

Synthesis, Spectral Characterisation of Novel Azo–Dye 4-((E)-(2-Hydroxy-3-Methoxy-5-((E)-Thiazol-2-Yldiazenyl)Benzylidene)Amino)-1,5-Dimethyl-2-Phenyl-1H-Pyrazol-3(2H)-One and its Transition Metal Complexes; Differential Pulse Voltammetric Detection of Nitrite and its Biological Activities

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Received: 3.10.2022; Accepted: 19.11.2022; Published: 5.02.2023

Abstract: The novel azo-azomethine 4-((E)-(2-hydroxy-3-methoxy-5-((E)-thiazol-2-ylidiazanyl)benzylidene)amino)-1,5-dimethyl-2-phenyl-1H-pyrazol-3(2H)-one (HTDP) and its Cu(II), Co(II) and Ni(II) complexes were synthesized, and their structure was confirmed by using physicochemical techniques. The spectrochemical data indicate that the synthesized ligand is tridentate in nature. The antimicrobial, anti-diabetes (A-DM), and anti-mycobacterium tuberculosis (M-TB) activities against ligands and their metal complexes were screened. The complexes exhibited enhanced bio-efficiency inhibitory results when compared to the HTDP. Excess nitrate poses harmful effects on the ecological system and causes the blue baby syndrome. The modified glassy carbon electrode prepared using [Co(HTDP)₂] is used for electrochemical sensing of nitrate under different concentrations and displayed good electro-catalytic activity with long linear range (10-80 μmolL⁻¹), sensitivity (0.172 μAμM⁻¹ cm⁻²) and the LOD (limit of detection) found to be 3.333 μML⁻¹ in PBS (pH 7) Further, the prepared sensor has more stability and showed less poisoning property.

Keywords: metal complexes; TGA; nitrate; cyclic voltammetry; DPV; antimicrobial; antidiabetic and anti-tuberculosis activities.

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1. Introduction

The heterocyclic compounds with at least two different donor atoms, like sulfur and nitrogen, have been studied widely due to their various biological activities [1-3]. The Schiff base (R₁-C=N-R₂) compounds have a group-specific condensation reaction for aldehyde/ketone with an amine. Coordination of ligand to metal ions occurs via azomethine nitrogen. The heterocyclic donor elements and azomethine group increase the potential applications for therapeutic activities. The azo Schiff bases are color chromophoric which are used in the synthetic colorant in cosmetics, fabrication, natural health products, insecticides,

recording layers of DVD, CD, textile cilium, plastics, leather-fell, metal sheets, food, and toys, etc. [4-6]. The chelating ligands coordinating with metal ions exhibit enhanced physicochemical and catalytic behavior. The transition metal based-drugs with lesser toxicity can be used as target-specific activities with a wide range of potential therapeutic applications, including antimicrobial, anticancer, antidiabetic, and anti-tuberculosis.

Antidiabetic and anti-tuberculosis are chronic diseases and are very dangerous. Tuberculosis is a communicable disease and is a major cause of ill health by bacillus mycobacterium grows tuberculosis in people when sick. Due to cough, the virus spreads through the air and affects the lungs (pulmonary). Most people suffer from the hazards of TB and related diabetes. Due to their high sensitivity, the metal complexes exhibited electrochemical properties due to redox behavior [7-9]. Thus, the modified glassy carbon electrode prepared using a metal complex can determine the nitrite [10]. Nitrate is an essential trace element for animals and plants. It is produced in the environment by the nitrogen cycle, fertilizer, and decomposition of animal waste. It refers to $-ONO$ group and is also called vasodilator medication. Medicine uses it to treat angina, cure ingredients, and preserve food. Excess nitrate causes harmful effects on the ecological system, water bodies, and especially on children (methemoglobinemia / blue baby syndrome).

Moreover, Befoul nitrate arising from muck has hazardous effects on the human living system [11-13]. Thus, the determination of nitrate is very important. Literature showed that cyclic voltammogram is a potential, sensitive, rapid, and reliable method for the detection of NO_2 . This method is used to investigate the electrochemical feature of a substance in solution.

Our research group has synthesized N, S heterocycles, and its metal complexes and evaluated their anticancer activity and electrochemical studies. In continuation of our research work, we reported here the synthesis of a novel [4-((E)-(2-hydroxy-3-methoxy-5-((E)-thiazol-2-ylidiazanyl)benzylidene)amino)-1,5-dimethyl-2-phenyl-1H-pyrazol-3(2H)one], and its Cu(II), Co(II), Ni(II) complexes followed by evaluation of their biological activities as well as the electrochemical determination NO_2 have been carried out [13].

2. Materials and Methods

2.1. Source and instrumentations.

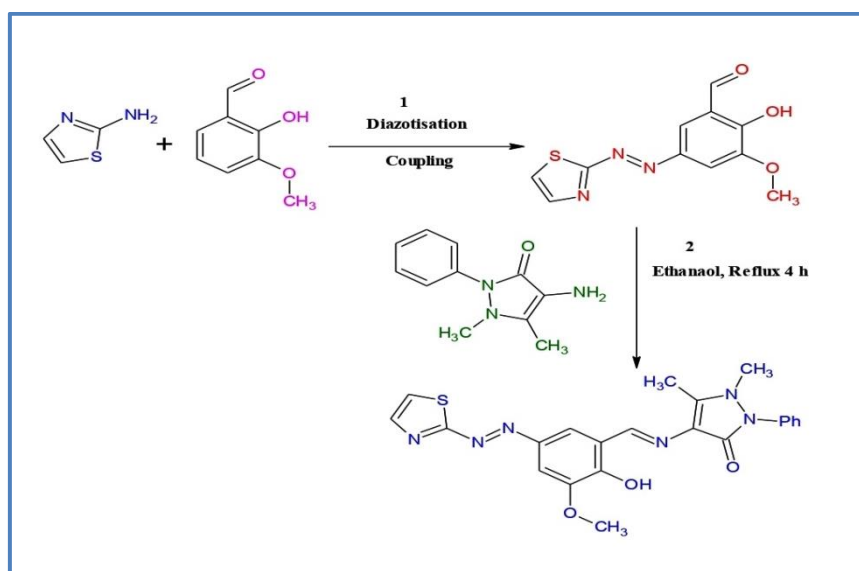
All the chemicals used were of analytical grade and purchased from Avra, Himedia, and Merck companies which were used without further purification. The melting point is determined by the digital melting point apparatus, electrothermal IA9100. The synthesized compounds were characterized by UV-Vis spectrophotometer in the range 200-800 nm using Systronics 119 model in DMF solvent. The elemental analysis was performed on a CHN analyzer Carlo Erba 1108 analyzer. The IR spectra were recorded using a KBr pellet in a Shimadzu FT-IR spectrophotometer in the range $4000-400\text{ cm}^{-1}$. 1H NMR spectrum of the ligand was determined by BRUKER Ascend 400MHz FTNMR spectrometer using tetramethylsilane as an internal reference standard. The electrochemical analysis was done by CHI 620E analyzer Inc. made in the U.S.A. The powder XRD was recorded on powder x-ray diffraction facility code XRD POWDER100. Mass spectra of the compounds were determined on LC-MS water AQUITY-2777C mass spectrometer. The molar conductance measurement was conducted using an ELICO-CM82 conductivity meter. The magnetic moment of the complexes was noted at $28\text{ }^\circ\text{C}$ by Gouy balance version 7550 using $Hg[Co(NCS)_4]$ as a

calibrant. The TGA of complexes was carried out on SII Exstar TG/DTA 6300 instrument from the laboratory temperature to 1000 °C with a scan rate of 10 °C/ min.

2.2. Synthesis of azo dye ligand (HTDP).

The ligand HTDP has been synthesized in two steps. The starting material was prepared using a diazonium salt solution and 2-hydroxy-3-methoxy benzaldehyde in the first step. The solution of diazonium salt was prepared by dissolving 3 mM/0.3g of 1,3-thiazole-2-amine in 10 mL of distilled water along with 5 mL of concentrated hydrochloric acid, which was maintained at 0-5 °C for 1 h and 4 mL of 3 mM/0.2g NaNO₂ solution was added dropwise in amine solution with continuous stirring. The obtained diazonium salt solution was added to a cold solution of 2-hydroxy-3-methoxy benzaldehyde (3 mM/0.46g) in basic media. The resulting solution was stirred for 1 h at the same temperature by maintaining pH 6-7. After completion of the reaction, the reaction solution was poured into ice-cold water. The solid product obtained was (2-hydroxy-3-methoxy-5-[(Z)-1,3-thiazol-2-ylidiazonyl]benzaldehyde.

In the second step, (2-hydroxy-3-methoxy-5-[(Z)-1,3-thiazol-2-ylidiazonyl]benzaldehyde (2 mM/0.6g) and 4-amino antipyrine (2 mM/0.5g) dissolved in 20 mL methanol. Acetic acid was added to the above solution's catalytic amount of (4 drops). The above mixture was stirred under reflux for 4 h. The progress of the reaction was monitored by TLC (ethyl acetate: petroleum ether, 4:6). After completion of the reaction; the solution was poured into ice-cold water to get [4-((E)-(2-hydroxy-3-methoxy-5-((E)-thiazol-2-ylidiazonyl)benzylidene)amino)-1,5-dimethyl-2-phenyl-1H-pyrazol-3(2H)one] (HTDP). The precipitate was filtered, washed with ethanol, and recrystallized from the methanol [13]. The synthetic route is presented in Scheme 1.

