24, 25]. In addition, to evaluate the binding energy involved in the interaction of compounds **2** to **7** with either 5fpo or 3zbi proteins surface, chloramphenicol was used as a control on a docking Server software [26, 27].

2.12. Statistical analysis.

Statistical analysis The obtained values are expressed as average \pm SE, using each heart as its control. The data obtained were put under variance (ANOVA) analysis using the

Bonferroni correction factor [28]. The differences were considered significant when p was equal to or smaller than 0.05.

3. Results and Discussion

Some chloramphenicol derivatives have been synthesized to treat different infectious diseases; however, the methods used involve some reagents that may be dangerous or require special conditions such as differences in pH or higher temperatures [18-22]. Analyzing these data, in this research, some chloramphenicol analogs (compounds **2** to **7**) were prepared using some chemical strategies as follows.

3.1. Synthesis of two dioxabicyclo (2 and 3).

There are several reports in the literature on the synthesis of dioxabicyclo derivatives which use some protocols that involve different reagents such as phenylmagnesium bromide [29], phenylpropanoid derivative [30], osmium tetroxide [31], D-proline [32], dianhydromannitol ditosylate [33] and others. In this research, two dioxabicyclo derivatives (**2** and **3**) were prepared *via* reaction of chloramphenicol with either 3,5-dinitrobenzoic acid or 3,5-dinitrofluorobenzene to form **2** or **3** in the presence of dimethyl sulfoxide (Figure 1).

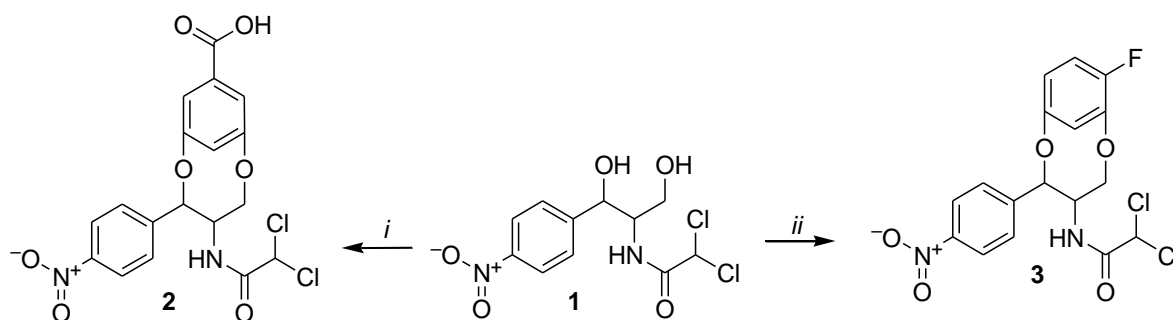


Figure 1. Synthesis of two dioxabicyclo derivatives (**2** and **3**). Reagents and Conditions: *i* = 3,5-dinitrobenzoic acid, potassium carbonate anhydrous, dimethyl sulfoxide, 72 h, 90 °C; *ii* = 3,5-dinitrofluorobenzene, potassium carbonate anhydrous, dimethyl sulfoxide, 72 h, 90 °C.

The ^1H NMR spectrum from **2** showed several bands at 3.32-6.02 and 6.60 and 6.68 ppm for 2,6-Dioxa-bicyclo[5.3.1]undeca-1(10),7(11),8-triene fragment; at 6.22 ppm for methylene bound to amide group; at 6.66 ppm for both carboxyl and amide groups; at 7.90-7.96 ppm for phenyl group bound to nitro group. ^{13}C NMR spectra display chemical shifts at 40.82, 78.54-112.52, 128.14, and 154.36-157.56 ppm for 2,6-Dioxa-bicyclo[5.3.1]undeca-1(10),7(11),8-triene fragment; at 64.42 ppm for ppm for methylene linked to amide group; at 122.94-125.50 and 145.30-146.40 ppm for phenyl group bound to the nitro group; at 161.60 ppm for amide group; at 168.10 ppm for carboxyl group. Besides, the mass spectrum from **2** showed a molecular ion (m/z) 440.01.

On the other hand, the ^1H NMR spectrum from **3** showed several bands at 3.72-5.60, and 6.50-6.96 ppm for 2,6-Dioxa-bicyclo[5.3.1]undeca-1(10),7(11),8-triene fragment; at 6.22 ppm for methylene linked to amide group; at 7.90-7.96 ppm for phenyl group bound to nitro group. ^{13}C NMR spectra display chemical shifts at 42.90, 80.34-116.90, and 149.12-151.30 ppm for 2,6-Dioxa-bicyclo[5.3.1]undeca-1(10),7(11),8-triene fragment; at 64.42 ppm for methylene bound to amide group; at 122.82-146.57 ppm from phenyl group linked to nitro group. In addition, the mass spectrum from **3** showed a molecular ion (m/z) 414.01.

3.2. Preparation of a [1,5]Dioxonane-6,9-dione analog (4).

Several dioxanone derivatives have been synthesized from some reagents such as *o*-phthalaldehyde [34], 2,3-trimethylene-5,6-dihydro-4H-pyran [35], fullerene/FeCl₃ [36]. In this study, a [1,5]Dioxonane-6,9-dione derivative was synthesized via reaction of chloramphenicol with succinic acid using *N,N'*-dicyclohexylcarbodiimide as catalyst (Figure 2). The ¹H NMR spectrum from **4** showed several bands at 2.22-6.22 ppm for 2[1,5]Dioxonane-6,9-dione fragment; at 6.26 ppm for methylene bound to amide group; at 7.74-7.80 ppm for phenyl group bound to the nitro group; at 7.96 ppm from amide group. ¹³C NMR spectra display chemical shifts at 29.70-49.16 and 69.50-79.73 ppm for [1,5]Dioxonane-6,9-dione fragment; at 64.40 ppm for methylene bound to amide group; at 122.85-146.90 ppm for phenyl bound to the nitro group; at 170.12-171.22 ppm for ester groups. Additionally, the mass spectrum from **4** showed a molecular ion (m/z) 404.01.

