










# A Renewed Concept on Diabetic Retinopathy: Polyphenols as a Choice of Solution

Talha Bin Emran <sup>1,2,\*</sup>, M. Akiful Haque <sup>3</sup>, Purnachandra Reddy Guntaka <sup>4</sup>, Laliteshwar Pratap Singh <sup>5</sup>, Mohd. Washid Khan <sup>6</sup>, Aziz Unnisa <sup>7</sup>, Mohammed H. Nafady <sup>8</sup>, Aasim A. Kazi <sup>9</sup>, Koula Doukani <sup>10</sup>, Firzan Nainu <sup>11,\*</sup>

<sup>1</sup> Department of Pharmacy, Faculty of Allied Health Sciences, Daffodil International University, Dhaka 1207, Bangladesh

<sup>2</sup> Department of Pharmacy, B.G.C. Trust University Bangladesh, Chittagong-4381, Bangladesh

<sup>3</sup> Department of Pharmaceutical Analysis, School of Pharmacy, Anurag University, Ghatkesar, Hyderabad 500088, India

<sup>4</sup> Gitam Institute of Pharmacy, Gitam Deemed to be University, Visakhapatnam, Andhra Pradesh, India

<sup>5</sup> Narayan Institute of Pharmacy, Faculty of Pharmacy, Gopal Narayan Singh University, Jamuhar, Sasaram (Rohtas) Bihar 821305, India

<sup>6</sup> Department of P. G. Studies and Research in Chemistry and Pharmacy, Rani Durgavati Vishwavidyalaya, Jabalpur, Madhya Pradesh, India

<sup>7</sup> Department of Pharmaceutical Chemistry, College of Pharmacy, University of Ha'il, Ha'il, Saudi Arabia

<sup>8</sup> Faculty of Applied Health Science Technology, Misr University for Science and Technology, Giza 12568, Egypt

<sup>9</sup> Department of Pharmaceutical Chemistry, NBS Institute of Pharmacy, AUSA 413520, Maharashtra, India

<sup>10</sup> Faculty of Nature and Life Sciences, University of Ibn Khaldoun –Tiaret, Algeria

<sup>11</sup> Department of Pharmacy, Faculty of Pharmacy, Hasanuddin University, Tamalanrea, Makassar 90245, Indonesia

\* Correspondence: talhabmb@bgctub.ac.bd (T.B.E); firzannainu@unhas.ac.id (F.N.);

Scopus Author ID 57120069200

Received: 8.01.2023; Accepted: 26.02.2023; Published: 7.04.2023

**Abstract:** Diabetic retinopathy (DR) is the widespread microvascular consequence of diabetes mellitus and the most common effect of blindness in people with diabetes. Existing drugs are only effective in advanced stages of DR, and long-term efficacy and safety results for these treatments have yet to be clarified in multiple clinical trials. Furthermore, a more trustworthy and effective medication that may be deemed more advantageous in managing DR is unavoidable. Polyphenols, which are plant-derived chemical compounds, may be helpful in the initial stages of DR treatment. Compounds rich in polyphenols have been shown to slow the progression of long-term consequences of diabetes, for example, heart disease, nerve damage, kidney damage, and retinopathy. Polyphenols could be used instead of traditional treatments to halt the disease's progression. It has been proposed that *in vitro* investigations on the effects of polyphenols on ocular vision physiology and antioxidant protection have a substantial bearing on this assertion. Among the benefits of polyphenols are scavenging the free radicals, lowering the production of advanced glycation end products, inhibiting aldose reductase, anti-inflammatory activity, and affecting ocular blood flow. The present review discussed the significance of polyphenols in preclinical and clinical research of DR-affected cellular and molecular pathways.

**Keywords:** polyphenols; diabetes retinopathy; oxidative stress; reactive oxygen species; a clinical study.

© 2023 by the authors. This open-access article is distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Diabetes mellitus (DM) is a metabolic condition triggered by an inability to produce or use insulin. Diabetes has significant socioeconomic effects since it affects many populations [1]. The diabetes epidemic severely influences individuals' health and financially impacts

society and healthcare. Cardiovascular system complications, such as diabetic retinopathy (DR), diabetic neuropathy, and diabetic nephropathy, are the most widespread reasons for death among people with diabetes. DR is an essential microvascular difficulty of diabetes, the primary trigger of vision loss and blindness worldwide, with a neurodegenerative component. As the disease becomes increasingly frequent, the number of people with diabetes-related kidney disease (DRKD) is anticipated to rise to 191 million by 2030 [2].

The retina's microvasculature has been destroyed, and penetration has increased due to DR, leading to blindness in the long run. According to studies, animal models with thinner retinas appear to have fewer retinal ganglion cells. Furthermore, apoptosis occurs more quickly in those retinas as dead cells increase, even before histopathological indications appear [3].

Many components and metabolites affect the diabetic animal's retinas, causing retinal cell destruction and influencing disease development. Oxidative stress (OS), which cooperates with many signaling routes, is one of the most critical components in the complicated process of diabetes [4]. DR has been linked to various metabolic processes that are difficult to comprehend, including the heightened development of advanced glycation end products (AGE), protein kinase C activation (PKC), and stimulation of the polyol and hexosamine pathways [5]. The diabetes retina's dysregulation of different metabolites affects a diversity of intermediaries, including cytokines, chemokines, growth factors, neurotrophic factors, and adhesion molecules. An expansion in OS in retinal cells causes DR [6].

Nature offers diverse therapeutically beneficial compounds with fewer side effects and lower costs. These naturally produced compounds, such as alkaloids, tannins, polyphenols, terpenoids, flavonoids, and steroids, have a variety of pharmacological activities that target many metabolic pathways [7]. These spontaneously produced molecules' improved intercellular medication distribution is critical in creating sound therapeutic effects [8,9]. The capacity of plant-derived chemical agent polyphenols to increase the intercellular drug transport mechanism is essential to establishing favorable treatment outcomes for DR [10,11]. This review highlighted the importance of polyphenols in treating various ailments and their potential relevance in developing novel therapeutics for diabetes-related retinal damage.

## **2. Risk Factors and Epidemiology**

Compared to their research in 2010 [12], the Visual Loss Expert Group's most recent report in 2015 found that DR has slipped from 5<sup>th</sup> to 6<sup>th</sup> position as the highly prevalent trigger of avoidable vision impairment globally [13]. Even though all reasons for blindness and vision impairment have decreased, DR was the only one that remained blind, an eye ailment that has been more common globally since 1990. DR mild-to-moderate vision impairment boosted by 7.7%, while DR moderate-to-severe vision impairment increased by 28.6%. This red flag in 2015 indicated that the existing screening system needed to be beefed up. Before the commencement of the illness, increase research programs directed at novel molecular targets and management strategies targets [13,14].

The worldwide predominance of DR was expected to be 34.6% for all types of DR in 2010, 6.81 % for diabetic macular edema (DME), and 6.96 % for proliferative diabetic retinopathy (PDR), and 10.2% for vision-threatening diabetic retinopathy (VDR) in 2010. Retinopathy includes conditions like PDR and DME. This equates to around a third of the total population. Because of DR, one-tenth of diabetics will require active therapy. The risk of acquiring a DR was 2.7 times superior in thin type 1 diabetes (T1DM) than in type 2 diabetes (T2DM). T2DM is a kind of diabetes that develops after at least 20 years [15]. The latest nine-

year prospective research study comprising T1 and T2 patients reported a 15.16 to 2.19% yearly incidence in T1DM, compared to 8.37 to 2.19% in T2DM. Low-density lipoprotein (LDL) cholesterol and arterial hypertension are all under control in DM duration and glycaemic management. In both T1DM and T2DM patients, high creatinine levels were revealed to be substantial risk factors for DR progress. The T1DM sample had a more significant predominance and occurrence of DR at the end of the trial. This study found a link between having diabetes for more extended and having lower basal and measured glucose levels [16].

There is proof that oily fish, dietary fiber, and Mediterranean help guard against DR in nutritional practices. Conversely, higher-calorie consumption has been related to a higher risk of DR. Furthermore, antioxidants, particularly vitamins C, E, and carotenoids, act as defensive agents, according to a recent study on antioxidants in typical diets [17].

### **3. Pathophysiology of Diabetic Retinopathy**

DR must be regarded as multifactorial since it develops directly from persistent hyperglycemia, despite being impacted by the risk factors previously discussed [18–20]. From a metabolic aspect, OS was discovered to play a vital role in the growth of DR, acting as a relationship between many variables. Glucose-mediated biochemical activities, an excess of glucose, cause the mitochondria to become activated. The polyol pathway, the manufacture of AGEs, and their activation improve PKC and increase the hexosamine pathway. These variables boost the reactive oxygen species (ROS) production [21–23]. These molecules are regularly generated for regular cell activity. The body's antioxidant capabilities assist in neutralizing the effects (Figure 1).

Furthermore, the accumulation of ROS leads to changes in mitochondrial DNA structure, which leads to changes in gene expression. ROS accumulation is exacerbated in mitochondrial malfunction [23,24] because OS has a long-term influence on diabetes. Even when correct glycaemic management is established, this phenomenon, known as metabolic memory, has been hypothesized to cause disease development [24]. Glycaemic-induced OS is exacerbated by arteriosclerosis, which worsens molecular damage [25]. Cellular dysfunction and apoptosis are caused by hyperglycemia-induced metabolic alterations and ROS increase [26].

#### *3.1. Inflammation.*

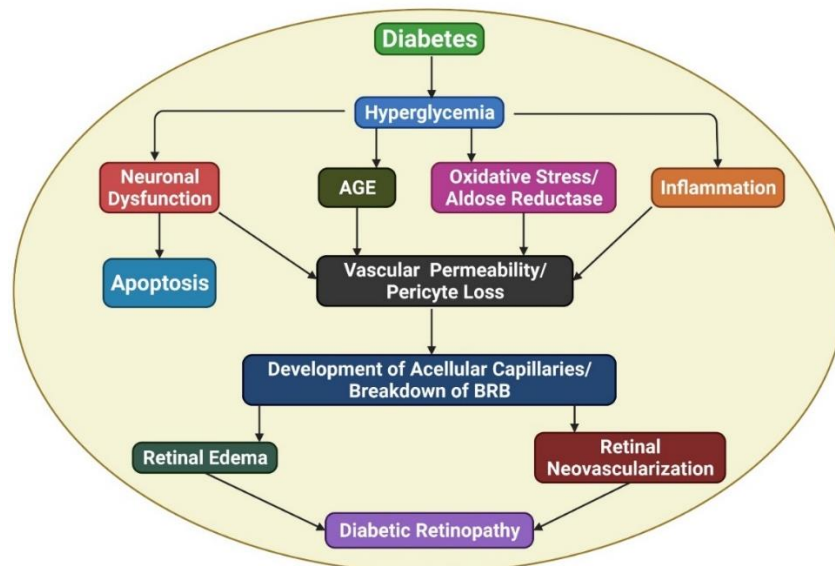
Microglial activation is a preliminary result of DR growth and can produce inflammatory mediators [27,28]. Pro-inflammatory cytokines are found at higher human concentrations (interleukins IL-1 $\beta$ , IL-6, IL-8, and TNF- $\alpha$ ). Vitreous samples have also been correlated with the seriousness of DR [29-31]. In response to inflammatory stimuli, endothelial cells enhance the manifestation of intracellular and extracellular proteins. Leukocyte adherence is facilitated by E-selectin and vascular adhesion molecules (ICAM-1 and VCAM-1). The formation of leukostasis, or the creation of white blood cells, is linked to linkage to endothelial cell walls—a critical factor in the progression of posterior microvascular destruction [32,33]. In diabetes blood samples, higher levels of the components mentioned above were found, and blocking ICAM-1 in diabetic retinal endothelial cells decreased cell death in cultured human retinal endothelial cells [34]. Surprisingly, administering an antioxidant agent reduced cellular loss while lowering the ICAM-1 level in those retinal cells [34].