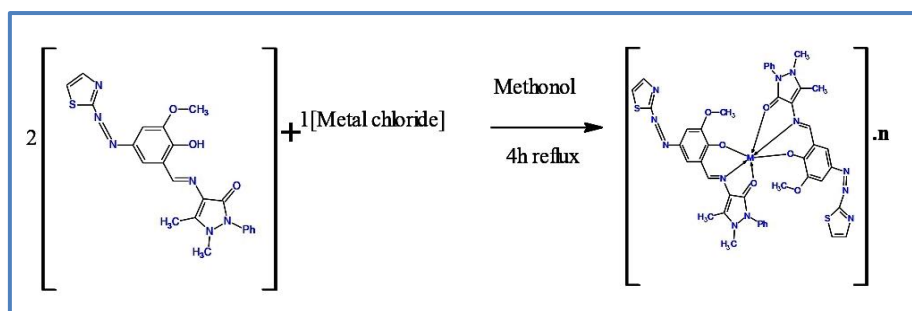


Scheme 1. [4-((E)-(2-hydroxy-3-methoxy-5-((E)-thiazol-2-ylidiazonyl)benzylidene)amino)-1,5-dimethyl-2-phenyl-1H-pyrazol-3(2H)one] (HTDP).

2.2.1. Synthesis of metal complexes.

All the complexes were prepared in the stoichiometric ratio 1:2 of metal: ligand. A solution of copper chloride was prepared in 20 mL methanol (1mM/0.15g) and added to the hot methanol solution of HTDP (2 mM/1g) with continuous stirring. The resulting solution was refluxed for 4 h in the temperature range of 50-60 °C. The solid precipitate was collected by filtration, washed several times with ethanol, rinsed with ethanol, then kept in a desiccator [13,

14]. A similar method was followed for preparing Co(II) and Ni(II) complexes by taking the solution of a respective metal chloride, as represented in Scheme 2. Yield is 75–80%.



Scheme 2. Preparation of [Cu(HTDP)₂]₂H₂O, [Co(HTDP)₂] and [Ni(HTDP)₂]₂H₂O.

2.3. Electrochemistry instrumentation.

Nitrate detection has been performed by cyclic voltammetry. It consists of the 3-electrode system; the working electrode is a glassy carbon electrode (GCE), platinum as a counter electrode (PCE), and Ag/AgCl electrode as a reference electrode in the redox system. The GC electrode was wiped with alumina powder and washed properly with deionized water. Subsequently, 10 μ L of [Co(HTDP)₂] in DMF drop coated on the electrode surface area. This was used to detect nitrate in phosphate buffer (pH 7) solution [15, 16].

2.4. Biological evaluation.

2.4.1. Antimicrobial activities.

The antimicrobial activity of HTDP and their metal complexes screened against bacterial cultures of gram +ve *S. aureus* (MTCC-7443), gram –ve *E. coli* (MTCC-7410), *Shigella* (MTCC-1457) and *clostridium* (MTCC-11078). The fungal Cultures like *Aspergillus niger* (MTCC-1207) and *Fusarium* (MTCC-1372) were used for dual-culture agar well diffusion assay along with some modifications. A Petri plate was prepared using 20 mL of sterilized nutrient agar for bacterial, and potato dextrose agar media for fungal cultures were adjusted under aseptic conditions. The 100 μ L of standardized test microbial was transferred by using an inoculum glass loop. Screening in various concentrations and 1 mg/mL were tested. Further, diluting the sample with DMSO was added to plates; after that, the plates were incubated under 37 °C for one day. Chloramphenicol was considered standard for bacterial assay, and nystatin was used for the antifungal assay. The diameter of the inhibition zone was measured in millimeters (mm), and three repeated trials were taken to assess the strength of antibacterial activity [17, 18].

2.4.2. Antidiabetic study.

Inhibition of alpha-amylase enzyme was carried out according to the standard methods with a small modification. In a test tube, the reaction mixture containing 500 μ L phosphate buffer (100 mM, pH 6.8), 100 μ L α -amylase (2 U/mL), and 200 μ L of extract of different concentrations of HTDP and its metal complexes from 25 to 100 μ g was pre-incubated at 37 °C for 20 min. Then, the 200 μ L of 1% soluble starch (100 mM phosphate buffer pH 6.8) was added as a substrate and incubated further at 37 °C for 30 min. 1 mL of the DNS color reagent was added and kept in a water bath for 10 min. The absorbance of the resulting mixture was

measured at 540 nm using a UV-Visible spectrophotometer. Acarbose (0.1–0.5 mg/mL) was used as a standard, and the experiment was executed in three replicates.

The percentage of Alpha-amylase inhibition was calculated using the equation is

$$= [AC - \frac{AE}{AC}] \times 100$$

AC: absorbance of control and AE: absorbance of extract [19, 20].

The dose efficiency IC₅₀ value was calculated via standard Excel linear regression method by the average value of the zone inhibition result in various concentrations.

2.4.3. Anti-tuberculosis activity.

Mycobacterium tuberculosis MTCC 300, the culture was developed, and sub-cultured functioning Middle brook 7H10 [21, 22] with some of the modification was prepared well-mannered in the nutrient broth at 37 °C for 48 h for Anti-tuberculosis assay. The growth was noted down spectrophotometrically at 600 nm. The MIC assay was detected using a tetrazolium salt indicator, MTT (3-4,5-dimethylthiazol-2-yl-2,5-diphenyl tetrazolium bromide). The MIC assay took an aliquot of 25, 50, and 100 µL of 0.1 g/mL of the ligand and its metal complexes. MTT was pipetted into each well (40 µL at 0.2 mg/mL), and once growth control wells revealed a purple color following incubation at 37 °C for 1 h viable by the agent. The color formation indicated growth inhibition in the assay. An average of three repeated tests was taken to assess the strength of the activity. Chloramphenicol was considered a standard [23, 24]. The minimum inhibitory concentration IC₅₀ value was evaluated by excels linear regression technique by the mean value of the activity in different concentrations region.

3. Result and Discussion

3.1. Characterization.

The reaction of an azo dye, HTDP, with metal chlorides produces the complexes of the formula [Cu(HTDP)₂]H₂O, [Co(HTDP)₂] and Ni(HTDP)₂·2H₂O. The synthesized ligand and its complexes were characterized by various spectroscopic techniques. The prepared complexes are colored, crystalline solid, and stable toward air and moisture at the laboratory temperature. The complexes are insoluble in a common organic solvent but soluble in DMF and DMSO solvents. The molar conductance measurements of the complexes used in DMF (1×10⁻³ M) solution reported 18-29 Ω⁻¹ cm² mol⁻¹ indicating non-electrolytic nature [25]. The experimental, analytical data of HTDP and their metal complexes are reported in Table 1.

Table 1. Physical and analytical data of HTDP and its complexes.

Compounds	Color	Mol. wt	M.P in °C	Elemental analysis in (%) Cal				Λm Ω ⁻¹ cm ² mol ⁻¹
				C	H	N	M	
HTDP	Reddish brown	449.25	≥ 166	59.9 (58.8)	4.46 (4.45)	18.7 (18.7)		
[Cu(HTDP) ₂]H ₂ O	Brick pink	976.5	≥ 177	54 (54)	4.9 (4.7)	17.2 (17.1)	6.4 (6.3)	24
[Co(HTDP) ₂]	Bluish brown	953.9	≥ 204	55.4 (55.2)	3.9 (3.82)	17.6 (17.5)	5.6 (5.5)	29
[Ni(HTDP) ₂]·2H ₂ O	Dark red	989.3	≥ 179	53.3 (53.13)	4.2 (4.14)	16.9 (16.4)	5.8 (5.79)	19

3.2. ¹H NMR spectral data.

The newly synthesized HTDP has been characterized by ¹H NMR in DMSO-d₆ solvent at room temperature, and the spectrum is represented in Figure 1. The Ar-OH group proton

signal appeared as a singlet at 14.25 ppm, and the proton of azomethine group resonance appeared at 9.80 ppm (s, 1H, CH=N). The signals from aromatic protons appeared in the region 8.09-7.39 ppm (m, 9H, Ar-H), and methyl protons of the 4-aminoantipyrine ring (s, 3H, C-CH₃) showed around 2.50 ppm. Another methyl group signal of N-CH₃ proton exhibited a singlet at 3.36 ppm. The methoxy group protons showed a singlet on 3.94 ppm (s, 3H, Ar-OCH₃) [25, 26].

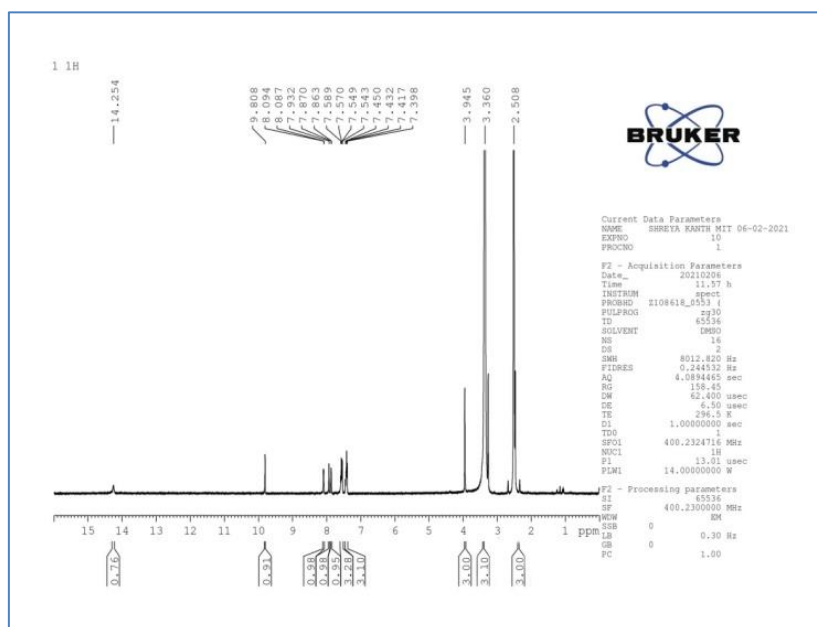


Figure 1. ¹H NMR spectrum of HTDP3.3. *Infrared spectra studies.*

The IR spectra of HTDP and their coordinated complexes are shown in Supplementary Figure S1, and the data are compiled in Table 2. The peak that appeared at 3002 cm⁻¹ is due to C-H stretching, and the ν(C=N) of the azomethine group exhibited a signal at 1553 cm⁻¹. The ν(-C=O) stretching was obtained at 1601 cm⁻¹. The peaks of azomethine and carbonyl groups shifted to 30-40 cm⁻¹ in their complexes due to the formation of ligand–metal coordination bonds. A band due to ν(OH) of ligand appeared at 3393 cm⁻¹, and this band is absent in the spectra of their complexes, indicating the elimination of hydrogen of the OH group; hence and it justified that the OH group is also involved in the coordination bond formation. Further, the complex formation is confirmed by the appearance bands around 480 cm⁻¹ for ν(M-N), and ν(M-O) obtained at 580-690 cm⁻¹. The band due to ν(N=N) appeared at 1441 cm⁻¹ in the spectrum of the uncoordinated ligand, and it appeared at the same position in their metal complexes, indicating the non-involvement of the N=N group in the coordination. Thus spectral results confirmed the tridentate nature of the ligand [27-29].