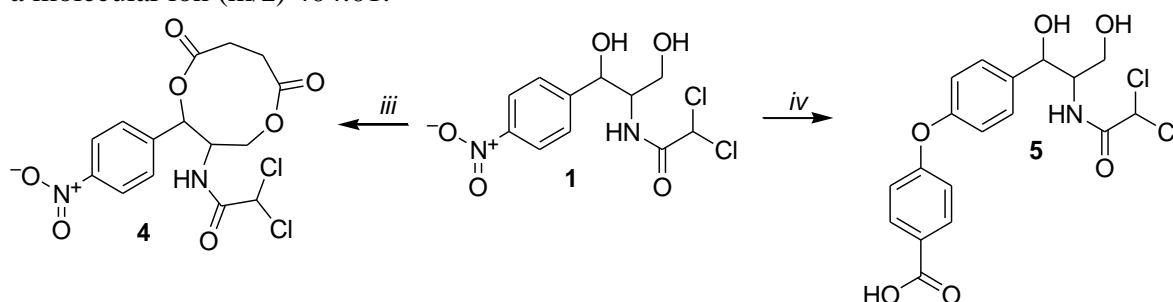


Figure 2. Synthesis of a [1,5]Dioxonane-6,9-dione analog (**4**) and a phenoxy-benzoic acid derivative (**5**). Reagents and Conditions: *iii* = succinic acid, *N,N'*-dicyclohexylcarbodiimide, EtOH, 72 h, room temperature; *iv* = 4-hydroxybenzoic acid, potassium carbonate anhydrous, dimethyl sulfoxide, 72 h, 90 °C.

3.3. Synthesis of an ether derivative (5).

Several protocols for synthesis of some ether derivatives have been reported using different reagents such as Ta/Al₂O₃ [37], palladium [38], tert-butyl nitrite [39], ceric ammonium nitrate [40], dimethyl sulfoxide/potassium carbonate [41]. In this way, compound **5** was synthesized from chloramphenicol, dimethyl sulfoxide, and potassium carbonate anhydrous (Figure 2). The ¹H NMR spectrum from **5** showed several bands at 4.06-5.02 ppm for methylene groups bound to hydroxyl groups; at 6.20 ppm for methylene bound to amide group; at 6.35 ppm for both hydroxyl and amide groups; at 6.66-8.10 ppm for phenyl groups. ¹³C NMR spectra display chemical shifts at 57.70-63.04 ppm for methylene groups linked to hydroxyl groups; at 64.44 ppm for methylene bound to amide group; at 114.82-158.30 ppm for phenyl groups; at 163.22 for amide group; at 168.40 ppm for carboxyl group. Besides, the mass spectrum from **5** showed a molecular ion (m/z) 413.04.

3.4. Synthesis of two acetamides analogs (6 and 7).

There are several method for synthesis of acetamide derivatives: these protocol use some reagents such as iron(II)acetate [42], methyl methoxyacetate [43], triethylphosphine [44], cobalt(III)chloropentaamine chloride [45], hydrochloric acid [46], α -pyridinium acetamide [47] and others. In this research, two acetamide derivatives were synthesized from chloramphenicol and either phenylhydrazine or hydrazine using copper(II) chloride as a catalyst to form the compounds **6** or **7** (Figure 3). The ¹H NMR spectrum from **6** showed several bands at 3.62 ppm for methylene bound to both amide and imino groups; at 4.00-5.00 ppm for methylene groups linked to hydroxyl groups; at 6.00 ppm for amino, hydroxyl, and amide

groups; at 6.44 for imino group; at 6.80-7.26 ppm for phenyl group bound to the amino group; at 7.60-7.80 ppm for phenyl group linked to nitro group. ^{13}C NMR spectra display chemical shifts at 59.90-72.32 ppm for methylene bound to both hydroxyl groups; at 115.60-121.10, 128.44, and 149.12 ppm for phenyl group linked to the amino group; at 122.54-126.94, 146.68 and 150.30 ppm for phenyl bound to the nitro group; at 158.60 ppm for amide group; at 128.22 ppm for imino group. Additionally, the mass spectrum from **6** showed a molecular ion (m/z) 358.12.

Finally, the ^1H NMR spectrum from **7** showed several bands at 4.00-5.00 ppm for methylene groups linked to both hydroxyl groups; at 5.18 ppm for amide, hydroxyl, and amino groups; at 6.00 ppm for imino group; at 7.60-7.80 ppm for phenyl group. ^{13}C NMR spectra display chemical shifts at 59.90-72.32 ppm for methylene groups linked to both hydroxyl groups; at 122.54-126.94 and 146.72-150.36 ppm for phenyl group; at 125.70 ppm for imino group; at 158.24 ppm for amide group. Finally, the mass spectrum from **7** showed a molecular ion (m/z) 282.09.

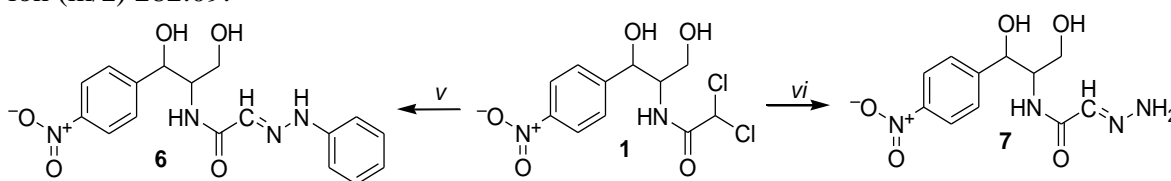


Figure 3. Synthesis of two acetamide derivatives (**6** and **7**). *Reagents and Conditions:* v = phenylhydrazine hydrochloride, Copper(II) chloride anhydrous, 72 h, room temperature; vi = hydrazine hydrochloride, Copper(II) chloride anhydrous, 72 h, room temperature.