### 3.2. Neurodegeneration.

Programmed cell death appears to influence neurons before vascular cells in neurodegeneration. Before the onset of DR clinical symptoms and microvascular changes, electroretinogram (ERG) tests have suggested remaining neuronal damage. Furthermore, diabetic animal models and human participants reduced the ganglion cell inner layer [35,36]. ERG tests were conducted on induced-diabetes mice pre and post-antioxidant treatment to study the defensive impact of lutein on visual purpose and histological neuronal variations related to OS about neurodegeneration [37].

### 3.3. Pericytes layer.

A basement membrane, an external pericytes layer, and an interior endothelial cells layer contribute to micro-vasculopathy in retinal capillaries. Pericyte loss occurs in hyperglycemic conditions [38], leading to localized microvascular dilatation and microaneurysms. Debris deposition and endothelial cell failure show the disturbing blood-retinal barrier (BRB), enhancing vascular absorptivity, exudation, and hemorrhages [39]. Leukostasis, which results from an inflammatory reaction, damages endothelial cells and leads to microvascular occlusions [40]. Following hypoxia, the transcription factor hypoxia-inducible factor 1 (HIF-1) is activated, which enhances the release of vascular endothelial growth factors (VEGFs) and leads to the creation of neovessels [41].

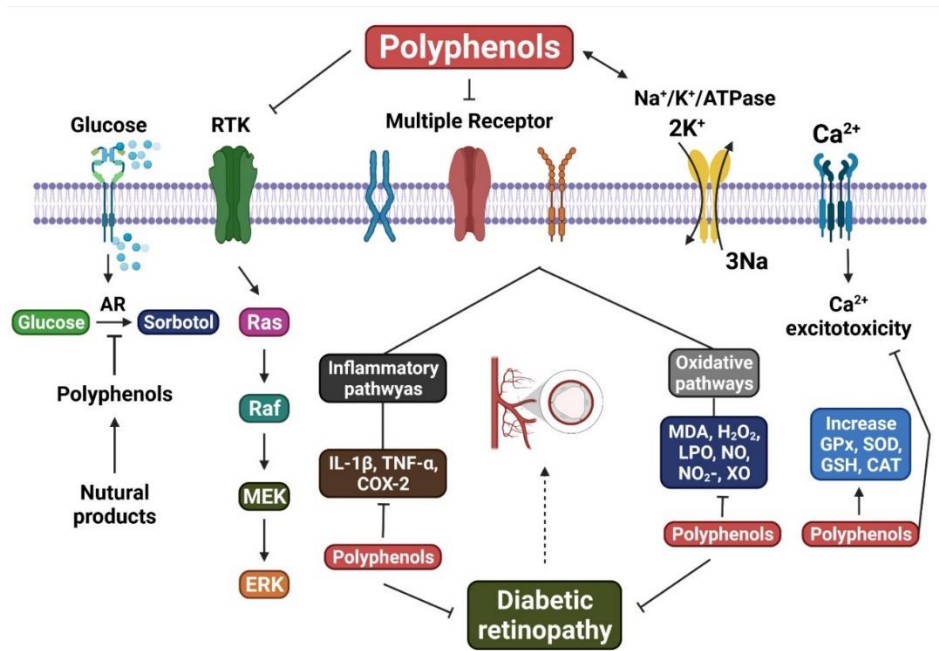


**Figure 1.** Diabetic retinopathy Growth and progress. A.G.E., advanced glycation end products; BRB, blood-retinal barrier.

## 4. Polyphenols and Diabetic Retinopathy

A group known as polyphenols or polyphenolics is naturally occurring and extensively dispersed in the kingdom of plants [42]. These compounds are distinguished by aromatic rings attached to phenol structural units. Regarding physical, biological, and metabolic qualities and functions, these secondary metabolites of plants are primarily defined by their phenol structural units. Polyphenols are split into two primary categories established on their molecular structure: flavonoids and non-flavonoids (Figure 2). Stilbenes, phenolic acids, lignans, and other polyphenols are subclassified as non-flavonoids [43]. Flavonoids are polyphenolic

molecules in vegetables and fruit, accounting for approximately 60% of total polyphenols [44]. Flavonoids can be divided into six subtypes: flavanol, flavones, flavanones, isoflavonols, flavonols, and anthocyanin-containing flavonols [45].



**Figure 2.** Mechanistic role of polyphenols against diabetic retinopathy. TNF- $\alpha$ , tumor necrosis factor alpha; COX-2, cyclooxygenase-2; ERK, extracellular signal-regulated kinase; LPO, lactoperoxide; NO, nitric oxide; XO, xanthine oxidase; MDA, malondialdehyde; CAT, catalase; SOD, superoxide dismutase; GSH, glutathione; GPx, glutathione peroxidase.

#### 4.1. Resveratrol.

Resveratrol (RSV) is abundant in blueberries, raspberries, mulberries, red wine, and grape skin [46]. Anti-angiogenic, antioxidant characteristics, antiproliferative, antiplatelet, endothelial, anti-inflammatory, and neurogenic capabilities are all found in RSV [47]. When administered orally, RSV is absorbed 75%, primarily by trans epithelial diffusion, but bioavailability is less than 1%. This is because glucuronidated chemicals are engaged and sulfated to easily remove essential metabolites in the colon and liver. On the other hand, bioavailability varies substantially between people due to considerations including age and gender [47–49]. Because RSV is a hydrophobic molecule, it is captivated by intestinal epithelial cells, hepatocytes, and breast carcinoma cell lines [47]. Resveratrol therapy was revealed to increase diminished glutathione (GSH) levels in erythrocytes and the ocular degree in rats, where GSH serves as an antioxidant [50,51]. RSV restrains the action of an enzyme associated with neovascularization, endothelial nitric oxide synthase, and inflammatory processes in rats' eyes [52,53]. In diabetes, RSV therapy reduces the number of vascular leaks, pericyte loss, and VEGF levels. According to Luna *et al.*, RSV reduced ROS production, which prevented the development of pro-inflammatory markers such as IL-1 $\beta$ , IL-6, and IL-8 [54].

#### 4.2. Curcumin.

Curcumin is a crystalline orange-yellow combination that is one of *Curcuma* spp. Main's components [55]. The WHO recommends daily consumption of 0-3 mg/kg as a dietary ingredient [56]. Recently, curcumin has been beneficial in treating DR. Curcumin performs as an antioxidant by diminishing free radicals [57]. Curcumin boosts the mRNA appearance of



antioxidant enzymes like SOD and catalase, decreasing oxidative and synchronizing nitrosative DNA destruction [58]. Curcumin stimulates a mitochondrial pathway by altering the respiratory activity of mitochondrial complexes I, II, III, and V while simultaneously activating erythroid 2-related factor 2 (Nrf2) [59]. Curcumin can enhance antioxidant capability and hypoglycemic and prophylactic anti-inflammatory action in diabetic rats' retinas by cutting pro-inflammatory cytokines levels like IL-1 $\beta$ , TNF- $\alpha$ , and VEGF [60,61], as well as 5-hydroxyeicosatetraenoic acid and a dual inhibitor of arachidonic acid [62]. Curcumin is an anti-angiogenic agent that lowers stromal cell-derived factor 1 alpha and inhibits retinal human endothelial cells [63].

#### 4.3. Quercetin.

Quercetin protects the retinas of diabetic patients from a range of injuries [64,65]. High glucose-stimulated irritation and programmed cell death generated IL-6, monocyte chemoattractant protein-1 (MCP-1), and ROS, which were decreased by quercetin. MiR-29b may be a good target for DR control since its anti-angiogenic impacts on diabetic animals' retinas by reducing RME production, relocation, and angiogenesis. Additionally, ARPE-19-treated cells have increased miR-29b expression, and lower miR-29b expression has been associated with a diminished quercetin protective effect in patients [66]. Therefore, the PTEN/Akt pathway stimulus and the inhabitation of the nuclear factor kappa B (NF- $\kappa$ B) path are primarily due to miR-29b expression [67]. *In vivo*, quercetin reduced caspase-3 and NF- $\kappa$ B expression, indicating antiapoptotic effects [68]. This flavonol is also responsible for inactivating neovascularization-related proteins like VEGF and MMP-99. Quercetin decreased TNF- $\alpha$  and IL-1 $\beta$  levels while increasing antioxidant enzymes (CAT, SOD, and GSH).

Expression of the AQP4 and GFAP were also inhibited by quercetin [68,69]. Another investigation discovered that quercetin has anti-diabetic effects in STZ-induced diabetic rats. An intraperitoneal quercetin dose significantly reduced IL-18, IL-1 $\beta$ , TNF- $\alpha$ , IL-6, and NLRP3 inflammasome over-activation at a 150 mg/kg dose for 16 days ganglion cells and thickness of the retinal cell layer were improved [65].

#### 4.4. Catechins, epicatechin, epigallocatechin and epigallocatechingallate.

Catechins are derived from natural sources (grapes, cocoa) and red wine, although green tea and chocolates have the highest concentrations. Catechin and epicatechin are key flavanols found, whereas galocatechin, fruits, epigallocatechin, and epigallocatechin gallate are primarily found in leguminous plant seeds [70]. The aggregation of advanced glycation end products (AGEs) plays a direct part in the turning on and improvement of DR [71]. Kim and colleagues studied the probable outcomes of AGE-injected rats in an *in vivo* and *in vitro* study. Combining (-)-epicatechin and glycated human serum albumin detached from diabetic patients increased AGE breakdown. The treatment of (-)-epicatechin in exogenously AGE-injected rats reduced retinal vascular apoptosis and the retina's AGE burden. As a result, (-)-epicatechin could be a promising treatment option for DN [72]. Skopinski *et al.* found that flavanols isolated from plants, including epigallocatechin (EGC) and epigallocatechin gallate (EGCG), inhibited the angiogenic actions in rats considered [73]. Green tea (*Camellia sinensis*) is a sustainable supplier of antioxidants, primarily due to its high flavonoid content, including EGCG [74]. Green tea has been shown to have therapeutic consequences in contrast to glutamate toxicity in diabetic rat retinas. Green tea extracts were found to reduce retinal

problems in diabetic rats; according to Silva *et al.*, Green tea reduced elevated glial fibrillary acidic protein (GFAP), glutamine synthetase, and oxidative retinal indicators. In diabetic rats treated with tea extract, N-methyl-D-aspartate R1 (NMDAr1), glutamate aspartate transporter (GLAST-1), and obstructing were likewise lowered. These results suggest that EGCG could be an effective treatment for DR. Due to high glucose levels, Müller cells produced too much glutamine synthetase and ROS but insufficient glutamate transporter, GSH, or glutamate receptor. Green tea treatment effectively reversed all changes mentioned above in diabetic rats' retinal cells [75].

Zhang *et al.* examined the effect of EGCG on the HREC line. The cell viability, cell cycle, and mortality were studied using the MTT assay and flow cytometry. According to the findings, medication of HRECs with EGCG dramatically reduced programmed cell death. EGCG also suppressed VEGF expression and lowered the expression of ERK1/2 and MAPK [76]. In the last stages of DR, metalloproteinase-9 (MMP-9) aids vascular penetrability and neovascularization. The effective primary constituent of green tea extract, EGCG, inhibited TNF- $\alpha$  and MMP-9 production in human retinal pigment epithelial cells 12-O-tetradecanoylphorbol-13-acetate (TPA) (HRPECs). In ARPE-19 cells, EGCG consumption inhibited apoptosis by dropping ROS levels and decreasing mRNA expression of VEGF, MMP-9, and VEGF Receptor-2. Moreover, in VEGF-treated HRPECs, EGCG administration dramatically inhibits proliferation, tube formation, and vascular permeability. EGCG reduced vascular leakage and permeability in animal studies [77]. In another study, EGCG improved autophagy in Müller cells by enabling autophagosome formation, boosting lysosomal acidity, and boosting autophagic flux. Furthermore, it increased cell proliferation, protected cells from apoptosis, and decreased retinal destruction produced by high glucose levels [78].