Table 2. FT-IR spectral data (cm⁻¹) of ligand and its metal complexes.

Compound	ν(-OH)	ν(Ar-CH)	ν(C=O)	ν(C=N)	(N=N)	ν(M-N)	ν(M-O)
HTDP	3393	3002	1601	1533	1441	-	-
[Cu(HTDP) ₂].H ₂ O	-	3000	1582	1549	1447	457	581
[Co(HTDP) ₂]	-	3001	1565	1519	1442	489	689
[Ni(HTDP) ₂].2H ₂ O	-	3002	1586	1551	1443	486	673

3.4. UV-Visible studies.

The Uv-visible absorption bands of synthesized HTDP and their coordinated metal complexes were noted in the DMF solution. The free HTDP exhibited an absorption band at

22,727 cm⁻¹, which is ascribed to n→π* of the (-N=N-) group. Further, the absorption band due to the (-N=N-) group in all the metal complexes is shifted to a higher wavelength indicating coordination of the metal chlorides with the HTDP. The electronic absorption spectrum of [Cu(HTDP)₂]H₂O showed two bands at 24,570, and 18518 cm⁻¹, which are ascribed to ²B_{1g}(F)→²B_{1g} and ²B_{1g}(F) →²B_{2g} respectively transitions suggesting octahedral geometry, and this is emphasized by its magnetic moment value of 1.76 BM [25, 26]. The [Co(HTDP)₂] exhibited two transition at 24,390 and 18,518 cm⁻¹ which are to ⁴T_{1g}(F)→⁴T_{2g}(P) (ν₂) and ⁴T_{1g}(F) →⁴T_{2g}(F) (ν₁) respectively. The magnetic moment value of 4.3BM and transitions are represented as octahedral geometry [25-30]. Similarly, in the case of [Ni(HTDP)₂]2H₂O a transition at 22,573 cm⁻¹ is assigned to the ³A_{2g}(F)→³T_{1g}(F) and another transition at 19,120 cm⁻¹ is assigned to A_{2g}(F)→³T_{1g}(F) [31, 32]. The observed electronic transitions and the magnetic moment value of 3.2BM suggest an octahedral geometry for the complex. The electronic transition data is represented in Table 3, and the transition shows in Supplementary Figure S2.

Table 3. Electronic spectral data for ligand and its complexes.

Compounds	Absorptions in cm ⁻¹	Transitions assignment	Geometry μ _{eff} (BM)
HTDP	24,691	n→π*	
[Cu(HTDP) ₂]H ₂ O	24,570 18518	² B _{1g} (F)→ ² B _{1g} ² B _{1g} (F) → ² B _{2g}	1.76BM
[Co(HTDP) ₂]	24,390 18,518	⁴ T _{1g} → ⁴ T _{2g} (P) ⁴ T _{1g} → ⁴ T _{2g} (F)	4.3BM
[Ni(HTDP) ₂]2H ₂ O	22,573 19,120	³ A _{2g} (F)→ ³ T _{1g} (F) A _{2g} (F)→ ³ T _{1g} (F)	3.2BM

3.5. Mass spectral studies.

The molecular ion peak of HTDP appeared at m/z 449 (M+1). The value matches the expected mass. The molecular peak (M+1) observed in the mass spectra of [Cu(HTDP)₂]H₂O, [Co(HTDP)₂], and [Ni(HTDP)₂]2H₂O are at 977.5, 954.5, and 990.3 respectively as shown in Figure 2.

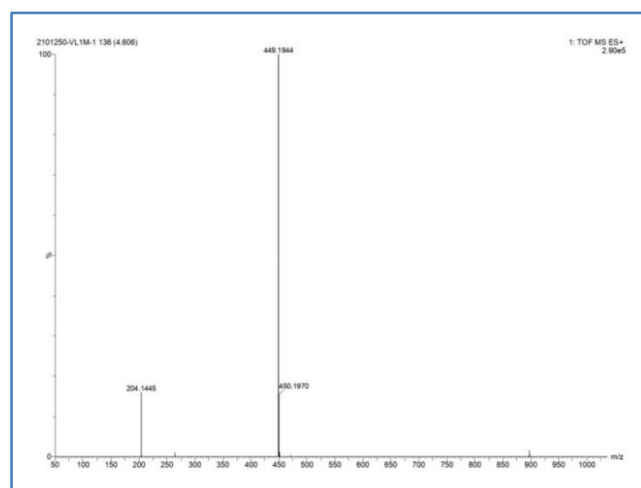


Figure 2. Mass spectrum of HTDP.

3.6. Powder XRD spectral studies.

The synthesized [Cu(HTDP)₂]H₂O, [Co(HTDP)₂], and [Ni(HTDP)₂]2H₂O complexes were studied, and the degree of crystallinity was determined from the powder X-ray diffraction patterns. The powder X-ray diffraction pattern of [Cu(HTDP)₂]H₂O showed 6 reflections in the range of 9-30°. The [Co(HTDP)₂] showed 5 peaks in the region 9.2-27°, and 9 peaks

appeared for [Ni(HTDP)₂]₂H₂O in the range at 6-28°, and their spectra are represented in Figure 3. The inter-planer spacing (d) has been calculated from the positions of intense peaks (2θ) using Bragg's equation $n\lambda=2d\sin\theta$ (where $\lambda= 1.5406 \text{ \AA}$). Further, the unit cell parameter was calculated for cubic symmetry from the obtained peaks, and $h^2 + k^2 + l^2$ values were determined and tabulated in Table 4. The calculated lattice parameter $a=b=c$ of [Cu(HTDP)₂]₂H₂O is 9.35, [Co(HTDP)₂] is 9.28, while for [Ni(HTDP)₂]₂H₂O is found to be 13.2. The presence of forbidden numbers 7 and 15 in the observed results indicates that the [Ni(HTDP)₂]₂H₂O complex may belong to hexagonal or tetragonal systems [28-34].

Table 4. XRD spectral data.

Compound	Point	2θ	Sinθ	Sin ² θ	Sin ² θ×1000	h ² +k ² +l ²	h k l	D		a in Å
								Obs	Cal	
[Cu(HTDP) ₂] ₂ H ₂ O	1	9.21	0.080	0.0064	6.4	1(1)	1 0 0	9.5	9.5	9.6
	2	9.5	0.082	0.0067	6.7	1.04(1)	1 0 0	9.2	9.2	9.5
	3	14.8	0.128	0.0160	16	2.5(3)	1 1 1	5.9	5.5	9.6
	4	19.3	0.167	0.0270	27	4.21(4)	2 0 0	4.5	4.9	9.9
	5	23.12	0.199	0.0390	39	6.09(6)	2 1 1	3.7	3.8	9.4
	6	26.1	0.220	0.0480	48	7.5(8)	2 2 0	3.4	3.2	9.6
[Co(HTDP) ₂]	1	9.28	0.080	0.0064	6.4	1(1)	1 0 0	9.5	9.4	9.4
	2	14.86	0.120	0.0144	14.4	2.11(2)	1 1 0	6.1	6.5	9.1
	3	17.2	0.140	0.0190	19	2.91(3)	1 1 1	5.1	5.4	9.3
	4	19.4	0.165	0.0272	27.2	4.27(4)	2 0 0	4.5	4.7	9.4
	5	26.5	0.228	0.0519	51.9	8.10(8)	2 2 0	3.3	3.2	9.2
[Ni(HTDP) ₂] ₂ H ₂ O	1	6.4	0.060	0.0036	3.6	1(1)	1 0 0	13	12.9	13
	2	9.2	0.080	0.0064	6.4	1.77(2)	1 1 0	9.2	9.5	13.7
	3	10.2	0.088	0.0077	7.7	2.13(2)	1 1 0	8.9	8.5	13
	4	14.8	0.128	0.0163	16.3	4.58(5)	2 1 0	5.9	5.9	13.6
	5	16.12	0.139	0.0193	19.3	5.3(5)	2 1 0	5.8	5.4	13
	6	17.2	0.149	0.0222	22.2	6.1(6)	2 1 1	5.4	5.2	13.3
	7	18.3	0.159	0.0252	25.2	7(7)	-	4.9	4.8	13.1
	8	19.4	0.168	0.0282	28.2	7.8(8)	2 2 0	4.6	4.5	13.1
	9	26.6	0.230	0.0529	52.9	14.6(15)	-	3.3	3.3	13

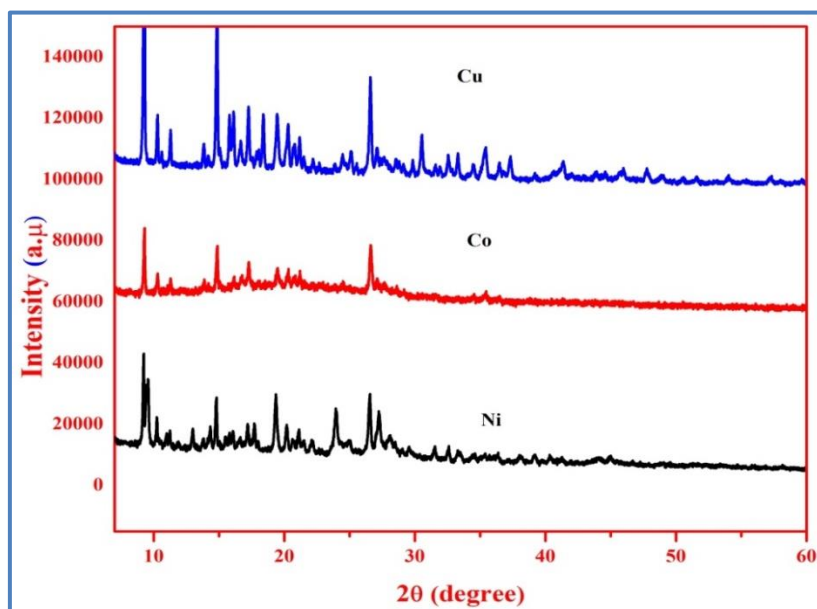


Figure 3. Powder XRD patterns of Cu, Co, and Ni complexes.