3.5. Biological activity.

For several years, chloramphenicol has been used to treat some infectious diseases; however, this drug can produce some secondary effects such as aplastic anemia [48], anaphylaxis [49], bone marrow suppression [50], Gray syndrome [51], acidosis [52]. Besides, some studies indicate that chloramphenicol can induce bacterial resistance in some bacterial strains such as *Staphylococcus aureus* [53] and *Escherichia coli* [54, 55]. To evaluate the biological activity of chloramphenicol derivatives (compounds **2** to **7**) on *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Escherichia coli*, and *Klebsiella pneumoniae*, chloramphenicol was used as a control using the minimum inhibitory concentration (MIC) method. It is important to mention that dilutions of the different compounds studied (mg/mL) (Table 1) are expressed in the results as mmol/mL. The results (Figure 4) showed that bacterial growth of either *Staphylococcus aureus* or *Streptococcus pneumoniae* was inhibited by chloramphenicol (1.54×10^{-3} mmol/mL) and compound **6** (2.4×10^{-3} mmol/mL); however, this effect was higher in the presence of compound **4** (1.20×10^{-3} mmol/mL).

These data suggest that phenylhydrazine fragment bound to acetamide group could be the responsibility of the antibacterial effect of compound **6**. This hypothesis is supported by other studies which indicate that some acetamide derivatives exert antibacterial activity against *Staphylococcus aureus* [56-58] and *Streptococcus pneumoniae* [59-62]. However, the biological activity of **4** was different to **6**; these data suggest that [1,5]Dioxane-6,9-dione fragment can exert higher antibacterial activity compared with acetamide groups. Here it is important to mention that some studies showed that some dioxo-dione derivatives decrease of bacterial growth of Gram-positive bacteria [63, 64].

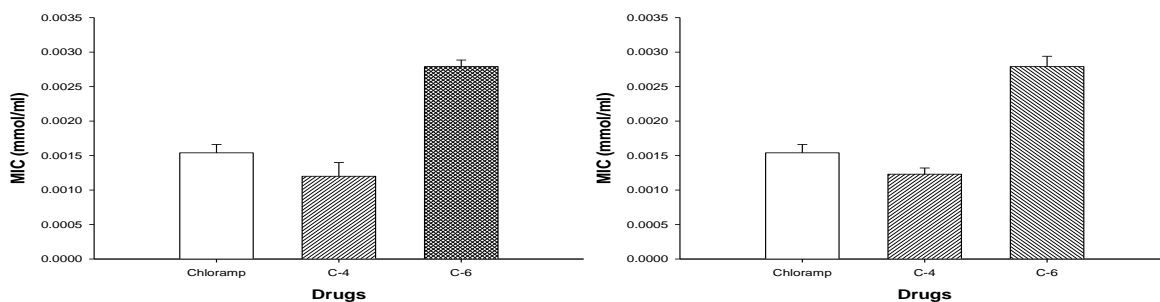


Figure 4. The biological activity produced by chloramphenicol (Chloramp) and compounds 4 and 6 against either *Staphylococcus aureus* (Left) or *Streptococcus pneumoniae* (Right). The results showed that bacterial growth of this microorganism was significantly inhibited ($p = 0.05$) by compound 4 compared with chloramphenicol and compound 6. Each bar represents the mean \pm S.E. of 9 experiments.

On the other hand, other experiments were carried out to evaluate the biological activity of chloramphenicol and compounds 2 to 7 against *Escherichia coli* and *Klebsiella pneumoniae*. The results (Figure 5) showed that compound 4 significantly decreased the bacterial growth of Gram-negative bacteria compared to compound 6 and chloramphenicol. All these data suggest that these chloramphenicol derivatives could interact with some biological molecule involved in the surface of Gram-positive or Gram-negative bacteria, which may decrease the bacterial growth of these microorganisms.

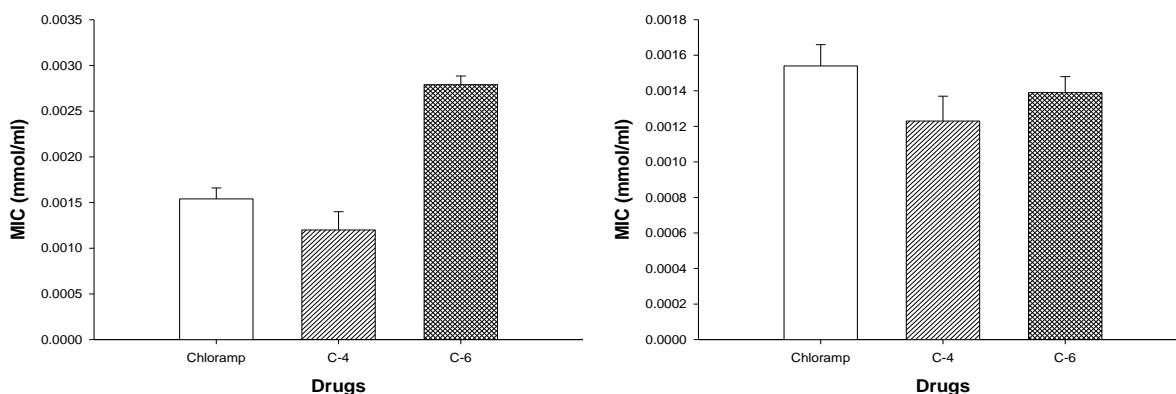


Figure 5. Antibacterial activity exerted by chloramphenicol (Chloramp) and compounds 4 and 6 against either *Klebsiella pneumoniae* (Left) or *Escherichia coli* (Right). The data showed that the bacterial growth of Gram-negative bacteria was significantly inhibited ($p = 0.05$) by compounds 4 and 6 compared with chloramphenicol. Each bar represents the mean \pm S.E. of 9 experiments.