#### 4.5. Rutin

Rutin reduces apoptosis in the diabetic retina by decreasing caspase-3 and raising neurotrophic elements (BDNF and nerve growth factor (NGF)) as well as Bcl-2 levels [79]. Furthermore, rutin therapy reduced TNF- $\alpha$ , aldose reductase protein, and VEGF expressions, restoring fluorescein vascular seepage. It also can boost overall antioxidant activity in the retina [80].

#### 4.6. Galangin

Galangin flavonol reduced blood-retina barrier (BRB) breakdown by reducing inflammation caused by microglia and so repaired BRB malfunction caused by TNF- $\alpha$  [81] via increasing the Nrf2 pathway. Galangin therapy boosted strong intersection enzyme representation and diminished BRB destruction in ARPE-19 cells and HRECs exposed to TNF- $\alpha$ . Galangin also mimicked the initiation of Nrf2 and the decrease of ROS construction, resulting in the alteration of many antioxidant genes [81].

#### 4.7. Kaempferol

Protein, mRNA for a placenta growth factor (PGF), and VEGF were raised in HRECs under hyperglycemic conditions. To reduce angiogenesis, kaempferol treatment reduced the expansion of angiogenic factors. Anti-angiogenic effects of kaempferol are further demonstrated by inhibiting phosphorylation processes of specific kinases (Akt1, Erk1/2, and Src) beside PI3K manifestation induced by hyperglycemia [82]. Digestive enzymes,  $\alpha$ -

glucosidase and  $\beta$ -amylase, are responsible for converting hydrolyzed carbohydrates into glucose that can be absorbed. Kaempferol slowed down these enzymes' activity leading to a lower and more stable blood sugar spike after a meal. This is a significant way to manage T2DM [83].

In addition, kaempferol was tested to protect RGCs from high-glucose-induced damage. At a concentration of 0, 20, 40, 60, 80, or 100  $\mu\text{mol/L}$ , RGC cells were exposed to kaempferol and increased glucose (55  $\text{mmol/L}$ ) for 48 h. After testing, it was discovered that treatment with kaempferol dramatically reduced ROS levels, caspase-3 activity, lactate dehydrogenase leakage, and cell death. The extracellular signal-regulated kinase (ERK) phosphorylation and expression of vasohibin-1 (VASH1) were also increased by kaempferol, which inhibited ERK phosphorylation [84] (Table 1).

**Table 1.** Preclinical investigations of polyphenols in the protection of diabetic retinopathy.

Bioactive compound	Sources	Dose/Concentration	Study model	Outcomes	Refs
Resveratrol	Grapes, berries and peanuts	5 mg/kg/day; Four months	STZ-nicotinamide-stimulated diabetic retinopathy	Decreased superoxide dismutase function and oxidative stress	[85]
Resveratrol	Red wine and grape skin	5, 50, 100, or 200 mg/kg; 5 days	Male C57/B6 mice	Lowered ICAM-1 and MCP-1 protein levels in the retina	[86]
Resveratrol	Grapes, berries and peanuts	5, 10, or 50 $\mu\text{g/kg/day}$ ; 12 weeks	STZ-mediated diabetic rats	Retinal vascular penetrability and inflammatory arbitrators such as Ox-LDL, LDL, MCP-1, A.G.E.s, IL-1 $\beta$ , IL-6, IFN $\gamma$ , and VEGF were reduced.	[87]
Resveratrol	Red wine and grape skin	20 mg/kg; 4 weeks	STZ-intervened diabetic rats	By suppressing retinal CAMKII activity, the NF- $\kappa$ B was downregulated.	[88]
Curcumin	<i>Curcuma longa</i>	100 and 200 mg/kg; 16 weeks	Diabetic rat model	Diabetes-stimulated retinal ultrastructure alterations include retinal ganglion cell death, retinal shrinkage, and photoreceptor cell membrane disruption.	[89]
Curcumin	<i>Curcuma longa</i>	80 mg/kg; daily	STZ-mediated diabetic rats	GSH levels were not depleted, and oxidative stress was reduced.	[90]
Curcumin	<i>Curcuma longa</i>	1 g/kg; 16 weeks	STZ-mediated diabetic rats	VEGF, TNF- $\alpha$ expression, and GSH, CAT, and SOD levels were reduced.	[61]
Quercetin	Fruits and vegetables	20, 40, 80 $\mu\text{mol/L}$ ; daily	High-glucose exposed HRECs	Diminished angiogenesis and apoptosis; lowered caspase-1, IL-18, Beclin-1, and NLRP3 expression.	[91]
Quercetin	Fruits and vegetables	150 mg/kg; 16 days	STZ-stimulated diabetic rats	TNF- $\alpha$ , IL-6, IL-18, and IL-1 $\beta$ expressions were decreased, and the	[65]



Bioactive compound	Sources	Dose/Concentration	Study model	Outcomes	Refs
				NLRP3 inflammasome was overactivated.	
Quercetin	Fruits and vegetables	10, 20, 30, 40, and 50 $\mu$ M; 1 day	ARPE-19 cells treated with high glucose	Reduced expression of Bax, p53, and cleaved caspase-3; reduced apoptosis, IL-6, MCP-1, and ROS generation	[92]
Catechin	<i>Camellia sinensis</i>	50, 100, and 200 mg/kg; 8 weeks	STZ-mediated diabetic rats	Suppressed the NF- $\kappa$ B pathway, lowering IL-1 $\beta$ , IL-6, and TNF- $\alpha$ levels while increasing HSP27 levels.	[93]
(-)-epicatechin	Cocoa and green tea	50 mg and 100 mg/kg; 2 weeks	AGE-treated rats	Increased AGE splitting action and reduced AGE load; enhanced retinal vascular programmed cell death	[94]
EGCG	<i>Camellia sinensis</i>	0, 10, 20, 30, 50 $\mu$ M; 24 hours	Glucose-exposed retinal Müller cells	Increased cell growth and apoptosis; reduced retinal degeneration	[78]
EGCG	<i>Camellia sinensis</i>	5.7 g/kg; 12 weeks	STZ-mediated diabetic rats	BRB breakdown is reduced; the occludin, GLAST, and NMDAr1 subunits are attenuated; electroretinography recordings are improved; reduced amounts of glutamine synthetase and GFAP	[75]
EGCG	<i>Convallaria majalis</i>	2, 20, and 200 $\mu$ g/ml; every day	Balb/c mice	Suppression of angiogenic functions	[73]
EGCG	<i>Camellia sinensis</i>	20 and 40 mM; 24 hours	High glucose-treated HRPECs	Decreased apoptosis; decreased ERK1/2 and MAPK expression; suppressed VEGF expression.	[73]
EGCG	<i>Camellia sinensis</i>	200 mg/kg EGCG; 4 days	VEGF-treated Sprague-Dawley and balb/c mice	BRB breakdown was reduced, and vascular leakage was also reduced.	[76]
Rutin	Onions, tea, apples, and red wine	100 mg/kg; 5 weeks	STZ-facilitated diabetic rats	Bcl-2, G.S.H., NGF, BDNF, and blood insulin levels were elevated, while TBARS, caspase-3, and blood sugar levels were reduced.	[77]
Rutin	Onions, tea, apples, and red wine	50 mg/kg; 24 weeks	STZ-facilitated diabetic rats	TNF- $\alpha$ , VEGF, and aldose reductase levels decreased, fluorescein vascular seepage lowered, and antioxidant potential increased.	[79]
Galangin	<i>Alpinia officinarum</i>	20 and 50 $\mu$ M; 6 hours	Microglial BV2 cells handled with D-glucose	Reduced NF- $\kappa$ B, TNF- $\alpha$ , and Egr1 protein levels; reduced ROS generation, phosphorylate n of ERK1/2, microglia cells, and BRB degradation.	[80]

Bioactive compound	Sources	Dose/Concentration	Study model	Outcomes	Refs
Kaempferol	<i>Kaempferia galanga</i>	0, 20, 40, 60, 80, or 100 µmol/L; 48 hours	High-glucose stimulated injury in RGC cells	Reduced R.O.S. levels, caspase-3 activity, lactate dehydrogenase permeability, and death; suppressed ERK phosphorylation while increasing ERK phosphorylation and production of vasohibin-1 (VASH1)	[95]
Kaempferol	Tea, grapefruit, broccoli	5 and 25 µM; 1 day	HREC's handled with high glucose	Lowered PI3K expression, PGF mRNA, and VEGF levels; Src, ERK1/2, and Akt1 initiation decreased.	[84]
Puerarin	Pueraria plants	250 and 500 mg/kg; 4 weeks	STZ-facilitated diabetic rats	MDA and STAT3 levels have decreased, while SOD activity has enhanced.	[96]
Puerarin	Pueraria plants	80 mg/kg; 4 weeks	STZ-mediated diabetic rats	Inhibited AGE-modified protein levels	[97]
Puerarin	<i>Pueraria lobata</i>	1, 5, and 10 µM; 1 hour	Bovine retinal pericytes exposed to AGE-BSA	Lowered ROS generation and NADPH oxidase function; lessened NF-κB stimulation and phosphorylation of p47phox and Rac1	[98]
Puerarin	<i>Radix puerariae</i>	10, 25, and 50 µM; 24 hours	TR-iBRB2 cells treated with IL-1β	Expression of ICAM-1, VCAM-1, and Bcl-2 is reduced; Cell death, leukostasis, and mitochondrial dysfunction are reduced, and NO and SOD synthesis are increased.	
Puerarin	Pueraria plants	125, 250, and 500 mg/kg; 4 weeks	STZ-facilitated diabetic rats	Reduced NF-κB p65 activation and apoptotic in retinal cells	[99]
Naringenin	Citrus fruits	50 mg/kg; 5 weeks	STZ-treated diabetic rats	G.S.H., Bcl-2, TrkB, BDNF, and synaptophysin levels were raised; Bax, TBARS, and caspase-3 levels were significantly lower.	[100]
Eriodictyol	<i>Eriodictyon californicum</i>	0.1, 1, and 10 mg/kg; 10 days	STZ-handled with diabetic rats	Reduced plasma lipid peroxidation; reduced ICAM-1, eNOS, TNF-α, and VEGF expression; inhibited BRB degradation.	[101]
Eriodictyol	<i>Eriodictyon californicum</i>	5, 10, and 20 µM, one day	RGC-5 cell line treated with high glucose	Reduced IL-8 and TNF-α; synthesis of ROS was reduced; decreased cell mortality; enhanced Nrf2 nuclear translocation C.A.T., GPx, and SOD activities have all	[102]

Bioactive compound	Sources	Dose/Concentration	Study model	Outcomes	Refs
				improved; increased expression of heme-oxygenase-1 and cell viability.	
Diosmin	Dehydrogenation of hesperidin	0.1, 1, and 10 µg/mL	ARPE-19 cells treated with high glucose	Phosphorylation of JNK and p38 is reduced; Caspase-3, R.O.S., and cytochrome c release were all inhibited; Bcl-2/Bax and cell growth have enhanced.	[103]
Chrysin	Honey, propolis, honeycomb, and passion flowers	1, 10, and 20 µM; 3 days	RPE cells treated with high glucose	Lowered ER stress; lower levels of IGF-1, VEGF, and PEDF; lessened AGE release and RAGE production	[104]
Chrysin	Honey, propolis, honeycomb, and passion flowers	10 mg/kg; 10 weeks	db/db mice	The development of RAGE and the release of AGE were both inhibited.	[105]

#### 4.8. Anthocyanins.

Anthocyanins are flavonoids found in berries with anti-inflammatory properties [106,107]. Anthocyanins lower several illnesses, including cardiovascular disease, neurological disease, and cancer [108]. Berry anthocyanins also improve brain neuronal and cognitive function. Anthocyanins can prevent OS and its products and the production of pro-inflammatory molecules [109,110]. Anthocyanins have been demonstrated to treat ocular diseases like DR [109] effectively. According to Song *et al.* [111], anthocyanins protect diabetic optic cells against oxidative damage and inflammation.