3.7. Thermal analysis of the compound.

The thermal analysis of [Cu(HTDP)₂]₂H₂O, [Co(HTDP)₂] and [Ni(HTDP)₂]₂H₂O were carried out to understand the thermal stability and the weight loss of the complexes from the laboratory temperature to 1000 °C at a heating rate of 10 °C/min in an N₂ atmosphere. The

temperatures versus weight loss percentage are plotted and shown in Figure 4. The [Cu(HTDP)₂]₂H₂O indicates three degradation steps. In the first step, a weight loss of 8.14 % occurred in the region 43-177 °C due to the loss of one H₂O and 4CH₃ [35]. In the second stage, the weight loss of 35.8% appeared in the temperature range 177-476 °C corresponding to loss of C₁₄H₁₀N₆S₂O moiety, and in the third stage, 50 % weight loss occurred from 476-667 °C due to the dissociation of C₂₆H₂₀N₆O₄ moiety. Finally, CuO remains.

In the case of [Co(HTDP)₂], the weight loss that occurred initially, amounting to 20.9 %, has appeared at 65-286 °C due to the dissociation of C₆N₄S₂. The second stage of weight loss, 36.5 %, occurred in the temperature region 286-481 °C because of the dissociation of C₁₇H₁₆N₄O₄ moiety. The third stage attributed to eliminating 36.2 % of C₂₁H₂₁N₄O moiety in the range 481-594 °C, leaving behind CoO as residue [36].

In the [Ni(HTDP)₂]₂H₂O, a weight loss in the region 42-179 °C of about 9.4 % corresponds to two lattice water molecules. In the second stage, 42.2 % weight loss due to the loss of C₂₀H₁₄N₆S₂O in the region at 179-476 °C. Further, the third stage of weight loss from 476 to 601 °C due to 41 % of C₂₀H₁₅N₆O₄ moiety. Finally, NiO residue remained [37, 38]. The weight loss data are given in Table 5, and degradation curves are shown in Figure 4.

Table 5. Thermal degradation.

Complex	Step	Decomposition Temp °C	Assignment	Loss of mass in (%)	Residue species
[Cu(HTDP) ₂] ₂ H ₂ O	1	43-177	H ₂ O & 4CH ₃	8.14 %	CuO
	2	177-476	C ₁₄ H ₁₀ N ₆ S ₂ O	35.8%	
	3	476-667	C ₂₆ H ₂₀ N ₆ O ₄	50.1%	
[Co(HTDP) ₂]	1	65-286	C ₆ N ₄ S ₂	20.9%	CoO
	2	286-481	C ₁₇ H ₁₆ N ₄ O ₄	36.5%	
	3	481-594	C ₂₁ H ₂₁ N ₄ O	36.2%	
[Ni(HTDP) ₂] ₂ H ₂ O	1	42-179	2H ₂ O & 4CH ₃	9.5%	NiO
	2	179-476	C ₂₀ H ₁₄ N ₆ S ₂ O	42.2%	
	3	476-601	C ₂₀ H ₁₅ N ₆ O ₄	41 %	

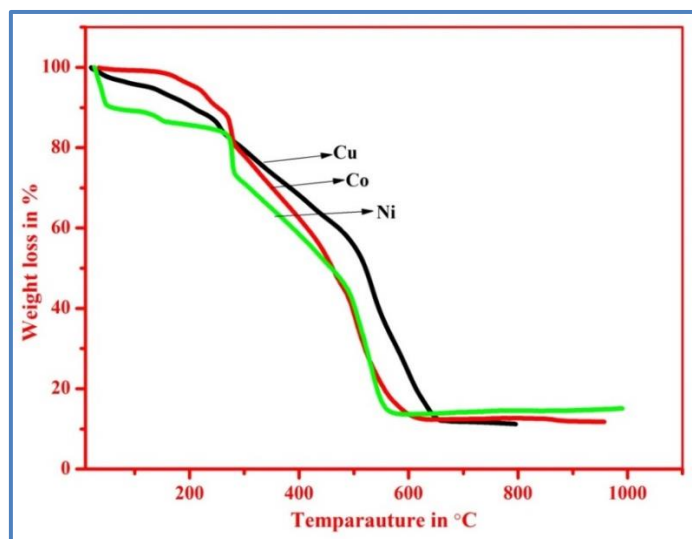


Figure 4. TGA curves for [Cu(HTDP)₂]₂H₂O, [Co(HTDP)₂] and [Ni(HTDP)₂]₂H₂O.

3.8. Electrochemical methods.

3.8.1. Charge transfer behavior of modified electrode.

The charge transfer behavior of [Co(HTDP)₂] modified sensor was investigated using K₄[Fe(CN)₆], which behaves as a redox system. The bare glassy carbon electrode (GCE)

displayed Ferri/ferrocyanide system in CV response. The electrochemical evaluation in 0.5 mM $K_4[Fe(CN)_6]$ solution at a scan rate of 50 mVs^{-1} between bare GCE and modified $[Co(HTDP)_2]/GC$ electrodes is represented in Figure 5. The modified GCE exhibited more current response in the redox system because the cobalt complex has good electro-conducting properties compared to bare GCE. The electrode acts as a bridge of electron transfer between electrode surface (GCE) and buffer solution in cyclic voltmeter [15, 39].

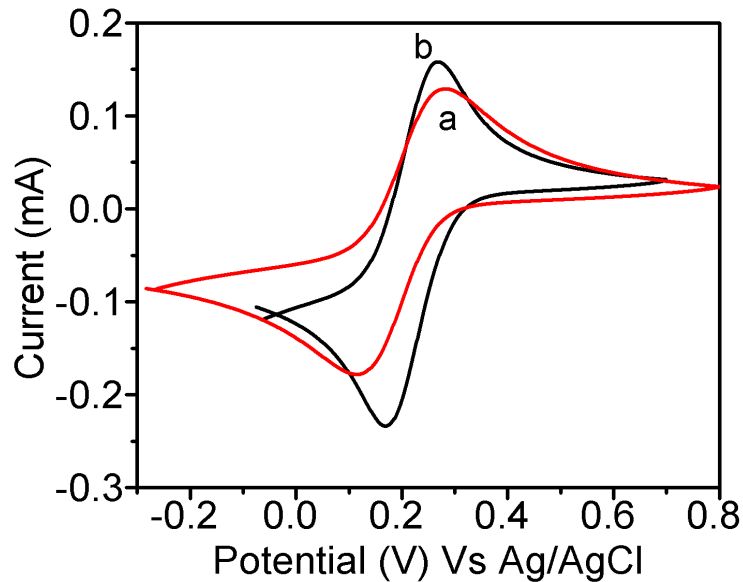


Figure 5. Cyclic voltammogram of 0.5M KCl solution containing 5 mM $K_3[Fe(CN)_6]/K_4[Fe(CN)_6]$ at the a) bare GCE and b) $[Co(HTDP)_2]/GCE$, the scan rate: 50 mVs^{-1} .

3.8.2. Electrochemical investigation of the different electrodes.

Electrochemical sensing of nitrate was studied in an electrolytic solution using a phosphate buffer solution of pH 7. The cyclic voltammogram of (i) bare GCE, (ii) $[Co(HTDP)_2]/GCE$, and (iii) $[Co(HTDP)_2]/GCE$ with $02\text{ }\mu\text{L}$ concentration of $NaNO_2$ at a scan rate of 50 mVs^{-1} . The modified $[Co(HTDP)_2]/GC$ electrode showed intensified peak current responses as compared to the bare glassy carbon electrode, and it is a good electrochemically active, as represented in Figure 6 [16, 39].

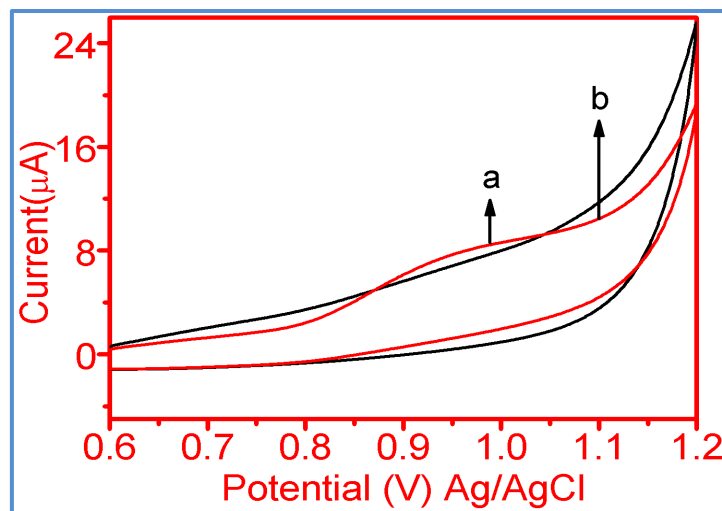


Figure 6. Cyclic voltammogram of different electrodes (a) Bare GCE and (b) Modified electrode in PBS pH 7 with the scan rate of 50 mVs^{-1} .