3.6. Ligand-protein interaction.

Various theoretical models are used to predict the interaction of different compounds with the surface of several proteins or enzymes; these models involve some parameters such as free binding energies and solvation energies [65-67]. For example, the Dockingserver software was used to analyze the interaction of some compounds such as coumarin-thiopropionic acids [68], (1,2,3-triazole) [69], anthranilic acid derivatives [70], Pyrazolo[5, 1-c][1, 2, 4]triazoles [71] with both Gram-positive and Gram-negative bacteria. In this way, it was decided to use 5fpo and 3zbi proteins as theoretical models (Protein Data Bank) [25, 26] to evaluate the interaction of compounds 2 to 7 with either *Staphylococcus aureus* or *Escherichia coli* bacteria. The results (Tables 1 and 2) showed different amino acid residues involved in the interaction of compounds 4 and 6 with either 5fpo or 3zbi proteins surface compared with compounds 2, 3, 5, 7, and chloramphenicol; this phenomenon could be due to differences in their chemical

structure. Besides, other theoretical data showed differences in some thermodynamic parameters involved in the interaction of **4** and **6** with either 5fpo or 3zbi proteins compared with compounds **2**, **3**, **5**, **7**, and chloramphenicol (Tables 3 and 4).

Table 1. Interaction of compound 4 and indomethacin (control) with 5fpo-protein surface.

Chloramphenicol	Compound 2	Compound 3	Compound 4	Compound 5	Compound 6	Compound 7
Met79	Leu82	Met79	Leu82	Leu80	Leu82	Leu80
Leu80	Asn84	Leu80	Glu110	Leu82	Glu110	Leu82
Leu82	Lys112	Ser81	Lys112	Leu111	Leu111	Lys112
Glu110	Ala117	Leu82	Ala117	Lys112	Lys112	Glu167
Lys112	Arg133	Lys112	Arg133	Arg133	Ile113	Tyr219
Ala117	Glu167	Ala117	Glu167	Glu167	Glu167	Val281
Arg133	Arg194	Arg133	Arg194	Tyr219	Phe217	Lys283
Glu167	Tyr219	Glu167	Tyr219	Asp278	Tyr219	
Tyr219	Lys307	Tyr219	Val281	Val281	Asn247	
Val281				Lys283	Val281	
				Lys307	Lys283	

Table 2. Interaction of chloramphenicol and compounds 2 to 7 with 3zbi -protein surface.

Chloramphenicol	Compound 2	Compound 3	Compound 4	Compound 5	Compound 6	Compound 7
Ile791	Ile791	Thr792	Phe803	Ile791	Ile791	Ile791
Thr792	Thr792	Phe803	Leu805	Thr792	Thr792	Thr792
Phe803	Phe803	Leu805	Ala823	Ile795	Gly793	Gly793
Val804	Val804	Ala823	Ala831	Lys796	Ile795	Val802
Leu805	Leu805		Phe888	Gln799	Val802	Phe803
Ala823	Ala823			Arg801	Phe803	Ala823
				Val802	Leu805	
				Phe803	Ala823	
				Ala823		

Table 3. Thermodynamic parameters involved in the interaction of chloramphenicol and their derivatives with the 5fpo-protein surface.

Compound	Est. Free energy of Binding (kcal/mol)	Free of Inhibition Constant (Ki)	Est. Inhibition Constant (Ki)	vdW + H-bond + desolv Energy (kcal/mol)	Electrostatic Energy (kcal/mol)	Total Interm. Energy	Interact Surface
Chloramphenicol	-7.84		1.78	-8.89	0.05	-8.84	702.91
2	-5.32		125.63	-6.76	-0.04	-6.81	853.63
3	-5.31		128.09	-6.94	0.03	-6.92	860.66
4	-7.07		6.62	-7.87	0.04	-7.83	803.93
5	-5.98		41.42	-6.36	-0.88	-7.24	851.29
6	-7.84		1.79	-7.71	-0.20	-7.91	759.21
7	-8.79		10.47	-7.94	-0.08	-8.02	692.93

Table 4. Thermodynamic parameters involved in the interaction of chloramphenicol and their derivatives with the 3zbi-protein surface.

Compound	Est. Free energy of Binding (kcal/mol)	Free of Inhibition Constant (Ki)	Est. Inhibition Constant (Ki)	vdW + H-bond + desolv Energy (kcal/mol)	Electrostatic Energy (kcal/mol)	Total Interm. Energy	Interact Surface
Chloramphenicol	-5.82		54.16	-5.99	0.01	-5.98	423.32
2	-5.46		98.85	-7.00	0.04	-6.95	550.00
3	-5.49		94.60	-6.57	0.01	-6.57	518.97
4	-6.55		15.82	-7.64	-0.02	-7.49	591.75
5	-5.74		61.87	-7.79	-0.09	-7.88	649.74
6	-5.86		50.62	-7.66	-0.01	-7.68	603.96
7	-5.29		133.07	-5.74	-0.02	-5.76	479.98

4. Conclusions

In this study, an easy synthesis of some chloramphenicol derivatives (compounds **2** to **7**) is reported: it is noteworthy that these methods do not require special conditions or dangerous reagents. Besides, both compounds **4** and **6** showed antibacterial activity against either Gram-positive or Gram-negative bacteria; therefore, these chloramphenicol derivatives could be a good candidate as the bacterial agent.

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Conflict of Interest

We declare that this manuscript does not have any conflict of financial interests (political, personal, religious, ideological, academic, intellectual, commercial, or otherwise) for its publication.