#### 4.9. Puerarin.

Teng and colleagues investigated Puerarin's antioxidant and antiapoptotic activities in diabetic retinopathy rats. The STZ. group, the STZ. + Puerarin group and the control group were among the three groups in the study. Reverse transcription-polymerase chain reaction (RT-PCR), retinal histopathology, and electron microscopy were used to investigate the consequences of Puerarin on VEGF and HIF-1 gene expression. The morphological modification in both the medial and lateral nuclear layers was downregulated by Puerarin, which decreased the DR produced by STZ. The VEGF and hypoxia-inducible factor (HIF-1) in DR rats were also influenced by puerarin therapy. According to another study, when linked to the non-treated group, puerarin therapy thickened the outer nuclear layer (ONL) and increased the b-wave amplitude. However, in diabetic rats' retinas, puerarin isoflavones increased SOD activity while decreasing AGE buildup, VEGF levels, and STAT3 activity [112].

Puerarin therapy prevented the manifestation of various genes and proteins linked to cell death in retinal cells, including complement 3, nitro-tyrosine (NT), inducible nitric oxide synthase (iNOS), mRNA, and Fas/Fasl [113]. In rat retinal pericytes, Puerarin similarly reduced BSA-induced apoptosis [98]. According to the findings by Zhu *et al.*, Puerarin's therapeutic benefits in RCEs against IL-1 $\beta$ -induced cell dysfunction were also examined. Puerarin suppressed both apoptosis and leukostasis [99]. Cai *et al.* studied the impacts of

Puerarin on diabetic rats. A control group, a low-dose puerarin group (250 mg/kg), and a high-dose puerarin group (500 mg/kg) of rats were randomly assigned. Following a 4-week treatment, malondialdehyde (MDA) and STAT3 levels declined considerably, while SOD activity improved. Puerarin also restored the amplitude of the b-wave in electroretinography [97]. Puerarin's effects on AGE-modified proteins in diabetic rats' blood and retinal tissues were studied by Liu *et al.* Puerarin treatment decreased the amount of AGE-modified proteins while preventing pathogenic changes [114].

Puerarin lowered MDA and ROS levels in NMDA-induced retinal cells, downregulated nNOS expression, and boosted SOD and NO generation. Caspase-3 activity and Bax expression were decreased, and p38 and JNK inhibition were blocked [115]. The buildup of intracellular sugar alcohols has been shown to boost aldose reductase activity in retinopathy patients. 3,6,7,4',5'-pentamethoxy-5,3'-dihydroxyflavone isoflavones isolated from *Caesalpinia pulcherrima* hampered aldose reductase, prolonging the start of DR. It also reduced the production of TBARs and proteins while boosting the action of antioxidant enzymes such as Gpx, C.A.T., and SOD [116].

#### 4.10. Naringenin.

Grapefruit, oranges, and lemons contain the major flavanone components: naringenin, eridictyol, and hesperetin. Furthermore, the chemical structures of such molecules are particularly reactive since they are subjected to o-methylation, hydroxylation, and glycosylation [117]. By reducing GSH and thiobarbituric acid reactive chemical concentrations in STZ-stimulated diabetic mice, Al-Dosari *et al.* discovered that naringenin had antioxidant, antiapoptotic, and neuroprotective properties [101]. Another study looked at the consequences of naringenin on sodium iodate (NaIO<sub>3</sub>)-induced rat retinopathy and ARPE-19 cells treated with NaIO<sub>3</sub>. Carbonyl protein, ROS, and lactic dehydrogenase (LDH) were all discovered. The researchers found that naringenin therapy improved retinal function while lowering Sirtuin 1 protein levels (SIRT1). Carbonyl protein, ROS, and LDH were reduced [118].

#### 4.11. Eriodictyol.

Because it modulates the Nrf2/HO-1 pathway, Eriodictyol possesses anti-inflammatory and antiapoptotic effects. Eriodictyol therapy reduced retinal inflammation in diabetic rats by lowering ICAM-1, eNOS, VEGF, and TNF- $\alpha$  levels. By lowering these mediators' activity, eriodictyol could restore BRB degradation. Eriodictyol [119] reduced ROS production and prevented plasma lipid peroxidation. *In vitro*, eriodictyol has lower ROS concentrations and OS while increasing S.O.D., GPx activity, and C.A.T. Eriodictyol also improved cell survival, nuclear translocation of Nrf2, and the synthesis of heme-oxygenase-1, all of which are significant OS regulators [103].

#### 4.12. Diosmin.

A flavone glycoside in sour fruits, diosmin(30,5,7-trihydroxy-40-methoxy flavone-7-rhamnoglucoside), can protect against cardiovascular disease [120]. Diosmin, the primary ingredient in Daflon nutraceuticals, can treat symptoms of venous insufficiency. This medication can also be used to treat hemorrhoidal crises [121]. Diosmin effectively protected BRB in an ischemic/reperfusion injury paradigm by lowering VEGF representation and

decreasing edema [122]. As a result, diosmin reversed the I/R-induced alterations in retinal ganglion cell count and electroretinogram, which were likewise reversed by diosmin. In the retina, diosmin administration decreased MDA and increased GSH-Px, C.A.T., and SOD activity [123]. Furthermore, diosmin treatment of ARPE-19 cells increased cell survival by lowering apoptosis, reducing ROS production, and increasing SOD and GSH-Px activity [104].

#### *4.13. Chrysin.*

Chrysin protects against visual cycle impairment initiated by hyperglycemia. Chrysin inhibited hyperglycemia-induced neovascularization in retinal pigment epithelium (RPE) cells by lowering IGF-1 and VEGF levels while increasing pigment epithelium-derived factor (PEDF) levels [105]. After Chrysin administration, the external nuclear layer thickness of the db/db mice retina improved, as did the stories of visual cycle-related enzymes (RDH5 and lecithin retinol acyltransferase (LRAT)) [105]. AGE and RAGE induction generation was diminished in a high-glucose RPE cell line treated with chrysin, but RPE65, RDH5, PEDF, and LRAT were elevated. Chrysin also inhibited endoplasmic reticulum stress sensor proteins (ER stress proteins), and the visual cycle was restored by controlling the AGE/RAGE pathway [105].

### **5. Clinical Trials**

Phytochemicals have been shown to protect against DR in various non-clinical investigations. Still, just a few clinical studies have looked into how phytochemicals help protect diabetes and DR. A clinical examination was conducted on 564 patients out of 10,054 to determine the likelihood of having diabetes. The flavonoids myricetin and quercetin were reported to reduce diabetes mellitus [126] dramatically. In addition, a meta-analysis of RCTs with 1584 participants looked at the consequences of catechins, including or not including caffeine, on diabetes biomarkers. Green tea catechins reduced fast blood glucose concentrations considerably however did not affect glycated hemoglobin (HbA1c) or fasting blood insulin levels [124]. The efficacy of oral Melilotus and Centella Asiatica flavonoids in treating diabetic cystoid macular edema was studied in another RCT comprising 70 T2DM patients. According to the study, flavonoids preserved retinal sensitivity but did not influence HbA1c levels, blood pressure, central retinal thickness, visual acuity, and microalbuminuria [125]. Furthermore, fasting glucose and serum adiponectin were significantly lowered and elevated in 160 diabetic individuals after 12 weeks of treatment with pure anthocyanins [126].

Between 2003 and 2006, Mahoney and colleagues used data from the NHANES to consider the effect of a flavonoid-rich diet on DR and diabetes-related biomarkers in 381 diabetic people. A flavonoid-rich food intake reduced the probability of having DR by 30%. They also reduced glucose, C-reactive protein, and hemoglobin A1C levels [127]. Another one-year study indicated that green tea regularly lowered developing DR by 50% [128]. Pycnogenol® has been found to alter the visual function growth in diabetic patients with DR in a multi-center field study. After six months of treatment with Pycnogenol®, no substantial improvement in patient visibility was reported in 1169 individuals [129,130].

Additionally, oxidative stress and central macular thickness were dramatically reduced after six months of Pycnogenol® treatment [131]. In addition, two months of Pycnogenol® treatment alleviated sensory disturbance in patients with DR in the initial phases [130]. Despite the positive restorative consequences of phytochemicals in diabetes, inconsistent or



contradictory data from clinical trials pose considerable barriers. According to reports, most current investigations focus on T2DM patients and patients at various stages of the condition. As a result, linking phytochemicals to the treatment of diabetes has become increasingly challenging [132] (Table 2).

**Table 2.** Clinical studies of polyphenols in the management of diabetic retinopathy (DR).

Bioactive compound/ Plant	Dose/Concentration	Duration	Participants	Outcomes	Refs
Chinese green tea	Ingested once a week	One year	100 subjects with DR and 100 diabetic subjects without DR.	DR incidence was reduced by 50% and associated with those not drinking green tea.	[128]
Catechins (Green tea)	-	-	1584 participants	FBG levels are lower; HbA1c and fasting blood insulin levels are unaffected.	[124]
Myricetin and quercetin	-	One year	The possibility of developing diabetes was investigated in 564 of 10,054 participants.	Diminished the possibility of improving T2DM	[133]
Pycnogenol®	60–120 mg, daily	Six months	1169 diabetic subjects having DR.	There was no discernible improvement in the patient's visibility due to the treatment.	[134]
Pycnogenol®	150 mg/day	Two months	46 subjects	Enhancement in appearance and baseline	[130]
Purified anthocyanins	320 mg, daily	12 weeks	160 subjects	Fasting glucose levels were lower, and serum adiponectin levels were higher.	[126]
Melilotus and <i>Centella Asiatica</i> flavonoids	Oral administration of 15 mg <i>C. asiatica</i> , 300 mg diosminand 160 mg Melilotus, daily	36 months	70 subjects	Blood pressure, visual acuity, and microalbuminuria did not change significantly.	[125]
Flavonoid-rich diet	-	-	381 diabetic patient	Lowered glucose, HbA1C, and C-reactive protein levels by 30%, reducing the progression of DR.	[127]

## 5. Concluding Remarks and Future Perspectives

Polyphenols can treat DR as a secure and effective substitute for current medications and treatments. Numerous studies in animals and cell cultures have demonstrated that phytochemicals can reduce neurodegeneration and programmed cell death *in vivo* and *in vitro*, improve the oxidative state, lower the construction of pro-inflammatory mediators, and restore retina thickness. Because of the presence of polyphenols, the medicinal impacts of their intervention have been well-established in various clinical studies. According to a few studies, polyphenol supplementation with a balanced diet can help people with diabetes avoid diabetes and associated consequences, such as DR. Another advantage of consuming polyphenols is that it helps prevent DR. Results from preclinical and clinical research indicated that including polyphenols in one's diet can help avoid the development of DR and other vision problems. Instead of treating nutraceuticals as magic bullets that may cure people of their ailments, they should augment a healthy diet.

Additional experimental trials are needed to demonstrate the purpose and relevance of polyphenols in DR. Because the outcomes of some clinical trials have been varied, it is impossible to say whether natural substances have a consistently positive effect on patients. As a result, drawing appropriate conclusions from these investigations takes time and effort. The bioavailability of most polyphenols and other nutraceuticals, such as resveratrol, curcumin, and others, is low, making developing a therapeutic molecule difficult.

Furthermore, the absence of readily available delivery systems for nutraceutical compounds is another possible flaw that must be addressed soon. The retina has only been the subject of many clinical studies due to the nutraceuticals' restrictive delivery paradigm. On the other hand, nanotechnology has substantially improved natural chemical delivery mechanisms through novel technologies. To cure DR more successfully, future research should focus on reducing the barriers limiting nutraceutical medicinal molecules' creation.

## Funding

The authors did not receive any external funding.

## Acknowledgments

None.

## Conflicts of Interest

The authors declare no conflict of interest.