3.8.3. Electrochemical determination of nitrate.

Electrochemical detection of nitrate using modified [Co(HTDP)₂]/GC electrode by CV technique. In the modified electrode in PBS pH 7.0, the CV curves showed an increase in peak current response compared to bare GCE. While adding the various concentrations of nitrate, the peak current increases gradually with a slight shift in the potential towards the negative side; therefore, the modified electrode can detect the nitrate with a long linear range, lower sensitivity, and LOD. The linear range of (10 to 80 μmolL⁻¹), sensitivity is (0.172 μAμM⁻¹cm⁻²), and LOD is 3.333 μML⁻¹. Thus, the modified electrode exhibits highly electroactive properties, as shown in Figure 7 [15, 39].

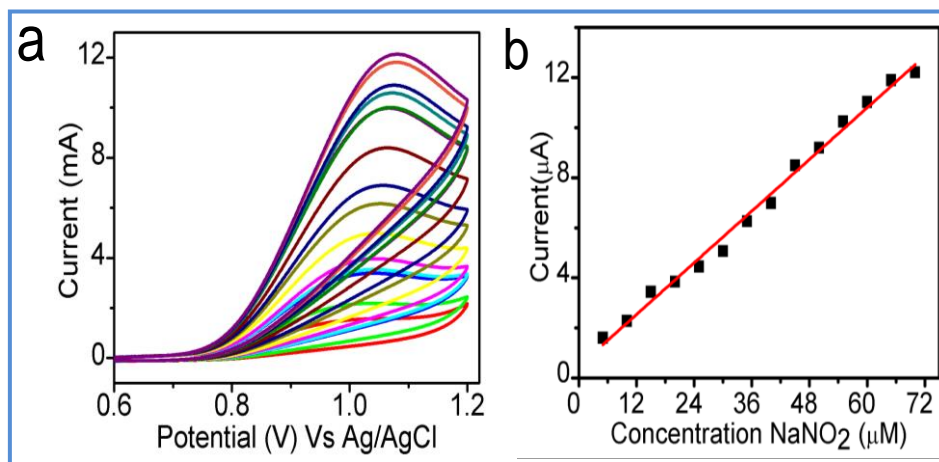


Figure 7. (a) Cyclic voltammogram of nitrite at [Co(HTDP)₂]/GCE concentration ranging from (10–80 μM); (b) Inset: Calibration plot of current vs. concentration of NaNO₂.

3.8.4. Effect of scan rate.

The modified GCE while subjected to a pH 7 buffer solution containing 10 μM of nitrate under various scan rates from 10 to 150 mVs⁻¹. As the scan rate increases, the peak current increases linearly as calculated by the linear regression equation, $I_{pa} (\mu A) = 0.918 (\mu M) + 0.0609$ and $R^2 = 0.9959$. These analytical results confirm [Co(HTDP)₂] that the modified electrode was a surface adsorbed process, as shown in Figure 8 [40].

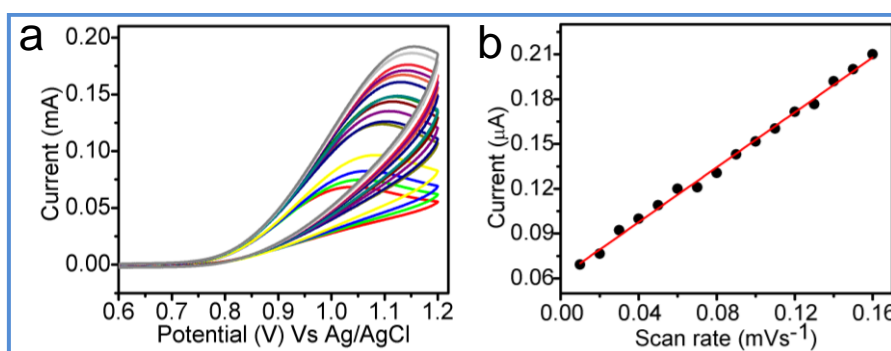


Figure 8. (a) Cyclic voltammogram of nitrite at [Co(HTDP)₂] modified GCE scan rate ranging from (10 - 150mVs⁻¹). (b) Inset plot of current (I) vs. Scan rate.

3.8.5. Differential pulse voltammetry (DPV) studies of nitrate.

DPV was one of the best techniques for detecting NaNO₂ using modified [Co(HTDP)₂]/GCE, and it gives accurate results for the detection of nitrate; it was conducted in PBS pH 7, and the results are collected in Table 6. The reaction which occurs, as mentioned

below, showed enhanced anodic peak current response compared to bare GCE. The increasing concentration of NaNO₂ in the range from 0.1 to 01 μmolL⁻¹ and the peak current enhanced at the fixed potential. The analytical profile of the modified electrode was long linear range 0.1-01 μmolL⁻¹, sensitivity 11.043 μAμM⁻¹cm⁻² and LOD is 0.0333μML⁻¹. Further, the modified sensor shows reproducible and anti-leaching properties, as shown in Figure 9; the electrochemical behavior of the electron is shown in the below equations [41].

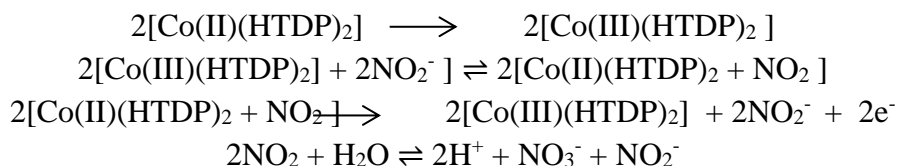


Table 6. Comparison of the different actuators for nitrite determination.

Compound	Linear range μmolL ⁻¹	Technique	LOD μML ⁻¹	Sensitivity (μA μM ⁻¹ cm ⁻²)	Reference
CoTM-QOPc /CNP/GCE	0.2-200	CV	0.06	2.298	15
	0.2-225	DPV	0.06	1.031	
CoL/MNSs/CPE	0.2-30.0	SWV	0.015	-	39
Co ₃ O ₄ /RGO	1-380	CV	0.14	29.5	40
CRS/BN-RGO/GCE	0.001- 1290	DPV	0.0159	1.5309	41
[Co(HTDP) ₂]/GCE	10-80	CV	3.333	0.172	This work
	0.1-1	DPV	0.0333	11.043	

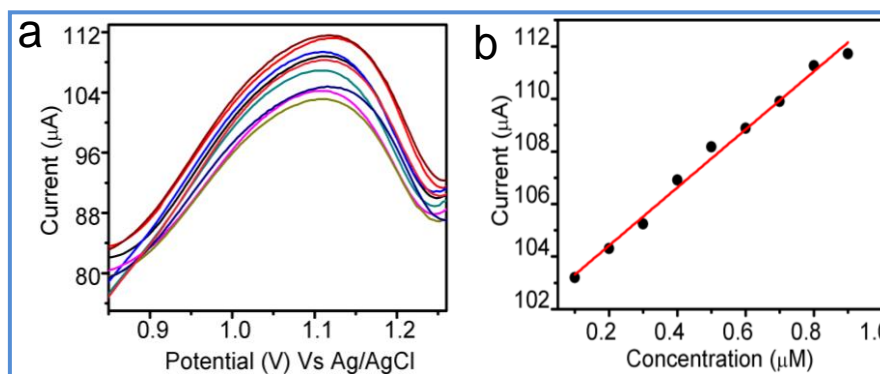


Figure 9. (a) DPV of [Co(HTDP)₂]/GCE at a concentration ranging from 0.1– 10 μM/L. (b) Inset: Calibration graph of current vs. concentration.

3.8.6. Real samples analysis.

The experimental feasibility of the [Co(HTDP)₂]/GCE was applied to examine the nitrite range in different milk sources using a standard addition method. The results were noted in Table 7. Initially, each individual milk source was filtered with 0.1 M of pH 7 PBS as described in the Chinese National Food Safety Standard [42] and appeared in 92–103% range. These results reveal the experimental feasibility of the drop-coated electrode for analysis of nitrite in real samples.

Table 7. Determination of nitrite in milk samples.

Sample	Added (μmolL ⁻¹)	Detected (μmolL ⁻¹)	Recovery (%)
Milk 1	5	4.60	92
	10	10.06	100.6
	15	14.84	98.93
Milk 2	5	4.45	89
	10	10.33	103.3
	15	14.94	99.6

3.9. Biological studies.

3.9.1. Antimicrobial activities.

The HTDP and their synthesized metal complexes were tested for antimicrobial cultures by agar well diffusion technique using Nystatin and Chloramphenicol as standard drugs. The zone of inhibition area in activities was measured by millimeters (mm) and was repeated three times to assess the effectiveness of the antimicrobial activity. Ligand showed a good positive result, and its complexes showed enhanced growth inhibition against an antimicrobial activity as compared to the uncoordinated ligand. Because metals were bonded with donating atoms of the ligand having delocalized electrons in the chelating rings system, the lipophilic property increases [43, 44]. The result was explained through Tweedy’s chelation theory and Overtone’s theory [45-47]. Moreover, the $[Cu(HTDP)_2]H_2O$ and $[Co(HTDP)_2]$ showed the best result with *S.aureus*, *E.coli*, and *fusarium* microbial cultures. The values are recorded in Tables (8 and 9), and the results are represented in Figures 10 to 12.



Figure 10. Antibacterial zone of inhibition by HTDP and its metal complexes.