References

1. Van-Kampen, J.; Van de Vijver, D.; Fraaij, P. et al. Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 (COVID-19). *Nature communications* **2021**, *12*, 267, <https://doi.org/10.1038/s41467-020-20568-4>.
2. Johnson, S.; Lavergne, V.; Skinner, A.; Gonzales-Luna, A.; Garey, K.W.; Kelly, C.P.; Wilcox, M.H. Clinical practice guideline by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA): 2021 focused update guidelines on management of Clostridioides difficile infection in adults. *Clinical Infectious Diseases* **2021**, *73*, e1029-e1044, <https://doi.org/10.1093/cid/ciab549>.
3. Planas, D.; Bruel, T.; Grzelak, L. et al. Sensitivity of infectious SARS-CoV-2 B.1.1.7 and B.1.351 variants to neutralizing antibodies. *Nature Medicine* **2021**, *27*, 917-924, <https://doi.org/10.1038/s41591-021-01318-5>.
4. Bruyndonckx, R.; Adriaenssens, N.; Hens, N.; Versporten, A.; Monnet, D.; Molenberghs, G.; Goossens, H.; Weist, K.; Coenen, S, the ESAC-Net study group. Consumption of penicillins in the community, European Union/European Economic Area, 1997-2017. *Journal of Antimicrobial Chemotherapy* **2021**, *76*, ii14-ii21, <https://doi.org/10.1093/jac/dkab173>.
5. Parsels, K.; Mastro, K.; Steele, J.; Thomas, S.; Kufel, W. Cefiderocol: a novel siderophore cephalosporin for multidrug-resistant Gram-negative bacterial infections. *Journal of Antimicrobial Chemotherapy* **2021**, *76*, 1379-1391, <https://doi.org/10.1093/jac/dkab015>.
6. Deelen, J.; Rottier, W.; Buiting, A.; Dorigo-Zetsma, J.; Kluytmans, J.; Van der Linden, P.; Thijsen, S.F.T.; Vlamincx, B.J.M.; Weersink, A.J.L.; Ammerlaan, H.S.M.; Bonten, M.J.M.; Van Werkhoven, C.H. Short-course aminoglycosides as adjunctive empirical therapy in patients with Gram-negative bloodstream infection, a cohort study. *Clinical Microbiology and Infection* **2021**, *27*, 269-275, <https://doi.org/10.1016/j.cmi.2020.04.041>.
7. Cegielski, J.; Chan, P.; Lan, Z.; Udawadia, Z.; Viiklepp, P.; Yim, J.; Menzies, D. Aminoglycosides and capreomycin in the treatment of multidrug-resistant tuberculosis: individual patient data meta-analysis of 12 030 patients from 25 countries, 2009–2016. *Clinical Infectious Diseases* **2021**, *73*, e3929-e3936. <https://doi.org/10.1093/cid/ciaa621>.
8. Ontong, J.; Ozioma, N.; Voravuthikunchai, S.; Chusri, S. Synergistic antibacterial effects of colistin in combination with aminoglycoside, carbapenems, cephalosporins, fluoroquinolones, tetracyclines, fosfomycin, and piperacillin on multidrug resistant *Klebsiella pneumoniae* isolates. *Plos one* **2021**, *16*, 0244673, <https://doi.org/10.1371/journal.pone.0244673>.
9. El-Shabrawy, M.; Abdel-Reheem, A.; Al-Hakeem, F.; Emam, M. Corrosion Resistance of Concrete Specimens subjected to Sulphate Solution. *Mansoura Engineering Journal* **1987**, *12*, 30-39.