## References

1. Harikumar, K.; Hemalatha, G.J.; Kumar, B.; Lado, S.F.S. A review on diabetes mellitus. *Int. J. Nov. trends Pharm. Sci.* **2014**, *4*, 201–217, <https://scienztech.org/index.php/ijntps/article/view/139>.
2. Ting, D.S.W.; Cheung, G.C.M.; Wong, T.Y. Diabetic retinopathy: global prevalence, major risk factors, screening practices and public health challenges: a review. *Clin. Exp. Ophthalmol.* **2016**, *44*, 260–277, <https://doi.org/10.1111/ceo.12696>.
3. E. Chepchumba Yego, D.J.F. Modes of Retinal Cell Death in Diabetic Retinopathy. *J. Clin. Exp. Ophthalmol.* **2013**, *04*, <https://doi.org/10.4172/2155-9570.1000298>.
4. Calderon, G.D.; Juarez, O.H.; Hernandez, G.E.; Punzo, S.M.; De La Cruz, Z.D. Oxidative stress and diabetic retinopathy: Development and treatment. *Eye* **2017**, *31*, 1122–1130, <https://doi.org/10.1038/eye.2017.64>.
5. Safi, S.Z.; Qvist, R.; Kumar, S.; Batumalaie, K.; Ismail, I.S. Bin Molecular mechanisms of diabetic retinopathy, general preventive strategies, and novel therapeutic targets. *Biomed Res. Int.* **2014**, *2014*, <https://doi.org/10.1155/2014/801269>.
6. Li, C.; Miao, X.; Li, F.; Wang, S.; Liu, Q.; Wang, Y.; Sun, J. Oxidative Stress-Related Mechanisms and Antioxidant Therapy in Diabetic Retinopathy. *Oxid. Med. Cell. Longev.* **2017**, *2017*, <https://doi.org/10.1155/2017/9702820>.
7. Alzamel, N.M. Bioactive Compounds in some Medicinal Plants from Different Habitats in KSA. *Pakistan J. Med. Heal. Sci.* **2022**, *16*, 1085, <https://doi.org/10.53350/pjmhs221621085>.
8. Jeetah, R.; Bhaw-Luximon, A.; Jhurry, D. Nanopharmaceutics: Phytochemical-based controlled or sustained drug-delivery systems for cancer treatment. *J. Biomed. Nanotechnol.* **2014**, *10*, 1810–1840, <https://doi.org/10.1166/jbn.2014.1884>.
9. Dewan, N.; Chowdhary, G.; Pandit, S.; Ahmed, P.; Dasgupta, D. Review on-Herbosomes, A new arena for drug delivery. *J. Pharmacogn. Phytochem.* **2016**, *5*, 104–108, <https://www.phytojournal.com/archives/2016/vol5issue4/PartB/5-3-8-286.pdf>.
10. Jugran, A.K.; Rawat, S.; Devkota, H.P.; Bhatt, I.D.; Rawal, R.S. Diabetes and plant-derived natural products: From ethnopharmacological approaches to their potential for modern drug discovery and development. *Phyther. Res.* **2021**, *35*, 223–245, <https://doi.org/10.1002/ptr.6821>.
11. Anwar, S.; Khan, S.; Almatroudi, A.; Khan, A.A.; Alsahli, M.A.; Almatroodi, S.A.; Rahmani, A.H. A review on mechanism of inhibition of advanced glycation end products formation by plant derived polyphenolic compounds. *Mol. Biol. Rep.* **2021**, *48*, 787–805, <https://doi.org/10.1007/s11033-020-06084-0>.
12. Bourne, R.R.A.; Stevens, G.A.; White, R.A.; Smith, J.L.; Flaxman, S.R.; Price, H.; Jonas, J.B.; Keeffe, J.; <https://biointerfaceresearch.com/>

- Leasher, J.; Naidoo, K.; *et al.* Causes of vision loss worldwide, 1990–2010: A systematic analysis. *Lancet Glob. Heal.* **2013**, *1*, e339–e349, <https://pubmed.ncbi.nlm.nih.gov/25104599/>.
13. Flaxman, S.R.; Bourne, R.R.A.; Resnikoff, S.; Ackland, P.; Braithwaite, T.; Cicinelli, M. V.; Das, A.; Jonas, J.B.; Keeffe, J.; Kempen, J.; *et al.* Global causes of blindness and distance vision impairment 1990–2020: a systematic review and meta-analysis. *Lancet Glob. Heal.* **2017**, *5*, e1221–e1234, [https://doi.org/10.1016/S2214-109X\(17\)30393-5](https://doi.org/10.1016/S2214-109X(17)30393-5).
  14. Rodríguez, M.L.; Pérez, S.; Mena-Mollá, S.; Desco, M.C.; Ortega, Á.L. Oxidative Stress and Microvascular Alterations in Diabetic Retinopathy: Future Therapies. *Oxid. Med. Cell. Longev.* **2019**, *2019*, <https://doi.org/10.1155/2019/4940825>.
  15. Yau, J.W.Y.; Rogers, S.L.; Kawasaki, R.; Lamoureux, E.L.; Kowalski, J.W.; Bek, T.; Chen, S.J.; Dekker, J.M.; Fletcher, A.; Grauslund, J.; *et al.* Global prevalence and major risk factors of diabetic retinopathy. *Diabetes Care* **2012**, *35*, 556–564, <https://doi.org/10.2337/dc11-1909>.
  16. Romero-Aroca, P.; Navarro-Gil, R.; Valls-Mateu, A.; Sagarra-Alamo, R.; Moreno-Ribas, A.; Soler, N. Differences in incidence of diabetic retinopathy between type 1 and 2 diabetes mellitus: A nine-year follow-up study. *Br. J. Ophthalmol.* **2017**, *101*, 1346–1351, <https://doi.org/10.1136/bjophthalmol-2016-310063>.
  17. Prasathkumar, M.; Becky, R.; Anisha, S.; Dhriya, C.; Sadhasivam, S. Evaluation of hypoglycemic therapeutics and nutritional supplementation for type 2 diabetes mellitus management: An insight on molecular approaches. *Biotechnol. Lett.* **2022**, 1–36, <https://doi.org/10.1007/s10529-022-03232-3>.
  18. Ansari, P.; Tabasumma, N.; Snigdha, NN; Siam, NH; Panduru, R.V.; Azam, S.; Hannan, J.M.A.; Abdel-Wahab, Y.H.A. Diabetic Retinopathy: An Overview on Mechanisms, Pathophysiology and Pharmacotherapy. *Diabetology* **2022**, *3*, 159–175, <https://doi.org/10.3390/diabetology3010011>.
  19. Cantó, A.; Martínez, J.; Perini-Villanueva, G.; Miranda, M.; Bejarano, E. Early Neural Changes as Underlying Pathophysiological Mechanism in Diabetic Retinopathy. *Int. J. Transl. Med.* **2021**, *2*, 1–16, <https://doi.org/10.3390/ijtm2010001>.
  20. Nibali, L.; Gkraniyas, N.; Mainas, G.; Di Pino, A. Periodontitis and implant complications in diabetes. *Periodontol. 2000* **2022**, *90*, 88–105, <https://doi.org/10.1111/prd.12451>.
  21. Brownlee, M. The pathobiology of diabetic complications: A unifying mechanism. *Diabetes* **2005**, *54*, 1615–1625, <https://doi.org/10.2337/diabetes.54.6.1615>.
  22. Kowluru, R.A.; Chan, P.S. Oxidative stress and diabetic retinopathy. *Exp. Diabetes Res.* **2007**, *2007*, <https://doi.org/10.1155/2007/43603>.
  23. Wu, M.Y.; Yang, G.T.; Lai, T.T.; Li, C.J. The oxidative stress and mitochondrial dysfunction during the pathogenesis of diabetic retinopathy. *Oxid. Med. Cell. Longev.* **2018**, *2018*, <https://doi.org/10.1155/2018/3420187>.
  24. Wang, Z.; Zhao, H.; Guan, W.; Kang, X.; Tai, X.; Shen, Y. Metabolic memory in mitochondrial oxidative damage triggers diabetic retinopathy. *BMC Ophthalmol.* **2018**, *18*, <https://doi.org/10.1186/s12886-018-0921-0>.
  25. Alfonso-Muñoz, E.A.; Burggraaf-Sánchez de las Matas, R.; Mataix Boronat, J.; Molina Martín, JC; Desco, C. Role of oral antioxidant supplementation in the current management of diabetic retinopathy. *Int. J. Mol. Sci.* **2021**, *22*, 4020, <https://doi.org/10.3390/ijms22084020>.
  26. Pal, S.; Rao, G.N.; Pal, A. Inflammation and apoptosis, two key events induced by hyperglycemia mediated reactive nitrogen species in RGC-5 cells. *Life Sci.* **2021**, *279*, 119693, <https://doi.org/10.1016/j.lfs.2021.119693>.
  27. Altmann, C.; Schmidt, M.H.H. The role of microglia in diabetic retinopathy: Inflammation, microvasculature defects and neurodegeneration. *Int. J. Mol. Sci.* **2018**, *19*, 110, <https://doi.org/10.3390/ijms19010110>.
  28. Vujosevic, S.; Micera, A.; Bini, S.; Berton, M.; Esposito, G.; Midena, E. Aqueous humor biomarkers of müller cell activation in diabetic eyes. *Investig. Ophthalmol. Vis. Sci.* **2015**, *56*, 3913–3918, <https://doi.org/10.1167/iovs.15-16554>.
  29. Simó-Servat, O.; Hernández, C.; Simó, R. Usefulness of the vitreous fluid analysis in the translational research of diabetic retinopathy. *Mediators Inflamm.* **2012**, *2012*, <https://doi.org/10.1155/2012/872978>.
  30. Murugeswari, P.; Shukla, D.; Rajendran, A.; Kim, R.; Namperumalsamy, P.; Muthukkaruppan, V. Pro-inflammatory cytokines and angiogenic and anti-angiogenic factors in vitreous of patients with proliferative diabetic retinopathy and eales' disease. *Retina* **2008**, *28*, 817–824, <https://doi.org/10.1097/IAE.0b013e31816576d5>.
  31. Koleva-Georgieva, D.N.; Sivkova, N.P.; Terzieva, D. Serum inflammatory cytokines IL-1beta, IL-6, TNF-alpha and VEGF have influence on the development of diabetic retinopathy. *Folia Med. (Plovdiv)*. **2011**, *53*, 44–50, <https://doi.org/10.2478/v10153-010-0036-8>.
  32. Giblin, M.J.; Ontko, C.D.; Penn, J.S. Effect of cytokine-induced alterations in extracellular matrix composition on diabetic retinopathy-relevant endothelial cell behaviors. *Sci. Rep.* **2022**, *12*, 1–14, <https://www.nature.com/articles/s41598-022-12683-7>.
  33. Kaabi, Y.A. Potential Roles of Anti-Inflammatory Plant-Derived Bioactive Compounds Targeting Inflammation in Microvascular Complications of Diabetes. *Molecules* **2022**, *27*, 7352, <https://doi.org/10.3390/molecules27217352>.