Table 8. Antibacterial activity of the HTDP and its metal complexes.

Bacterial cultures	Cont. in mg/ml	Growth inhibition in mm				STD
		HTDP	$[Cu(HTDP)_2]H_2O$	$[Co(HTDP)_2]$	$[Ni(HTDP)_2]2H_2O$	
<i>S.aureus</i>	0.25	2.03 ± 0.05	8.03 ± 0.05	6.03 ± 0.05	2.03 ± 0.05	16 ± 0
	0.5	2.2 ± 0.2	9.03 ± 0.05	10.03 ± 0.05	8.03 ± 0.05	16 ± 0
	0.75	2.3 ± 0.35	09.2 ± 0.25	11.06 ± 0.1	8.3 ± 0.6	16 ± 0
	1	2.4 ± 0.3	10.03 ± 0.05	14 ± 0	9.03 ± 0.05	16 ± 0
<i>Clostridium</i>	0.25	3 ± 0	8.03 ± 0.05	7.03 ± 0.05	5.03 ± 0.05	16 ± 0
	0.5	4.06 ± 0.1	10.03 ± 0.05	12.03 ± 0.05	8.03 ± 0.05	16 ± 0
	0.75	4.13 ± 0.15	10.16 ± 0.28	11.16 ± 0.28	8.8 ± 0.07	16 ± 0
	1	4.2 ± 0.25	12.06 ± 0.11	13.03 ± 0.05	9.03 ± 0.05	16 ± 0
<i>E. coli</i>	0.25	3.1 ± 0.1	9.03 ± 0.05	5.03 ± 0.05	5.03 ± 0.05	18 ± 0
	0.5	4.1 ± 0.1	9.3 ± 0.05	11.03 ± 0.05	7.03 ± 0.05	18 ± 0
	0.75	4.2 ± 0.15	12.03 ± 0.05	13.03 ± 0.05	9.03 ± 0.05	18 ± 0
	1	4.2 ± 0.15	12.03 ± 0.05	13.03 ± 0.05	9.03 ± 0.05	18 ± 0
<i>Shigella</i>	0.25	2.2 ± 0.34	8.03 ± 0.05	9.03 ± 0.05	5.1 ± 0.17	16 ± 0
	0.5	4.1 ± 0.1	8.16 ± 0.15	9.16 ± 0.28	8.06 ± 0.1	16 ± 0
	0.75	4.13 ± 0.1	9.16 ± 0.15	11.26 ± 0.4	9.1 ± 0.3	16 ± 0
	1	4.16 ± 0.15	10.03 ± 0.05	12.03 ± 0.05	9.16 ± 0.2	16 ± 0

*Each value is displayed average ± SD (standard deviation) of three replicates for the zone of inhibition.

*STD: Chloramphenicol.

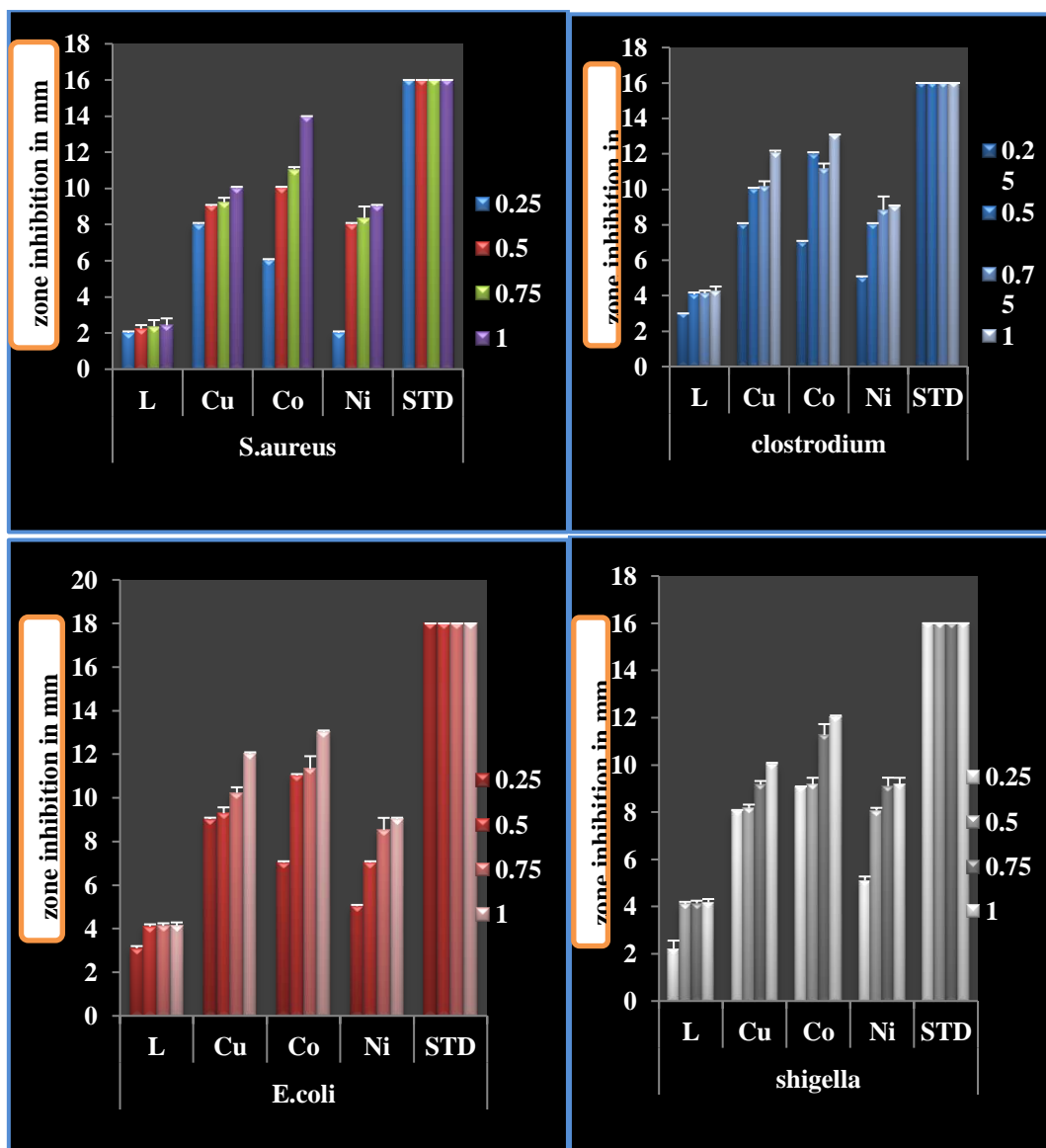


Figure 11. Graphical representation of antibacterial zone of inhibition activity of HTDP and its metal complexes.

Table 9. Antifungal activity of HTDP and its metal complexes.

Fungal cultures	Cont. in mg/ml	Growth inhibition in mm				
		HTDP	[Cu(HTDP) ₂] ₂ H ₂ O	[Co(HTDP) ₂]	[Ni(HTDP) ₂] ₂ H ₂ O	STD
<i>Aspergillus</i>	0.25	1 ± 0	4.03 ± 0.05	6.03 ± 0.05	3.03 ± 0.05	12 ± 0
	0.5	1.13 ± 0.05	6.2 ± 0.26	8.06 ± 0.11	5.03 ± 0.05	12 ± 0
	0.75	2.03 ± 0.05	8.03 ± 0.05	10.03 ± 0.07	6.06 ± 0.11	12 ± 0
	1	2.2 ± 0.26	8.33 ± 0.2	10.1 ± 0.17	8.03 ± 0.05	16 ± 0
<i>Fusarium</i>	0.25	1.03 ± 0.05	5.03 ± 0.05	7.05 ± 0.05	2.03 ± 0.05	10 ± 0
	0.5	1.03 ± 0.05	5.03 ± 0.05	7.05 ± 0.05	2.03 ± 0.05	10 ± 0
	0.75	1.23 ± 0.25	8.06 ± 0.11	11.06 ± 0.11	6.1 ± 0.17	10 ± 0
	1	4 ± 0	9.03 ± 0.05	12.03 ± 0.05	10.03 ± 0.05	12 ± 0

*Each value is displayed average ± SD (standard division) of three replicates for the zone of inhibition.

*STD: Nystatin

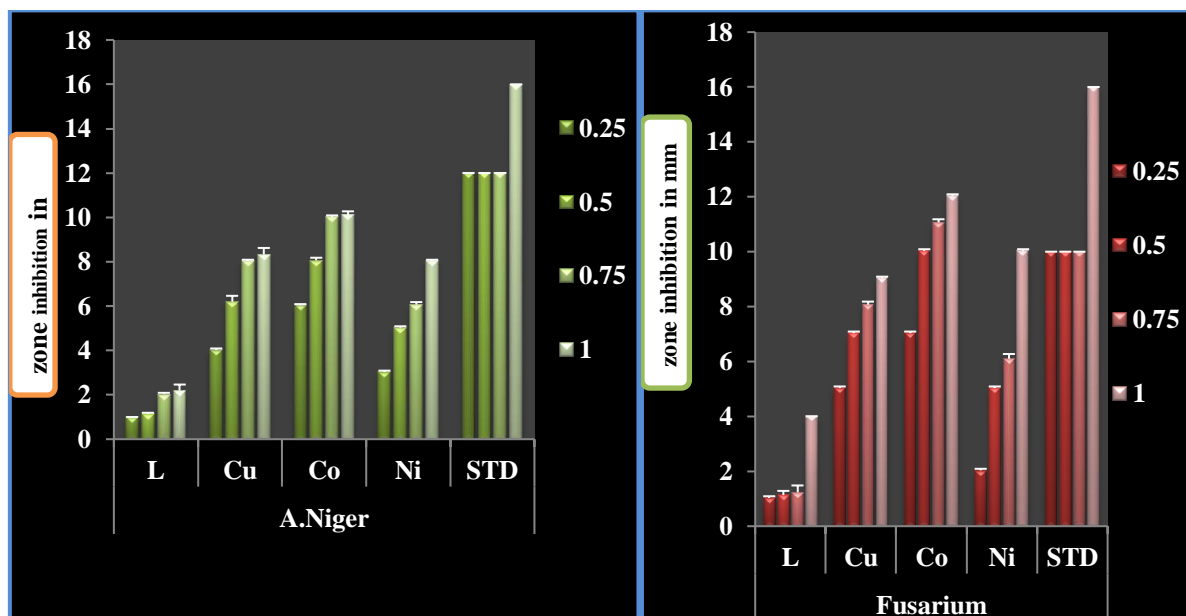


Figure 12. Graphical representation of the antifungal activity of HTDP and its metal complexes.