10. Theriault, N.; Tillotson, G.; Sandrock, C. Global travel and Gram-negative bacterial resistance; implications on clinical management. *Expert Review of Anti-infective Therapy* **2021**, *19*, 181-196, <https://doi.org/10.1080/14787210.2020.1813022>.
11. Zhang, Q.; Chu, X.; Buckling, A. Overcoming the growth–infectivity trade-off in a bacteriophage slows bacterial resistance evolution. *Evolutionary Applications* **2021**, *14*, 2055-2063, <https://doi.org/10.1111/eva.13260>.
12. Islam, M. Bacterial resistance to antibiotics: access, excess, and awareness in Bangladesh. *Expert Review of Anti-infective Therapy* **2021**, *19*, 973-981, <https://doi.org/10.1080/14787210.2021.1865804>.
13. Fraiture, M.; Deckers, M.; Papazova, N.; Roosens, N. Detection strategy targeting a chloramphenicol resistance gene from genetically modified bacteria in food and feed products. *Food Control* **2020**, *108*, 106873, <https://doi.org/10.1016/j.foodcont.2019.106873>.
14. Ruan, S.; Luo, J.; Zhang, Y.; Nordhaus, I.; Diao, X.; Zhao, H. Spatial and seasonal variations of chloramphenicol resistance genes and their co-occurring analysis with bacteria in a typical mangrove area of Hainan Island, China. *Estuarine, Coastal and Shelf Science* **2020**, *233*, 106541, <https://doi.org/10.1016/j.ecss.2019.106541>.
15. Ma, X.; Liang, B.; Qi, M.; Yun, H.; Shi, K.; Li, Z.; Guo, Y.; Yan, P.; Liu, S.-J.; Wang, A. Novel pathway for chloramphenicol catabolism in the activated sludge bacterial isolate *Sphingobium* sp. CAP-1. *Environment Science & Technology* **2020**, *54*, 7591-7600, <https://doi.org/10.1021/acs.est.9b07324>.
16. Wang, C.; Dong, X.; Yang, Y. et al. Metabolites in the TCA Cycle Promote Resistance to Chloramphenicol of *Edwardsiella tarda*. *Journal of Proteome Research* **2020**, *20*, 972-981, <https://doi.org/10.1021/acs.jproteome.0c00725>.
17. Goodale, A.; Michailidis, F.; Watts, R.; Chok, S.; Hayes, F. Characterization of permissive and non-permissive peptide insertion sites in chloramphenicol acetyltransferase. *Microbial Pathogenesis* **2020**, *149*, 104395, <https://doi.org/10.1016/j.micpath.2020.104395>.
18. Neu, H.; Fu, K. In vitro activity of chloramphenicol and thiamphenicol analogs. *Antimicrobial Agents and Chemotherapy* **1980**, *18*, 311-316, <https://doi.org/10.1128/AAC.18.2.311>.
19. Vince, R.; Almquist, R.; Ritter, C.; Daluge, S. Chloramphenicol binding site with analogues of chloramphenicol and puromycin. *Antimicrobial Agents and Chemotherapy* **1975**, *8*, 439-443, <https://doi.org/10.1128/AAC.8.4.439>.
20. Tereshchenkov, A.; Dobosz-Bartoszek, M.; Osterman, I.A. et al. Binding and action of amino acid analogs of chloramphenicol upon the bacterial ribosome. *Journal of Molecular Biology* **2018**, *430*, 842-852, <https://doi.org/10.1016/j.jmb.2018.01.016>.
21. Kono, M.; O'Hara, K.; Nagawa, M.; Mitsuhashi, S. Antibacterial activity of chloramphenicol-related compounds toward a chloramphenicol-resistant strains of *Staphylococcus aureus*. *Japanese Journal of Microbiology* **1972**, *16*, 461-467, <https://doi.org/10.1111/j.1348-0421.1972.tb00685.x>.
22. Nasrin, S.; Ganji, S.; Kakirde, K. et al. Chloramphenicol derivatives with antibacterial activity identified by functional metagenomics. *Journal of Natural Products* **2018**, *81*, 1321-1332, <https://doi.org/10.1021/acs.jnatprod.7b00903>.
23. Figueroa-Valverde, L.; Díaz-Cedillo, F.; Camacho-Luis, A.; Ramos, M.; Cervera, E. Synthesis of a dihydrotestosterone–ciprofloxacin conjugate: relationship between descriptors log P, π , R m, and V m and its antibacterial activity in *S. aureus* and *E. coli*. *Monatshefte für Chemie-Chemical Monthly* **2010**, *141*, 373-380, <http://dx.doi.org/10.1007/s00706-010-0263-y>.
24. Ludlow, R.; Verdonk, M.; Saini, H.; Tickle, I.; Jhoti, H. Detection of secondary binding sites in proteins using fragment screening. *Proceedings of National Academy Sciences* **2015**, *112*, 15910-15915, <https://doi.org/10.1073/pnas.1518946112>.
25. Rivera-Calzada, A.; Fronzes, R.; Savva, C.; Chandran, V.; Lian, P.; Laeremans, T.; Pardon, E.; Steyaert, J.; Remaut, H.; Waksman, G.; Orlova, E. Structure of a bacterial type IV secretion core complex at subnanometre resolution. *The EMBO Journal* **2013**, *32*, 1195-1204, <https://doi.org/10.1038/emboj.2013.58>.
26. Torchala, M.; Gerguri, T.; Chaleil, R.; Gordon, P.; Russell, F.; Keshani, M.; Bates, P.A. Enhanced sampling of protein conformational states for dynamic cross-docking within the protein-protein docking server SwarmDock. *Proteins: Structure, Function, and Bioinformatics* **2020**, *88*, 962-972, <https://doi.org/10.1002/prot.25851>.
27. Kong, R.; Yang, G.; Xue, R.; Liu, M.; Wang, F.; Hu, J.; Chang, S. COVID-19 Docking Server: A meta server for docking small molecules, peptides and antibodies against potential targets of COVID-19. *Bioinformatics* **2020**, *36*, 5109-5111, <https://doi.org/10.1093/bioinformatics/btaa645>.
28. Hocht, C.; Opezzo, J.; Gorzalczany, S.; Bramuglia, G.; Tiara, C. Una aproximación cinética y dinámica de metildopa en ratas con coartación aórtica mediante microdiálisis. *Revista Argentina de Cardiología* **1999**, *67*, 769-773.
29. Eiden, F.; Denk, F. Synthesis of CNS-activity of pyran derivatives: 6, 8-dioxabicyclo (3,2,1) octane. *Arch Pharm (Weinheim)* **1991**, *324*, 353-354, <https://doi.org/10.1002/ardp.19913240606>.
30. Yin, G.; Ren, T.; Rao, Y.; Zhou, Y.; Li, Z.; Shu, W.; Wu, A. Stereoselective synthesis of 2, 8-dioxabicyclo [3.3.1] nonane derivatives via a sequential Michael addition/bicyclization reaction. *Journal of Organic Chemistry* **2013**, *78*, 3132-3141, <https://doi.org/10.1021/jo400081q>.