34. Blum, A.; Pastukh, N.; Socea, D.; Jabaly, H. Levels of adhesion molecules in peripheral blood correlat with stages of diabetic retinopathy and may serve as bio markers for microvascular complications. *Cytokine* **2018**, *106*, 76–79, <https://doi.org/10.1016/j.cyto.2017.10.014>.
35. Little, K.; Llorián-Salvador, M.; Scullion, S.; Hernández, C.; Simó-Servat, O.; Del Marco, A.; Bosma, E.; Vargas-Soria, M.; Carranza-Naval, M.J.; Van Bergen, T. Common pathways in dementia and diabetic retinopathy: Understanding the mechanisms of diabetes-related cognitive decline. *Trends Endocrinol. Metab.* **2022**, *33*, 50–71, <https://doi.org/10.1016/j.tem.2021.10.008>.
36. Lechner, J.; Medina, R.J.; Lois, N.; Stitt, A.W. Advances in cell therapies using stem cells/progenitors as a novel approach for neurovascular repair of the diabetic retina. *Stem Cell Res. Ther.* **2022**, *13*, 1–16, <https://stemcellres.biomedcentral.com/articles/10.1186/s13287-022-03073-x>.
37. Sasaki, M.; Ozawa, Y.; Kurihara, T.; Kubota, S.; Yuki, K.; Noda, K.; Kobayashi, S.; Ishida, S.; Tsubota, K. Neurodegenerative influence of oxidative stress in the retina of a murine model of diabetes. *Diabetologia* **2010**, *53*, 971–979, <https://doi.org/10.1007/s00125-009-1655-6>.
38. Mrugacz, M.; Bryl, A.; Zorena, K. Retinal vascular endothelial cell dysfunction and neuroretinal degeneration in diabetic patients. *J. Clin. Med.* **2021**, *10*, 458, <https://doi.org/10.3390/jcm10030458>.
39. Durham, J.T.; Herman, I.M. Microvascular modifications in diabetic retinopathy. *Curr. Diab. Rep.* **2011**, *11*, 253–264, <https://doi.org/10.1007/s11892-011-0204-0>.
40. Miyamoto, K.; Khosrof, S.; Bursell, S.E.; Rohan, R.; Murata, T.; Clermont, A.C.; Aiello, L.P.; Ogura, Y.; Adamis, A.P. Prevention of leukostasis and vascular leakage in streptozotocin-induced diabetic retinopathy via intercellular adhesion molecule-1 inhibition. *Proc. Natl. Acad. Sci. U. S. A.* **1999**, *96*, 10836–10841, <https://doi.org/10.1073/pnas.96.19.10836>.
41. Zhang, D.; Lv, F.L.; Wang, G.H. Effects of HIF-1 $\alpha$  on diabetic retinopathy angiogenesis and VEGF expression. *Eur. Rev. Med. Pharmacol. Sci.* **2018**, *22*, 5071–5076, [https://doi.org/10.26355/eurrev\\_201808\\_15699](https://doi.org/10.26355/eurrev_201808_15699).
42. Anand, S.; Sowbhagya, R.; Ansari, M.A.; Alzohairy, M.A.; Alomary, M.N.; Almalik, A.I.; Ahmad, W.; Tripathi, T.; Elderderly, A.Y. Polyphenols and Their Nanoformulations: Protective Effects against Human Diseases. *Life* **2022**, *12*, 1639, <https://doi.org/10.3390/life12101639>.
43. Rothwell, J.A.; Perez-Jimenez, J.; Neveu, V.; Medina-Remón, A.; M'Hiri, N.; García-Lobato, P.; Manach, C.; Knox, C.; Eisner, R.; Wishart, D.S.; et al. Phenol-Explorer 3.0: A major update of the Phenol-Explorer database to incorporate data on the effects of food processing on polyphenol content. *Database* **2013**, *2013*, <https://doi.org/10.1093/database/bat070>.
44. Neveu, V.; Perez-Jiménez, J.; Vos, F.; Crespy, V.; du Chaffaut, L.; Mennen, L.; Knox, C.; Eisner, R.; Cruz, J.; Wishart, D.; et al. Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. *Database (Oxford)*. **2010**, *2010*, <https://doi.org/10.1093/database/bap024>.
45. Grosso, G.; Godos, J.; Currenti, W.; Micek, A.; Falzone, L.; Libra, M.; Giampieri, F.; Forbes-Hernández, T.Y.; Quiles, J.L.; Battino, M. The effect of dietary polyphenols on vascular health and hypertension: Current evidence and mechanisms of action. *Nutrients* **2022**, *14*, 545, <https://doi.org/10.3390/nu14030545>.
46. Zhang, L.-X.; Li, C.-X.; Kakar, M.U.; Khan, M.S.; Wu, P.-F.; Amir, R.M.; Dai, D.-F.; Naveed, M.; Li, Q.-Y.; Saeed, M. Resveratrol (RV): A pharmacological review and call for further research. *Biomed. Pharmacother.* **2021**, *143*, 112164, <https://doi.org/10.1016/j.biopha.2021.112164>.
47. Abu-Amero, K.K.; Kondkar, A.A.; Chalam, K. V. Resveratrol and ophthalmic diseases. *Nutrients* **2016**, *8*, <https://doi.org/10.3390/nu8040200>.
48. Goldberg, D.M.; Yan, J.; Soleas, G.J. Absorption of three wine-related polyphenols in three different matrices by healthy subjects. *Clin. Biochem.* **2003**, *36*, 79–87, [https://doi.org/10.1016/S0009-9120\(02\)00397-1](https://doi.org/10.1016/S0009-9120(02)00397-1).
49. Walle, T. Bioavailability of resveratrol. *Ann. N. Y. Acad. Sci.* **2011**, *1215*, 9–15, <https://doi.org/10.1111/j.1749-6632.2010.05842.x>.
50. Doganay, S.; Borazan, M.; Iraz, M.; Cigremis, Y. The effect of resveratrol in experimental cataract model formed by sodium selenite. *Curr. Eye Res.* **2006**, *31*, 147–153, <https://doi.org/10.1080/02713680500514685>.
51. Hightower, K.R.; McCready, J.P. Effect of selenite on epithelium of cultured rabbit lens. *Investig. Ophthalmol. Vis. Sci.* **1991**, *32*, 406–409, <https://pubmed.ncbi.nlm.nih.gov/1825204/>.
52. Bola, C.; Bartlett, H.; Eperjesi, F. Resveratrol and the eye: Activity and molecular mechanisms. *Graefe's Arch. Clin. Exp. Ophthalmol.* **2014**, *252*, 699–713, <https://doi.org/10.1007/s00417-014-2604-8>.
53. Kim, Y.H.; Kim, Y.S.; Roh, G.S.; Choi, W.S.; Cho, G.J. Resveratrol blocks diabetes-induced early vascular lesions and vascular endothelial growth factor induction in mouse retinas. *Acta Ophthalmol.* **2012**, *90*, <https://doi.org/10.1111/j.1755-3768.2011.02243.x>.
54. Luna, C.; Li, G.; Liton, P.B.; Qiu, J.; Epstein, D.L.; Challa, P.; Gonzalez, P. Resveratrol prevents the expression of glaucoma markers induced by chronic oxidative stress in trabecular meshwork cells. *Food Chem. Toxicol.* **2009**, *47*, 198–204, <https://doi.org/10.1016/j.fct.2008.10.029>.
55. Zhang, H.A.; Kitts, D.D. Turmeric and its bioactive constituents trigger cell signaling mechanisms that protect against diabetes and cardiovascular diseases. *Mol. Cell. Biochem.* **2021**, *476*, 3785–3814, <https://doi.org/10.1007/s11010-021-04201-6>.
56. E.C. on F.A. Evaluation of certain food additives. *World Heal. Organ. - Tech. Rep. Ser.* **1975**, 576. Available



online:

[https://apps.who.int/iris/bitstream/handle/10665/44062/WHO\\_TRS\\_952\\_eng.pdf;jsessionid=330AB127F5D0762ADEC6FE3922D7B82E?sequence=1](https://apps.who.int/iris/bitstream/handle/10665/44062/WHO_TRS_952_eng.pdf;jsessionid=330AB127F5D0762ADEC6FE3922D7B82E?sequence=1).

57. Priyadarsini, K.I.; Maity, D.K.; Naik, G.H.; Kumar, M.S.; Unnikrishnan, MK; Satav, J.G.; Mohan, H. Role of phenolic O-H and methylene hydrogen on the free radical reactions and antioxidant activity of curcumin. *Free Radic. Biol. Med.* **2003**, *35*, 475–484, [https://doi.org/10.1016/S0891-5849\(03\)00325-3](https://doi.org/10.1016/S0891-5849(03)00325-3).
58. Pinlaor, S.; Yongvanit, P.; Prakobwong, S.; Kaewsamut, B.; Khoontawad, J.; Pinlaor, P.; Hiraku, Y. Curcumin reduces oxidative and nitrate DNA damage through balancing of oxidant-antioxidant status in hamsters infected with *Opisthorchis viverrini*. *Mol. Nutr. Food Res.* **2009**, *53*, 1316–1328, <https://doi.org/10.1002/mnfr.200800567>.
59. Molina-Jijón, E.; Tapia, E.; Zazueta, C.; El Hafidi, M.; Zatarain-Barrón, Z.L.; Hernández-Pando, R.; Medina-Campos, O.N.; Zarco-Márquez, G.; Torres, I.; Pedraza-Chaverri, J. Curcumin prevents Cr(VI)-induced renal oxidant damage by a mitochondrial pathway. *Free Radic. Biol. Med.* **2011**, *51*, 1543–1557, <https://doi.org/10.1016/j.freeradbiomed.2011.07.018>.
60. Kowluru, R.A.; Kanwar, M. Effects of curcumin on retinal oxidative stress and inflammation in diabetes. *Nutr. Metab.* **2007**, *4*, <https://doi.org/10.1186/1743-7075-4-8>.
61. Gupta, S.K.; Kumar, B.; Nag, T.C.; Agrawal, S.S.; Agrawal, R.; Agrawal, P.; Saxena, R.; Srivastava, S. Curcumin prevents experimental diabetic retinopathy in rats through its hypoglycemic, antioxidant, and anti-inflammatory mechanisms. *J. Ocul. Pharmacol. Ther.* **2011**, *27*, 123–130, <https://doi.org/10.1089/JOP.2010.0123>.
62. Flynn, D.L.; Rafferty, M.F.; Boctor, A.M. Inhibition of 5-hydroxy-eicosatetraenoic acid (5-HETE) formation in intact human neutrophils by naturally-occurring diarylheptanoids: inhibitory activities of curcuminoids and yakuchinones. *Prostaglandins, Leukot. Med.* **1986**, *22*, 357–360, [https://doi.org/10.1016/0262-1746\(86\)90146-0](https://doi.org/10.1016/0262-1746(86)90146-0).
63. Sameermahmood, Z.; Balasubramanyam, M.; Saravanan, T.; Rema, M. Curcumin modulates SDF-1 $\alpha$ /CXCR4-induced migration of human retinal endothelial cells (HRECs). *Investig. Ophthalmol. Vis. Sci.* **2008**, *49*, 3305–3311, <https://doi.org/10.1167/iovs.07-0456>.
64. Li, D.L.; Mao, L.; Gu, Q.; Wei, F.; Gong, Y.-Y. Quercetin protects retina external barrier from oxidative stress injury by promoting autophagy. *Cutan. Ocul. Toxicol.* **2021**, *40*, 7–13, <https://doi.org/10.1080/15569527.2020.1860082>.
65. Chai, G.R.; Liu, S.; Yang, H.W.; Chen, X.L. Quercetin protects against diabetic retinopathy in rats by inducing heme oxygenase-1 expression. *Neural Regen. Res.* **2021**, *16*, 1344–1350, <https://doi.org/10.4103/1673-5374.301027>.
66. Tang, W.; Guo, J.; Gu, R.; Lei, B.; Ding, X.; Ma, J.; Xu, G. MicroRNA-29b-3p inhibits cell proliferation and angiogenesis by targeting VEGFA and PDGFB in retinal microvascular endothelial cells. *Mol. Vis.* **2020**, *26*, 64–75, <https://pubmed.ncbi.nlm.nih.gov/32165827/>.
67. Wang, X.; Li, H.; Wang, H.; Shi, J. Retraction: Paper withdrawn: Quercetin attenuates high glucose-induced injury in human retinal pigment epithelial cell line ARPE-19 by up-regulation of miR-29b. <https://doi.org/10.1093/jb/mvaa001>. *J. Biochem.* **2020**, *168*, 677, <https://doi.org/10.1093/jb/mvaa096>.
68. Kumar, B.; Gupta, S.K.; Nag, T.C.; Srivastava, S.; Saxena, R.; Jha, K.A.; Srinivasan, B.P. Retinal neuroprotective effects of quercetin in streptozotocin-induced diabetic rats. *Exp. Eye Res.* **2014**, *125*, 193–202, <https://doi.org/10.1016/j.exer.2014.06.009>.
69. Chen, B.; He, T.; Xing, Y.; Cao, T. Effects of quercetin on the expression of MCP-1, MMP-9 and VEGF in rats with diabetic retinopathy. *Exp. Ther. Med.* **2017**, *14*, 6022–6026, <https://doi.org/10.3892/etm.2017.5275>.
70. Rudra, A.; Arvind, I.; Mehra, R. Polyphenols: types, sources and therapeutic applications. *Int J Home Sci* **2021**, *7*, 69–75, <https://www.homesciencejournal.com/archives/2021/vol7issue3/PartB/7-2-68-221.pdf>.
71. Bikbova, G.; Oshitari, T.; Baba, T.; Yamamoto, S. Mechanisms of Neuronal Cell Death in AGE-exposed Retinas - Research and Literature Review. *Curr. Diabetes Rev.* **2017**, *13*, 280–288, <https://doi.org/10.2174/1573399812666160519111333>.
72. Kim, J.; Kim, C.S.; Moon, M.K.; Kim, J.S. Epicatechin breaks preformed glycated serum albumin and reverses the retinal accumulation of advanced glycation end products. *Eur. J. Pharmacol.* **2015**, *748*, 108–114, <https://doi.org/10.1016/J.EJP.2014.12.010>.
73. Skopinski, P.; Szaflik, J.; Duda-Król, B.; Nartowska, J.; Sommer, E.; Chorostowska-Wynimko, J.; Demkow, U.; Skopinska-Rózewska, E. Suppression of angiogenic activity of sera from diabetic patients with non-proliferative retinopathy by compounds of herbal origin and sulindac sulfone. *Int. J. Mol. Med.* **2004**, *14*, 707–711, <https://pubmed.ncbi.nlm.nih.gov/15375605/>.
74. Kodama, D.H.; Gonçalves, A.E. de SS; Lajolo, F.M.; Genovese, M.I. Flavonoids, total phenolics and antioxidant capacity: comparison between commercial green tea preparations. *Ciência e Tecnol. Aliment.* **2010**, *30*, 1077–1082, <https://doi.org/10.1590/s0101-20612010000400037>.
75. Silva, K.C.; Rosales, M.A.B.; Hamassaki, D.E.; Saito, K.C.; Faria, A.M.; Ribeiro, P.A.O.; Lopes de Faria, J.B.; Lopes de Faria, J.M. Green tea is neuroprotective in diabetic retinopathy. *Invest. Ophthalmol. Vis. Sci.* **2013**, *54*, 1325–1336, <https://doi.org/10.1167/IOVS.12-10647>.