3.9.2. Antidiabetic studies.

The synthesized HTDP and its metal complexes screened for antidiabetic activity. Inhibitions of enzymes manage the stage of insulin and glucose level in blood over a long period and also control the risk of hyperglycemia. Inhibition screening was recorded with various concentrations of 25, 50, and 100 $\mu\text{g/mL}$ shown in Figure 13. Further, HTDP exhibited 10.2, 24.03, and 33.23% inhibition activity, and its complexes showed enhanced inhibitory percentage results [48].

Table 10. Antidiabetic activity of the ligand and their metal complexes.

Cont. in $\mu\text{g/mL}$	Alpha amylase inhibitory activity in %				
	HTDP	$[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$	$[\text{Co}(\text{HTDP})_2]$	$[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$	STD
25	10.2 \pm 0.25	22.5 \pm 0.43	35.16 \pm 1.6	34.06 \pm 0.11	36.66 \pm 2.08
50	24.03 \pm 0.05	29 \pm 0	45.2 \pm 0.34	46.3 \pm 0.51	49.66 \pm 3.05
1	33.23 \pm 0.40	57 \pm 0.6	84.03 \pm 0.57	78.76 \pm 0.58	92.6 \pm 0.6

*STD Acarbose (Acb) is the standard of alpha-amylase enzyme activity, M \pm SD.

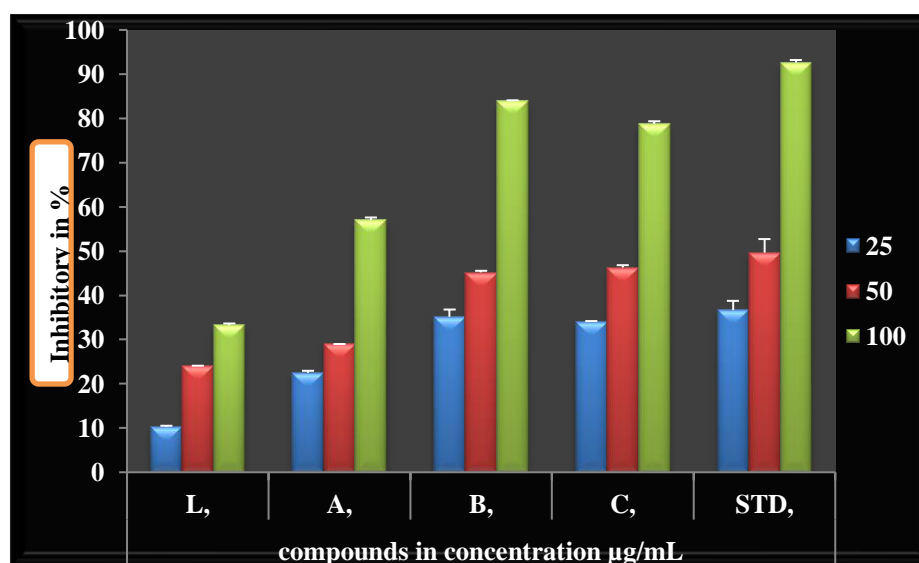


Figure 13. Graph representation of antidiabetic activities of ligand and its metal complexes.

Among the synthesized complexes, the $[\text{Co}(\text{HTDP})_2]$ showed higher inhibitory growth, as shown in Table 10. The potency of the synthesized compound was measured by the IC_{50} method. It indicates how much drug is needed to inhibit a biological process by half. The IC_{50} of HTDP showed at $153.50 \mu\text{g/mL}$, and its $[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$ showed $87.28 \mu\text{g/mL}$, $[\text{Co}(\text{HTDP})_2]$ exhibited $51.16 \mu\text{g/mL}$ while $[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$ showed IC_{50} of $47.08 \mu\text{g/mL}$, standard acarbose drug which was found to have $45.68 \mu\text{g/mL}$ as shown in figure 14 [48-52]. The data is given in table 11.

Table 11. IC_{50} value of ligands and their metal complexes against an alpha-amylase antidiabetic agent in $\mu\text{g/mL}$.

Compounds	IC_{50} value in $\mu\text{g/mL}$	R^2
HTDP	153.50 ± 0.26	0.915
$[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$	87.29 ± 0.51	0.971
$[\text{Co}(\text{HTDP})_2]$	51.16 ± 0.58	0.985
$[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$	47.08 ± 0.05	0.991
Acb	45.68 ± 0.39	0.988

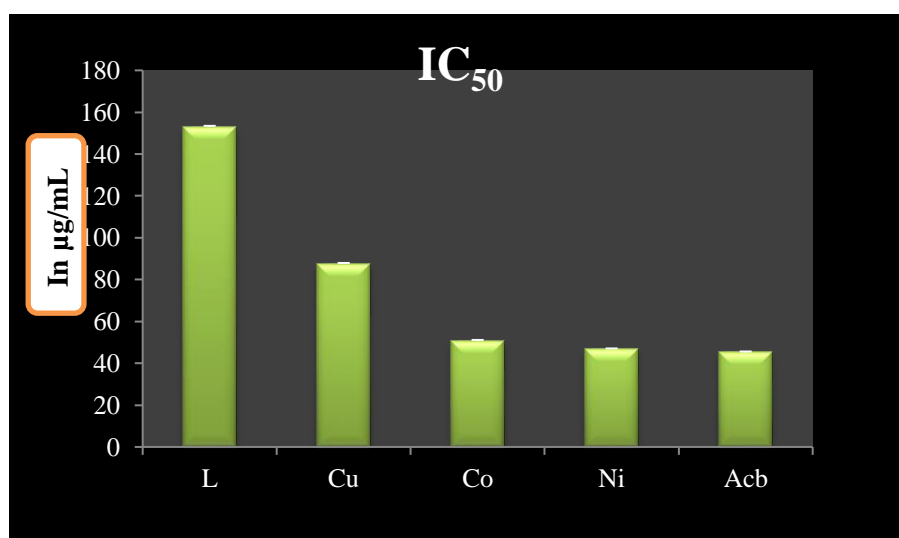


Figure 14. Graph representation of IC_{50} of ligand and its metal complexes in $\mu\text{g/mL}$ against alpha-amylase enzyme activity.

3.9.3. Anti-mycobacterium studies.

Synthesized HTDP and its metal complexes were screened for Mycobacterium tuberculosis strain using tetrazolium salt indicator (MTT) for anti-mycobacterial (M-TB) activity it spread in the eukaryotic area. The color change in the well revealed inhibition of bacterial growth. The MIC percentages were recorded at 37°C incubation in different concentrations of 25, 50, and $100 \mu\text{g/mL}$ with organic anti-infective bacterium drug agent Chloramphenicol (CHL) as standard. The HTDP showed 12, 15, and 22 % activity, and its metal complexes exhibited enhanced inhibition because HTDP act as a charge transfer to the central metal atom. Thus, the complexes form a good binding; therefore, They are stable bioactive molecules. Moreover, the $[\text{Co}(\text{HTDP})_2]$ and $[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$ showed better percentage inhibition at $50 \mu\text{g/mL}$, as shown in Figure 15. The MIC was evaluated by three replicate measurements, as shown in Table 12, and its mean inhibition values find the dose efficiency of compounds as calculated by $Y = \text{MX} + \text{C}$, $\text{IC}_{50}(\text{X}) = \text{Y} - \text{C} / \text{M}$. The HTDP IC_{50} has $317.04 \mu\text{g/mL}$, and their $[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$, $[\text{Co}(\text{HTDP})_2]$, and $[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$ had better half-maximum inhibitory concentration values as tabulated in table13 and the graph showed in figure16 [53].

Table 12. Anti-tuberculosis zone inhibition of ligands and their metal complexes against *M. Tuberculosis*.

Cont. in $\mu\text{g/mL}$	Mycobacterial tuberculosis activity in %				
	HTDP	$[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$	$[\text{Co}(\text{HTDP})_2]$	$[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$	CHL
25	12 \pm 0.25	22.5 \pm 0.43	39.16 \pm 1.6	43.06 \pm 0.11	13.66 \pm 2.08
50	15.03 \pm 0.05	35 \pm 0	52.2 \pm 0.34	54.3 \pm 0.51	30.66 \pm 3.05
100	22.23 \pm 0.40	57 \pm 0.6	81.03 \pm 0.57	60.76 \pm 0.58	72.6 \pm 0.6

*Chloramphenicol (CHL) is considered a standard antibiotic drug. M \pm SD.

Table 13. IC₅₀ value of ligand and their metal complexes against anti-mycobacterium tuberculosis agent in $\mu\text{g/mL}$.