31. Valente, P.; Avery, T.; Taylor, D.; Tiekink, E. Synthesis and Chemistry of 2, 3-Dioxabicyclo [2.2. 2] octane-5, 6-diols. *Journal of Organic Chemistry* **2009**, *74*, 274-282, <https://doi.org/10.1021/jo8020506>.
32. Kim, I.; Kim, S.; Choi, J.; Lee, G. Facile synthesis of benzo-fused 2, 8-dioxabicyclo[3.3.1]nonane derivatives via a domino Knoevenagel condensation/hetero-Diels-Alder reaction sequence. *Tetrahedron* **2008**, *64*, 664-671, <https://doi.org/10.1016/j.tet.2007.11.036>.
33. Archibald, T.; Baum, K. Synthesis of Polynitro-2, 6-dioxabicyclo [3.3.0] octanes. *Synthetic Communications* **1989**, *19*, 1493-1498, <https://doi.org/10.1080/00397918908051042>.
34. Lemiègre, L.; Lesetre, F.; Combret, J.; Maddaluno. Synthesis of α , β -unsaturated dioxanes, dioxolanes and dioxepanes by trans-acetalisation of dimethylacetals with meso or C2-symmetrical 1,2-,1,3-and 1,4-diols. *Tetrahedron* **2004**, *60*, 415-427, <https://doi.org/10.1016/j.tet.2003.11.009>.
35. Gagan J.M.F. Compounds with Larger Heterocyclic Rings: Compounds with Rings of More than Eight Atoms. In: *Rodd's Chem Carbon Comp.* **1964**, 427-466, Elsevier, <https://doi.org/10.1016/B978-044453345-6.50812-9>.
36. Zhai, W.; Jiang, S.; Peng, R.; Jin, B.; Wang, G. Facile Access to Novel [60] Fulleranyl Diethers and [60] Fullerene-Sugar Conjugates via Annulation of Diol Moieties. *Organic Letters* **2015**, *17*, 1862-1865, <https://doi.org/10.1021/acs.orglett.5b00536>.
37. Herrera, C.; Cortes-Reyes, M.; Larrubia, M.; Domínguez-Barroso, M.; Díaz-Rey, M.; Alemany, L. Dimethyl ether synthesis via methanol dehydration over Ta-supported catalysts. *Applied Catalysis A: General* **2019**, *582*, 117088, <https://doi.org/10.1016/j.apcata.2019.05.022>.
38. Chen, X.; Xiao, X.; Sun, H.; Li, Y.; Cao, H.; Zhang, X.; Yang, S.; Lian, Z. Transition-Metal-Catalyzed Transformation of Sulfonates via S-O Bond Cleavage: Synthesis of Alkyl Aryl Ether and Diaryl Ether. *Organic Letters* **2019**, *21*, 8879-8883, <https://doi.org/10.1021/acs.orglett.9b02858>.
39. Ma, X.; Song, Q. Tert-Butyl Nitrite Mediated Synthesis of Fluorinated O-Alkyloxime Ether Derivatives. *Organic Letters* **2019**, *21*, 7375-7379, <https://doi.org/10.1021/acs.orglett.9b02689>.
40. Chavan, S.; Kadam, A.; Lasonkar, P.; Gonnade, R. Synthesis of 3-Azidopiperidine Skeleton Employing Ceric Ammonium Nitrate (CAN)-Mediated Regioselective Azidoalkoxylation of Enol Ether: Total Synthesis of D₂ Receptor Agonist (\pm)-Quinagolide. *Organic Letters* **2018**, *20*, 7011-7014, <https://doi.org/10.1021/acs.orglett.8b02900>.
41. López-Ramos, M.; Figueroa-Valverde, L.; Herrera-Meza, S.; Rosas-Nexticapa, M.; Díaz-Cedillo, F.; García-Cervera, E.; Cahuich-Carrillo, R. Design and synthesis of a new steroid-macrocyclic derivative with biological activity. *Journal of Chemical Biology* **2017**, *10*, 69-84, <https://doi.org/10.1007/s12154-017-0165-0>.
42. Wenjun, T.; Nitinchandra, P.; Cappacci, A. et al. Synthesis of N-Acetyl Enamide by Reductive Acetylation of Oximes Mediated with Iron(II) Acetate: N-(4-Bromophenyl)vinyl)acetamide. *Organic Synth.* **2013**, *90*, 62-73, <https://doi.org/10.15227/orgsyn.090.0062>.
43. Thalen, L.K.; Rosch, C.; Backvall, J.E. Synthesis of 2-Methoxy-N-(1-Phenylethyl)Acetamide Via Dynamic Kinetic Resolution. *Organic Synth.* **2012**, *89*, 255-266, <https://doi.org/10.1002/0471264229.os089.25>.
44. Zhao, H.; Vandenbossche, C.; Koenig, S.G. et al. Synthesis of Enamides from Ketones: Preparation of N-(3,4-Dihydronaphthalene-1-yl)Acetamide. *Organic Synth.* **2010**, *87*, 275-287, <https://doi.org/10.1002/0471264229.os087.30>.
45. Anderson, C.; Overman, L.; Watson, M. Asymmetric Rearrangement of Allylic Trichloroacetamides: Preparation of (S)-2,2,2-Trichloro-N-(1-Propylallyl)Acetamide. *Organic Synth.* **2005**, *82*, 134-139, <https://doi.org/10.15227/orgsyn.082.0134>.
46. Hocker, J.; Gieseke, H. N-(2,4-diformyl-5-Hydroxyphenyl)Acetamide. *Organic Synth.* **1991**, *60*, 49, <https://doi.org/10.1002/0471264180.os060.11>.
47. Kojima, S.; Hiroike, K.; Ohkata, K. Stereoselective synthesis of activated cyclopropanes with an α -pyridinium acetamide bearing an 8-phenylmenthyl group as the chiral auxiliary. *Tetrahedron Letters* **2004**, *45*, 3565-3568, <https://doi.org/10.1016/j.tetlet.2004.03.044>.
48. Feder, H.; Osier, C.; Maderazo, E. Chloramphenicol: a review of its use in clinical practice. *Reviews of Infectious Diseases* **1981**, *3*, 479-491, <https://doi.org/10.1093/clinids/3.3.479>.
49. Palchick, B.; Funk, E.; McEntire, J.; Hamory, B. Anaphylaxis due to chloramphenicol. *American Journal of the Medical Sciences* **1984**, *288*, 43-45, <https://doi.org/10.1097/0000441-198407000-00011>.
50. Yunis, A. Chloramphenicol toxicity: 25 years of research. *American Journal of Medicine* **1989**, *87*, 44N-48N.
51. Geme, J. On the "gray syndrome" and chloramphenicol toxicity. *Pediatrics* **1960**, *25*, 1088-1090, <https://doi.org/10.1542/peds.25.6.1088a>.
52. Evans, I.; Kleiman, M. Acidosis as a presenting feature of chloramphenicol toxicity. *Journal of Pediatrics* **1986**, *108*, 475-477, [https://doi.org/10.1016/S0022-3476\(86\)80905-2](https://doi.org/10.1016/S0022-3476(86)80905-2).
53. Shaw, W.; Brodsky, R. Characterization of chloramphenicol acetyltransferase from chloramphenicol-resistant *Staphylococcus aureus*. *Journal of Bacteriology* **1968**, *95*, 28-36, <https://doi.org/10.1128/jb.95.1.28-36.1968>.
54. Crofts, T.; Sontha, P.; King, A.; Wang, B.; Bidy, B.; Zanolli, N.; Gaumnitz, J.; Dantas, G. Discovery and characterization of a nitroreductase capable of conferring bacterial resistance to chloramphenicol. *Cell Chemical Biology* **2019**, *26*, 559-570, <https://doi.org/10.1016/j.chembiol.2019.01.007>.