76. Zhang, L.; Zhang, Z.K.; Liang, S. Epigallocatechin-3-gallate protects retinal vascular endothelial cells from high glucose stress in vitro via the MAPK/ERK-VEGF pathway. *Genet. Mol. Res.* **2016**, *15*, <https://doi.org/10.4238/GMR.15027874>.
77. Lee, H.S.; Jun, J.H.; Jung, E.H.; Koo, B.A.; Kim, Y.S. Epigallocatechin-3-gallate inhibits ocular neovascularization and vascular permeability in human retinal pigment epithelial and human retinal microvascular endothelial cells via suppression of MMP-9 and VEGF activation. *Molecules* **2014**, *19*, 12150–12172, <https://doi.org/10.3390/MOLECULES190812150>.
78. Wang, L.; Sun, X.; Zhu, M.; Du, J.; Xu, J.; Qin, X.; Xu, X.; Song, E. Epigallocatechin-3-gallate stimulates autophagy and reduces apoptosis levels in retinal Müller cells under high-glucose conditions. *Exp. Cell Res.* **2019**, *380*, 149–158, <https://doi.org/10.1016/J.YEXCR.2019.04.014>.
79. Ola, M.S.; Ahmed, M.M.; Ahmad, R.; Abuhashish, H.M.; Al-Rejaie, S.S.; Alhomida, A.S. Neuroprotective Effects of Rutin in Streptozotocin-Induced Diabetic Rat Retina. **2015**, *56*, 440–448, <https://doi.org/10.1007/s12031-015-0561-2>.
80. Gupta, S.K.; Sharma, H.P.; Das, U.; Velpandian, T.; Saklani, R. Effect of rutin on retinal VEGF, TNF- $\alpha$ , aldose reductase, and total antioxidant capacity in diabetic rats: molecular mechanism and ocular pharmacokinetics. *International Ophthalmology* **2020**, *40*, 159–168, <https://doi.org/10.1007/s10792-019-01165-x>.
81. Zhang, T.; Mei, X.; Ouyang, H.; Lu, B.; Yu, Z.; Wang, Z.; Ji, L. Natural flavonoid galangin alleviates microglia-triggered blood–retinal barrier dysfunction during the development of diabetic retinopathy. *J. Nutr. Biochem.* **2019**, *65*, 1–14, <https://doi.org/10.1016/j.jnutbio.2018.11.006>.
82. Xu, X.H.; Zhao, C.; Peng, Q.; Xie, P.; Liu, Q.H. Kaempferol inhibited VEGF and PGF expression and in vitro angiogenesis of hrecs under diabetic-like environment. *Brazilian J. Med. Biol. Res.* **2017**, *50*, <https://doi.org/10.1590/1414-431X20165396>.
83. Hua, F.; Zhou, P.; Wu, H.Y.; Chu, G.X.; Xie, Z.W.; Bao, G.H. Inhibition of  $\alpha$ -glucosidase and  $\alpha$ -amylase by flavonoid glycosides from Lu'an GuaPian tea: molecular docking and interaction mechanism. *Food Funct.* **2018**, *9*, 4173–4183, <https://doi.org/10.1039/c8fo00562a>.
84. Zhao, L.; Sun, J.; Shi, S.; Qin, X.; Zhang, K.; Xu, J. Kaempferol protects retinal ganglion cells from high-glucose-induced injury by regulating vasohibin-1. *Neuroscience Letters* **2020**, *716*, <https://doi.org/10.1016/j.neulet.2019.134633>.
85. Soufi, F.G.; Mohammad-nejad, D.; Ahmadi, H. Resveratrol improves diabetic retinopathy possibly through oxidative stress - nuclear factor  $\kappa$ B - apoptosis pathway. *Pharmacol. Rep.* **2012**, *64*, 1505–1514, [https://doi.org/10.1016/S1734-1140\(12\)70948-9](https://doi.org/10.1016/S1734-1140(12)70948-9).
86. Sohn, E.; Kim, J.; Kim, C.S.; Lee, Y.M.; Kim, J.S. Extract of *Polygonum cuspidatum* Attenuates Diabetic Retinopathy by Inhibiting the High-Mobility Group Box-1 (HMGB1) Signaling Pathway in Streptozotocin-Induced Diabetic Rats. *Nutrients* **2016**, *8*, <https://doi.org/10.3390/NU8030140>.
87. Chen, Y.; Meng, J.; Li, H.; Wei, H.; Bi, F.; Liu, S.; Tang, K.; Guo, H.; Liu, W. Resveratrol exhibits an effect on attenuating retina inflammatory condition and damage of diabetic retinopathy via PON1. *Exp. Eye Res.* **2019**, *181*, 356–366, <https://doi.org/10.1016/J.EXER.2018.11.023>.
88. Kim, Y.H.; Kim, Y.S.; Kang, S.S.; Cho, G.J.; Choi, W.S. Resveratrol inhibits neuronal apoptosis and elevated Ca<sup>2+</sup>/calmodulin-dependent protein kinase II activity in diabetic mouse retina. *Diabetes* **2010**, *59*, 1825–1835, <https://doi.org/10.2337/DB09-1431>.
89. Yang, F.; Yu, J.; Ke, F.; Lan, M.; Li, D.; Tan, K.; Ling, J.; Wang, Y.; Wu, K.; Li, D. Curcumin Alleviates Diabetic Retinopathy in Experimental Diabetic Rats. *Ophthalmic Res.* **2018**, *60*, 43–54, <https://doi.org/10.1159/000486574>.
90. Zuo, Z.F.; Zhang, Q.; Liu, X.Z. Protective effects of curcumin on retinal Müller cell in early diabetic rats. *Int. J. Ophthalmol.* **2013**, *6*, 422–424, <https://doi.org/10.3980/J.ISSN.2222-3959.2013.04.02>.
91. Li, R.; Chen, L.; Yao, G.M.; Yan, H.L.; Wang, L. Effects of quercetin on diabetic retinopathy and its association with NLRP3 inflammasome and autophagy. *Int. J. Ophthalmol.* **2021**, *14*, 42–49, <https://doi.org/10.18240/IJO.2021.01.06>.
92. Wang, X.; Li, H.; Wang, H.; Shi, J. Quercetin attenuates high glucose-induced injury in human retinal pigment epithelial cell line ARPE-19 by up-regulation of miR-29b. *J. Biochem.* **2020**, *167*, 495–502, <https://doi.org/10.1093/JB/MVAA001>.
93. Wang, W.; Zhang, Y.; Jin, W.; Xing, Y.; Yang, A. Catechin weakens diabetic retinopathy by inhibiting the expression of nf- $\kappa$ b signaling pathway-mediated inflammatory factors. *Ann. Clin Lab Sci* **2018**, *48*, 594–600, <https://pubmed.ncbi.nlm.nih.gov/30373863/>.
94. Attoub, S.; Hassan, A.H.; Vanhoecke, B.; Iratni, R.; Takahashi, T.; Gaben, A.M.; Bracke, M.; Awad, S.; John, A.; Kamalboor, H.A.; *et al.* Inhibition of cell survival, invasion, tumor growth and histone deacetylase activity by the dietary flavonoid luteolin in human epithelioid cancer cells. *Eur. J. Pharmacol.* **2011**, *651*, 18–25, <https://doi.org/10.1016/J.EJPHAR.2010.10.063>.
95. Zhang, T.; Mei, X.; Ouyang, H.; Lu, B.; Yu, Z.; Wang, Z.; Ji, L. Natural flavonoid galangin alleviates microglia-triggered blood-retinal barrier dysfunction during the development of diabetic retinopathy. *J. Nutr. Biochem.* **2019**, *65*, 1–14, <https://doi.org/10.1016/J.JNUTBIO.2018.11.006>.
96. Xu, X.H.; Zhao, C.; Peng, Q.; Xie, P.; Liu, Q.H. Kaempferol inhibited VEGF and PGF expression and in