Compounds	IC ₅₀ value in $\mu\text{g/mL}$	R ²
HTDP	317.04 \pm 0.05	0.997
$[\text{Cu}(\text{HTDP})_2]\text{H}_2\text{O}$	84.14 \pm 0.165	0.996
$[\text{Co}(\text{HTDP})_2]$	45.17 \pm 0.156	0.998
$[\text{Ni}(\text{HTDP})_2]2\text{H}_2\text{O}$	48.17 \pm 0.282	0.904
CHL	73.04 \pm 0.05	0.997

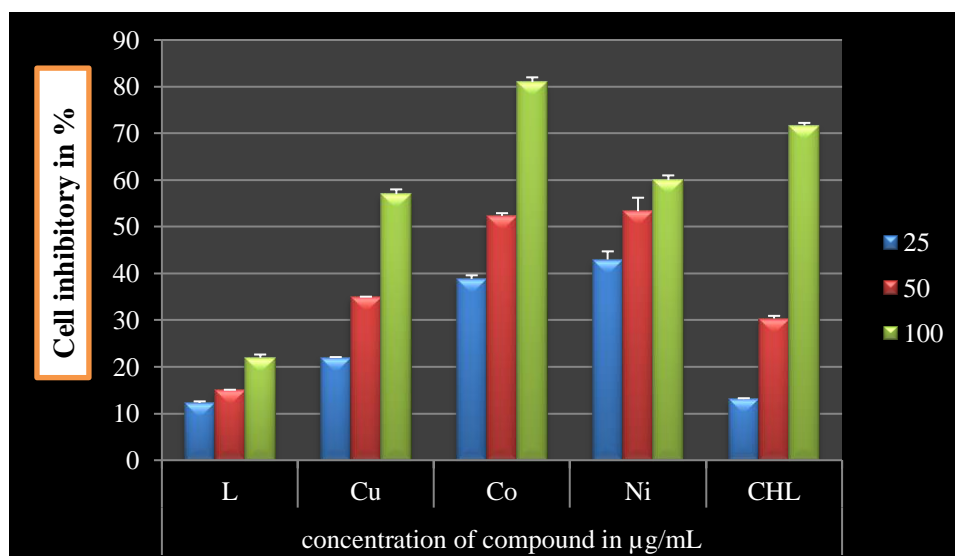


Figure 15. Graphical representation of anti-tuberculosis activities of ligand and its metal complexes.

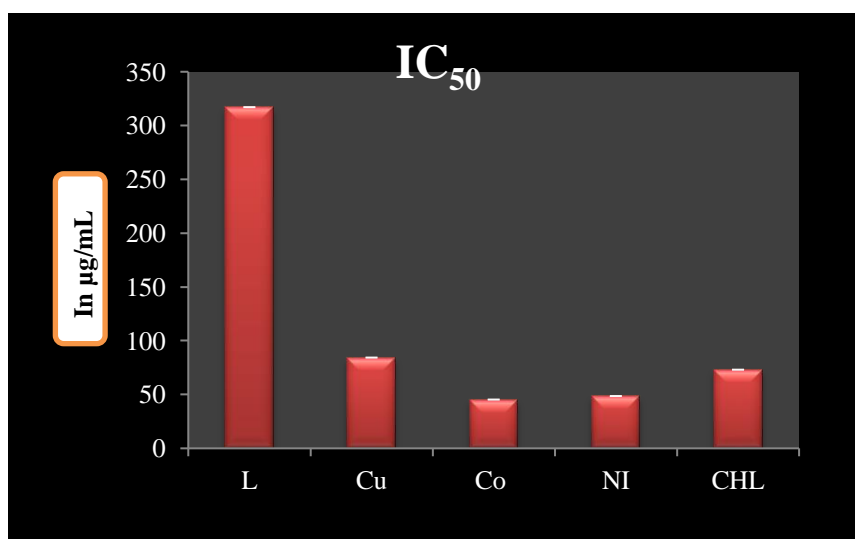


Figure 16. Graph represented IC₅₀ values of ligand and its metal complexes in $\mu\text{g/mL}$ against for mycobacterium agent.

4. Conclusions

In this present investigation, novel HTDP and its coordinated Cu(II), Co(II), and Ni(II) complexes have been synthesized and characterized by various analytical techniques. The molar conductance values indicate the non-electrolytic nature of complexes. Based on UV-Vis

and magnetic susceptibility data, the synthesized complexes proposed to have octahedral geometry. The thermal analysis data showed the mode of degradation of the metal complexes. The modified [Co(HTDP)₂]/GCE showed highly electroactive with nitrate sensing is 0.172 $\mu\text{A } \mu\text{M}^{-1}\text{cm}^{-2}$ by the cyclic voltammeter, and the differential pulse voltammeter showed 11.043 $\mu\text{A } \mu\text{M}^{-1}\text{cm}^{-2}$. The uncoordinated HTDP and their complexes were screened for antimicrobial culture. The clostridium E. coli and fusarium showed better growth of zone inhibition and [Cu(HTDP)₂]H₂O and [Co(HTDP)₂] exhibiting promising results. Inhibition of α -amylase showed positive activity with a potential IC₅₀ value for [Ni(HTDP)₂]2H₂O with 47.08 % inhibition activity, and it is best for diabetic patient and known as starch blockers because they assist dietary of glucose in the antidiabetic study. The [Co(HTDP)₂] and [Ni(HTDP)₂]2H₂O complexes showed good drug potency when compared to the uncoordinated ligand. The HTDP and its complexes showed enhanced activity and can be used as a potent agent against selective pathogens.

Funding

This research received no funding.

Acknowledgments

The authors thank the Manipal Institute of Technology, SAIF Dharwad, SJM College of Pharmacy, Chitradurga, and the University of Mysore for providing spectral facilities. The authors also thank Biotech Pvt Mysore, Karnataka, India, for biological studies.

Conflicts of Interest

The research authors declared no conflict of interest.

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Supplementary materials

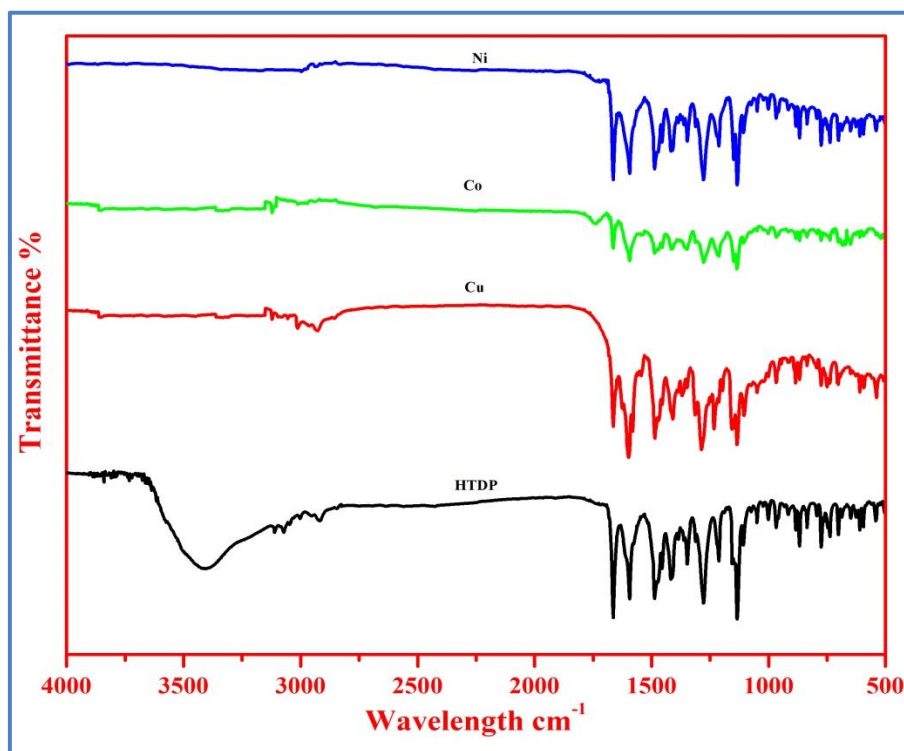


Figure S1. FT-IR spectra of synthesized compounds.

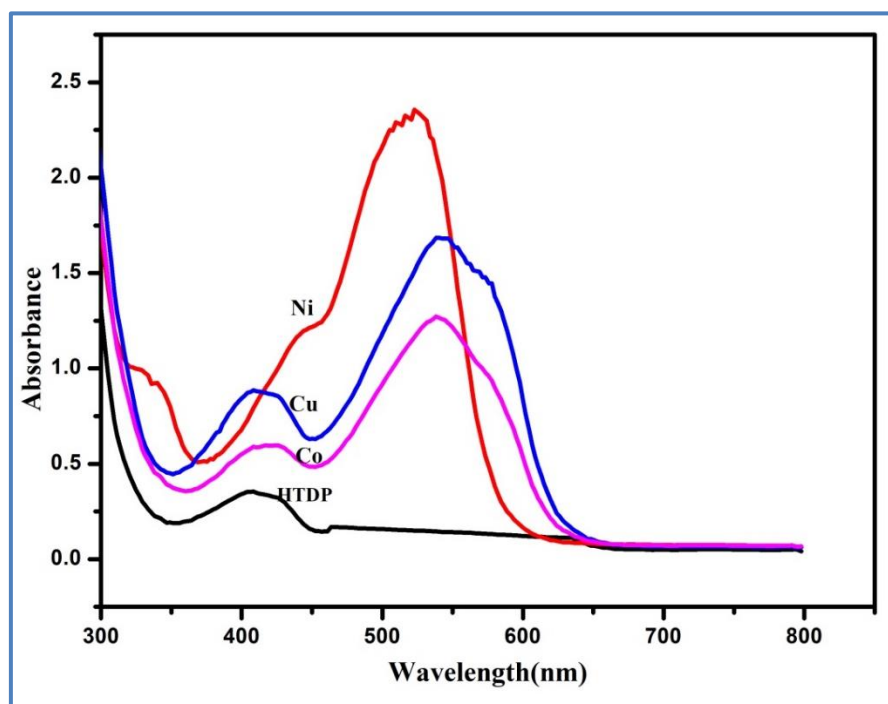


Figure S2. Electronic absorption spectra of synthesized compounds.