55. Eagle, H.; Fleischman, R.; Levy, M. Development of increased bacterial resistance to antibiotic I: Continuous Spectrum of Resistance to Penicillin, Chloramphenicol, and Streptomycin. *Journal of Bacteriology* **1952**, *63*, 623-638, <https://doi.org/10.1128/jb.63.5.623-638.1952>.
56. Rybak, M.; Gudzera, O.; Gorbatiuk, O.; Usenko, M.; Yarmoluk, S.; Tukalo, M.; Volynets, G. Rational Design of Hit Compounds Targeting *Staphylococcus aureus* Threonyl-tRNA Synthetase. *ACS Omega* **2021**, *6*, 24910-24918, <https://doi.org/10.1021/acsomega.1c03789>.
57. Hakim, F.; Salfidoer, S. Antibacterial and Antitubercular Activity of Novel Benzothiazole-Aryl Amine Derivatives Tethered through Acetamide Functionality. *Asian Journal of Chemistry* **2021**, *33*, 1757-1763.
58. Vlasov, S.; Borysov, O.; Severina, H.; Kovalenko, S.; Osolodchenko, T.; Vlasov, V.; Georgiyants, V. The synthesis, antimicrobial activity and docking studies of 6-(1H-benzimidazol-2-yl)-5-methylthieno [2, 3-d] pyrimidin-4 (3H)-ones with acetamide and 1, 2, 4-oxadiazol-5-ylmethyl substituents. *Journal of Organic and Pharmaceutical Chemistry* **2021**, *19*, 15-20, <http://dx.doi.org/10.24959/ophcj.21.240775>.
59. Yan, F.; Dang, Q.; Liu, C.; Yan, J.; Wang, T.; Fan, B.; Cha, D.; Li, X.; Liang, S.; Zhang, Z. 3,6-O-[N-(2-Aminoethyl)-acetamide-yl]-chitosan exerts antibacterial activity by a membrane damage mechanism. *Carbohydrate Polymers* **2016**, *149*, 102-111, <https://doi.org/10.1016/j.carbpol.2016.04.098>.
60. Paget, S.; Boggs, C.; Foleno, B.; Goldschmidt, R.; Hlasta, D.; Weidner-Wells, M.; Werblood, H.; Bush, K.; Macielag, M. Antibacterial activity of pyrrolopyridine-substituted oxazolidinones: synthesis and in vitro SAR of various C-5 acetamide replacements. *Bioorganic & Medicinal Chemistry Letters* **2006**, *16*, 4537-4542, <https://doi.org/10.1016/j.bmcl.2006.06.023>.
61. Ghazzali, M.; El-Faham, A.; Abdel-Megeed, A.; Al-Farhan, K. Microwave-assisted synthesis, structural elucidation and biological assessment of 2-(2-acetamidophenyl)-2-oxo-N phenyl acetamide and N-(2-(2-oxo-2 (phenylamino) acetyl) phenyl) propionamide derivatives. *Journal of Molecular Structure* **2012**, *1013*, 163-167, <https://doi.org/10.1016/j.molstruc.2012.01.028>.
62. Nomura, T.; Iwaki, T.; Yasukata, T.; Nishi, K.; Narukawa, Y.; Uotani, K.; Hori, T.; Miwa, H. A new type of ketolides bearing an N-aryl-alkyl acetamide moiety at the C-9 iminoether synthesis and structure-activity relationships. *Bioorganic & Medicinal Chemistry* **2005**, *13*, 6615-6628, <https://doi.org/10.1016/j.bmc.2005.07.041>.
63. Nomura, T.; Iwaki, T.; Narukawa, Y.; Uotani, K.; Hori, T.; Miwa, H. A new type of ketolide bearing an N-aryl-alkyl acetamide moiety at the C-9 iminoether: Synthesis and structure-activity relationships. *Bioorganic & Medicinal Chemistry* **2006**, *14*, 3697-3711, <https://doi.org/10.1016/j.bmc.2006.01.036>.
64. Kanagarajan, V.; Thanusu, J.; Gopalakrishnan, M. Synthesis and in vitro microbiological evaluation of an array of biolabile 2-morpholino-N-(4, 6-diarylpyrimidin-2-yl) acetamides. *European Journal of Medicinal Chemistry* **2010**, *45*, 1583-1589, <https://doi.org/10.1016/j.ejmech.2009.12.068>.
65. Seyedi, S.; Sadeghian, A.; Sadeghian, H.; Hazrathoseyni, A.; Sadeghian, M. Synthesis and biological evaluation of some new thioether-ester crown ethers. *Phosphorus, Sulfur and Silicon and the Related Elements* **2007**, *182*, 265-272, <https://doi.org/10.1080/10426500600917060>.
66. Shoichet, B.; Leach, A.; Kuntz, I. Ligand solvation in molecular docking. *Proteins: Structure, Function and Bioinformatics* **1999**, *34*, 4-16, [https://doi.org/10.1002/\(sici\)1097-0134\(19990101\)34:1%3C4::aid-prot2%3E3.0.co;2-6](https://doi.org/10.1002/(sici)1097-0134(19990101)34:1%3C4::aid-prot2%3E3.0.co;2-6).
67. Jackson, R.; Gabb, H.; Sternberg, M. Rapid refinement of protein interfaces incorporating solvation: application to the docking problem. *Journal of Molecular Biology* **1998**, *276*, 265-285, <https://doi.org/10.1006/jmbi.1997.1519>.
68. Palanisamy, P.; Subramanian, R.; Sankari, S.; Kumaresan, S. Coumarin thiopropionic Acids: Syntheses, Characterization, and Comparison of Docking-DFT Studies on their Antimicrobial Activity. *World Journal of Pharmaceutical Sciences* **2017**, *1*, 96-103.
69. Bitla, S.; Gayatri, A.; Puchakayala, M. et al. Design and synthesis, biological evaluation of bis-(1, 2, 3-and 1, 2, 4)-triazole derivatives as potential antimicrobial and antifungal agents. *Bioorganic of Medicinal Chemistry Letters* **2021**, *41*, 128004, <https://doi.org/10.1016/j.bmcl.2021.128004>.
70. Grant, N. Anthranilic Acid Derivatives as Novel Antibiotics against MRSA and other Gram-positive Microorganisms: Combating Antibiotic Resistance. *McNair Scholars Journal* **2013**, *17*, 6-12.
71. Setiawan, T.; Rizarullah, R. Predicting Multi-Epitope Peptide Cancer Vaccine from Novel TAA Topo48. *Journal of Science and Applicative Technology* **2021**, *5*, 171-178.