- vitro angiogenesis of HRECs under diabetic-like environment. *Brazilian J. Med. Biol. Res. = Rev. Bras. Pesqui. medicas e Biol.* **2017**, *50*, <https://doi.org/10.1590/1414-431X20165396>.
97. Cai, Y.; Zhang, X.; Xu, X.; Yu, Y. Effects of Puerarin on the retina and STAT3 expression in diabetic rats. *Exp. Ther. Med.* **2017**, *14*, 5480–5484, <https://doi.org/10.3892/ETM.2017.5203>.
98. Kim, J.; Kim, K.M.; Kim, C.S.; Sohn, E.; Lee, Y.M.; Jo, K.; Kim, J.S. Puerarin inhibits the retinal pericyte apoptosis induced by advanced glycation end products in vitro and in vivo by inhibiting NADPH oxidase-related oxidative stress. *Free Radic. Biol. Med.* **2012**, *53*, 357–365, <https://doi.org/10.1016/J.FREERADBIOMED.2012.04.030>.
99. Zhu, X.; Xie, M.; Wang, K.; Zhang, K.; Gao, Y.; Zhu, L.; Zhou, F. The effect of Puerarin against IL-1 $\beta$ -mediated leukostasis and apoptosis in retinal capillary endothelial cells (TR-iBRB2). *Mol. Vis.* **2014**, *20*, 1815, <https://pubmed.ncbi.nlm.nih.gov/25593509/>.
100. More, S.V.; Kim, I.S.; Choi, D.K. Recent Update on the Role of Chinese Material Medica and Formulations in Diabetic Retinopathy. *Molecules* **2017**, *22*, <https://doi.org/10.3390/MOLECULES22010076>.
101. Al-Dosari, D.I.; Ahmed, M.M.; Al-Rejaie, S.S.; Alhomida, A.S.; Ola, M.S. Flavonoid Naringenin Attenuates Oxidative Stress, Apoptosis and Improves Neurotrophic Effects in the Diabetic Rat Retina. *Nutrients* **2017**, *9*, <https://doi.org/10.3390/NU9101161>.
102. Bucolo, C.; Leggio, G.M.; Drago, F.; Salomone, S. Eriodictyol prevents early retinal and plasma abnormalities in streptozotocin-induced diabetic rats. *Biochem. Pharmacol.* **2012**, *84*, 88–92, <https://doi.org/10.1016/J.BCP.2012.03.019>.
103. Lv, P.; Yu, J.; Xu, X.; Lu, T.; Xu, F. Eriodictyol inhibits high glucose-induced oxidative stress and inflammation in retinal ganglial cells. *J. Cell. Biochem.* **2019**, *120*, 5644–5651, <https://doi.org/10.1002/JCB.27848>.
104. Liu, W.Y.; Liou, S.S.; Hong, T.Y.; Liu, I.M. The Benefits of the Citrus Flavonoid Diosmin on Human Retinal Pigment Epithelial Cells under High-Glucose Conditions. *Molecules* **2017**, *22*, <https://doi.org/10.3390/MOLECULES22122251>.
105. Kang, M.K.; Lee, E.J.; Kim, Y.H.; Kim, D.Y.; Oh, H.; Kim, S. II; Kang, Y.H. Chrysin Ameliorates Malfunction of Retinoid Visual Cycle through Blocking Activation of AGE-RAGE-ER Stress in Glucose-Stimulated Retinal Pigment Epithelial Cells and Diabetic Eyes. *Nutrients* **2018**, *10*, <https://doi.org/10.3390/NU10081046>.
106. Yuan, Z.; Zhang, J.; Tu, C.; Wang, Z.; Xin, W. The protective effect of blueberry anthocyanins against perfluorooctanoic acid-induced disturbance in planarian (*Dugesia japonica*). *Ecotoxicol. Environ. Saf.* **2016**, *127*, 170–174, <https://doi.org/10.1016/j.ecoenv.2016.01.019>.
107. Pap, N.; Fidelis, M.; Azevedo, L.; do Carmo, MAV; Wang, D.; Mocan, A.; Pereira, E.P.R.; Xavier-Santos, D.; Sant'Ana, A.S.; Yang, B. Berry polyphenols and human health: Evidence of antioxidant, anti-inflammatory, microbiota modulation, and cell-protecting effects. *Curr. Opin. Food Sci.* **2021**, *42*, 167–186, <https://doi.org/10.1016/j.cofs.2021.06.003>.
108. Esposito, D.; Chen, A.; Grace, M.H.; Komarnytsky, S.; Lila, M.A. Inhibitory effects of wild blueberry anthocyanins and other flavonoids on biomarkers of acute and chronic inflammation in vitro. *J. Agric. Food Chem.* **2014**, *62*, 7022–7028, <https://doi.org/10.1021/jf4051599>.
109. Zafra-Stone, S.; Yasmin, T.; Bagchi, M.; Chatterjee, A.; Vinson, J.A.; Bagchi, D. Berry anthocyanins as novel antioxidants in human health and disease prevention. *Mol. Nutr. Food Res.* **2007**, *51*, 675–683, <https://doi.org/10.1002/mnfr.200700002>.
110. Del Bo', C.; Roursgaard, M.; Porrini, M.; Loft, S.; Møller, P.; Riso, P. Different effects of anthocyanins and phenolic acids from wild blueberry (*Vaccinium angustifolium*) on monocytes adhesion to endothelial cells in a TNF- $\alpha$  stimulated pro-inflammatory environment. *Mol. Nutr. Food Res.* **2016**, *60*, 2355–2366, <https://doi.org/10.1002/mnfr.201600178>.
111. Song, Y.; Huang, L.; Yu, J. Effects of blueberry anthocyanins on retinal oxidative stress and inflammation in diabetes through Nrf2/HO-1 signaling. *J. Neuroimmunol.* **2016**, *301*, 1–6, <https://doi.org/10.1016/j.jneuroim.2016.11.001>.
112. Teng, Y.; Cui, H.; Yang, M.; Song, H.; Zhang, Q.; Su, Y.; Zheng, J. Protective effect of Puerarin on diabetic retinopathy in rats. *Mol. Biol. Rep.* **2009**, *36*, 1129–1133, <https://doi.org/10.1007/s11033-008-9288-2>.
113. Hao, L.N.; Wang, M.; Ma, J.L.; Yang, T. Puerarin decreases apoptosis of retinal pigment epithelial cells in diabetic rats by reducing peroxynitrite level and iNOS expression. *Sheng Li Xue Bao* **2012**, *64*, 199–206, <https://pubmed.ncbi.nlm.nih.gov/22513471/>.
114. Liu, J.; Zhang, Y.; Cheng, J.; Tang, X.; Fu, S.; Ma, L.; Tian, Y. Puerarin reduces the levels of AGE-modified proteins in serum and retinal tissues to improve the retinal damage in diabetic rats. *Lat. Am. J. Pharm.* **2019**, *38*, 396–405, [http://www.latamjpharm.org/resumenes/38/2/LAJOP\\_38\\_2\\_1\\_24.pdf](http://www.latamjpharm.org/resumenes/38/2/LAJOP_38_2_1_24.pdf).
115. Lv, B.; Huo, F.; Dang, X.; Xu, Z.; Chen, T.; Zhang, T.; Yang, X. Puerarin attenuates N-methyl-d-aspartic acid-induced apoptosis and retinal ganglion cell damage through the JNK/p38 MAPK pathway. *J. Glaucoma* **2016**, *25*, e792–e801, <https://doi.org/10.1097/IJG.0000000000000505>.
116. Kumar, M.P.; Sankeshi, V.; Naik, R.R.; Thirupathi, P.; Das, B.; Raju, T.N. The inhibitory effect of Isoflavones isolated from *Caesalpinia pulcherrima* on aldose reductase in STZ induced diabetic rats. *Chem. Biol. Interact.* **2015**, *237*, 18–24, <https://doi.org/10.1016/j.cbi.2015.05.010>.

117. Manach, C.; Scalbert, A.; Morand, C.; Rémésy, C.; Jiménez, L. Polyphenols: food sources and bioavailability. *Am. J. Clin. Nutr.* **2004**, *79*, 727–747, <https://doi.org/10.1093/ajcn/79.5.727>.
118. Chen, W.; Lin, B.; Xie, S.; Yang, W.; Lin, J.; Li, Z.; Zhan, Y.; Gui, S.; Lin, B. Naringenin protects RPE cells from NaIO<sub>3</sub>-induced oxidative damage in vivo and in vitro through up-regulation of SIRT1. *Phytomedicine* **2021**, *80*, <https://doi.org/10.1016/j.phymed.2020.153375>.
119. Mancino, R.; di Pierro, D.; Varesi, C.; Cerulli, A.; Feraco, A.; Cedrone, C.; Pinazo-Duran, M.D.; Coletta, M.; Nucci, C. Lipid peroxidation and total antioxidant capacity in vitreous, aqueous humor, and blood samples from patients with diabetic retinopathy. *Mol. Vis.* **2011**, *17*, 1298–1304, <https://pubmed.ncbi.nlm.nih.gov/21633716/>.
120. Zheng, Y.; Zhang, R.; Shi, W.; Li, L.; Liu, H.; Chen, Z.; Wu, L. Metabolism and pharmacological activities of the natural health-benefiting compound diosmin. *Food Funct.* **2020**, *11*, 8472–8492, <https://doi.org/10.1039/d0fo01598a>.
121. Bogucka-Kockaa, A.; Wozniaka, M.; Feldob, M.; Kockic, J.; Szweczyk, K. Diosmin -isolation techniques, determination in plant material and pharmaceutical formulations, and clinical use. *Nat. Prod. Commun.* **2013**, *8*, 545–550, <https://doi.org/10.1177/1934578x1300800435>.
122. Tong, N.; Zhang, Z.; Zhang, W.; Qiu, Y.; Gong, Y.; Yin, L.; Qiu, Q.; Wu, X. Diosmin Alleviates Retinal Edema by Protecting the Blood-Retinal Barrier and Reducing Retinal Vascular Permeability during Ischemia/Reperfusion Injury. *PLoS One* **2013**, *8*, <https://doi.org/10.1371/journal.pone.0061794>.
123. Tong, N.; Zhang, Z.; Gong, Y.; Yin, L.; Wu, X. Diosmin protects rat retina from ischemia/reperfusion injury. *J. Ocul. Pharmacol. Ther.* **2012**, *28*, 459–466, <https://doi.org/10.1089/jop.2011.0218>.
124. Zheng, X.X.; Xu, Y.L.; Li, S.H.; Hui, R.; Wu, Y.J.; Huang, X.H. Effects of green tea catechins with or without caffeine on glycemic control in adults: a meta-analysis of randomized controlled trials. *Am. J. Clin. Nutr.* **2013**, *97*, 750–762, <https://doi.org/10.3945/AJCN.111.032573>.
125. Forte, R.; Cennamo, G.; Bonavolontà, P.; Pascotto, A.; De Crechchio, G.; Cennamo, G. Long-term follow-up of oral administration of flavonoids, Centella asiatica and Melilotus, for diabetic cystoid macular edema without macular thickening. *J. Ocul. Pharmacol. Ther.* **2013**, *29*, 733–737, <https://doi.org/10.1089/JOP.2013.0010>.
126. Yang, L.; Ling, W.; Qiu, Y.; Liu, Y.; Wang, L.; Yang, J.; Wang, C.; Ma, J. Anthocyanins increase serum adiponectin in newly diagnosed diabetes but not in prediabetes: a randomized controlled trial. *Nutr. Metab. (Lond)*. **2020**, *17*, <https://doi.org/10.1186/S12986-020-00498-0>.
127. Mahoney, S.E.; Loprinzi, P.D. Influence of flavonoid-rich fruit and vegetable intake on diabetic retinopathy and diabetes-related biomarkers. *J. Diabetes Complications* **2014**, *28*, 767–771, <https://doi.org/10.1016/J.JDIACOMP.2014.06.011>.
128. Ma, Q.; Chen, D.; Sun, H.P.; Yan, N.; Xu, Y.; Pan, C.W. Regular Chinese Green Tea Consumption is Protective for Diabetic Retinopathy: A Clinic-Based Case-Control Study. *J. Diabetes Res.* **2015**, *2015*, <https://doi.org/10.1155/2015/231570>.
129. Schönlaue, F.; Rohdewald, P. Pycnogenol® for diabetic retinopathy: A review. *Int. Ophthalmol.* **2001**, *24*, 161–171, <https://doi.org/10.1023/A:1021160924583>.
130. Steigerwalt, R.; Belcaro, G.; Cesarone, M.R.; Di Renzo, A.; Grossi, M.G.; Ricci, A.; Dugall, M.; Cacchio, M.; Schönlaue, F. Pycnogenol improves microcirculation, retinal edema, and visual acuity in early diabetic retinopathy. *J. Ocul. Pharmacol. Ther.* **2009**, *25*, 537–540, <https://doi.org/10.1089/JOP.2009.0023>.
131. Domanico, D.; Fragiotta, S.; Cutini, A.; Carnevale, C.; Zompatori, L.; Vingolo, E. Circulating levels of reactive oxygen species in patients with nonproliferative diabetic retinopathy and the influence of antioxidant supplementation: 6-Month follow-up. *Indian J. Ophthalmol.* **2015**, *63*, 9–14, <https://doi.org/10.4103/0301-4738.151455>.
132. Garcia-Medina, J.J.; Rubio-Velazquez, E.; Foulquie-Moreno, E.; Casaroli-Marano, R.P.; Pinazo-Duran, M.D.; Zanon-Moreno, V.; Del-Rio-vellosillo, M. Update on the effects of antioxidants on diabetic retinopathy: In vitro experiments, animal studies and clinical trials. *Antioxidants* **2020**, *9*, 1–23, <https://doi.org/10.3390/ANTIOX9060561>.
133. Kunkel, B.N.; Brooks, D.M. Cross talk between signaling pathways in pathogen defense. *Curr. Opin. Plant Biol.* **2002**, *5*, 325–331, [https://doi.org/10.1016/S1369-5266\(02\)00275-3](https://doi.org/10.1016/S1369-5266(02)00275-3).
134. Schönlaue, F.; Rohdewald, P. Pycnogenol for diabetic retinopathy. A review. *Int. Ophthalmol.* **2001**, *24*, 161–171, <https://doi.org/10.1023/A:1021160924583>.