

**Influence of magnetic MWCNTs on the antimicrobial activity of cephalosporins**  
**Alexandru Mihai Grumezescu<sup>1</sup>, Ecaterina Ilinca<sup>2</sup>, Carmen Chifiriuc<sup>2\*</sup>, Dan Mihaiescu<sup>1</sup>, Paul Balaure<sup>1</sup>, Vanessa Traistaru<sup>1</sup>, Grigore Mihaiescu<sup>2</sup>**

**ABSTRACT**

The purpose of this study was to analyze the antimicrobial activity of cephalosporin antibiotics adsorbed on the surface of magnetic MWCNTs, against microbial cells in suspension and adhered to the substrate, in order to highlight potential improvements of antimicrobial activity parameters of these antibiotics. Magnetic MWCNTs were obtained by plasma processing and characterized by HR-TEM. The need for the discovery of such strategies is imposed by the numerous mechanisms developed by pathogenic microorganisms manage to gain resistance to the majority of currently available antibiotics.

**Keywords:** *magnetic MWCNTs, cephalosporins, antimicrobial activity, nanomaterials*

## 1. Introduction

Carbon nanotubes (CNTs) are one of the new nanomaterials that have excellent optical, electronic, thermal [1], chemical [2], and mechanical [3] properties arising from their unique chemical [4] structure and size [5]. The interest in CNTs is gradually increasing due to their special properties related to electrical conduction and chemical affinity for various chemical species [6]. The applications of these nanomaterials cover a large spectrum, especially in biosciences: drug delivery, nanomedicine [7], cell growth [8] etc. Nanotubes are classified as single-walled nanotubes (SWNTs) and multi-walled nanotubes (MWNTs) and both categories could be functionalized and non functionalized in order to assure good adherence for different substrates [9] as well as improved biocompatibility. Nanopharmaceutical products based on carbon nanotube technologies will require a full understanding of the physical and chemical characteristics of CNTs and their interaction with the biological systems [10]. The application of CNTs as carriers is increased by their propensity to penetrate cells [11]. Cationic functionalized CNTs can be bound to active molecules via stable covalent bonds or supramolecular assemblies based on electrostatic attractions. Two possibilities exist: the more energetically feasible attachment onto the exterior either by covalent or non covalent interactions, and the encapsulation of these molecular assemblies within CNTs [12,13,14,15]. Cephalosporins are antibiotics with a structure and activity similar to penicillins [16]. They are resistant to penicillinase, but susceptible to cephalosporinases. They have a large action spectrum, including Gram-positive (*Staphylococcus sp.*, *Streptococcus sp.*, *Corynebacterium diphtheriae*) and

<sup>1</sup> Faculty of Applied Chemistry and Materials Science, Politehnica University of Bucharest, Romania

\*Corresponding author e-mail address: [carmen\\_balotescu@yahoo.com](mailto:carmen_balotescu@yahoo.com)

<sup>2</sup> Faculty of Biology, University of Bucharest, Bucharest, Romania

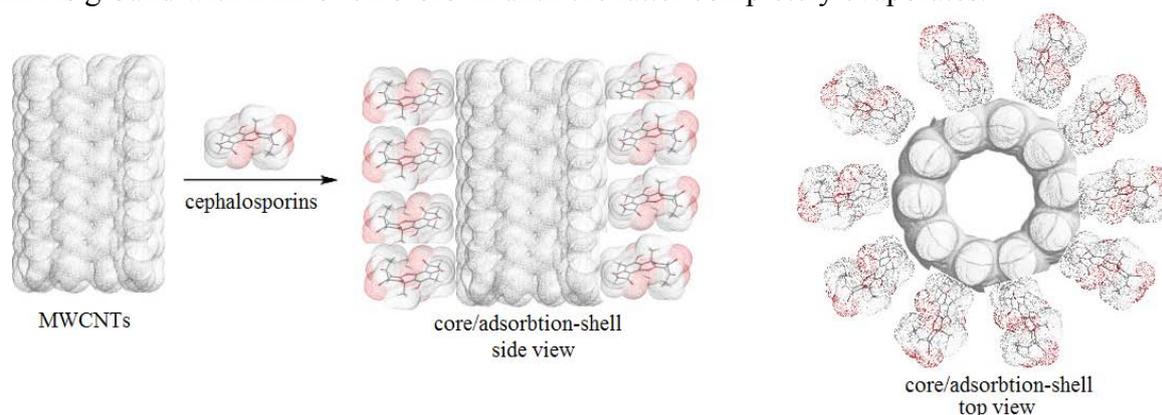
Gram-negative (*Neisseria gonorrhoeae*, *Haemophilus influenzae*, *Escherichia coli*, *Proteus sp.*, *Klebsiella sp.*) microorganisms.

In this context our aim was to study the influence of magnetic MWCNTs on the antimicrobial activity of cephalosporins from II<sup>nd</sup>, III<sup>rd</sup> and IV<sup>th</sup> generation on Gram-positive and Gram-negative bacterial strains with known antibiotic susceptibility patterns.

## 2. Experimental section

**2.1. Synthesis and characterization of magnetic MWCNTs.** Magnetic nanoparticles were obtained by toluene plasma processing [17], and the morphology was determined by transmission electronic microscopy (TEM). The transmission electron images were obtained on finely powdered samples using a Tecnai<sup>TM</sup> G2F30 S-TWIN high resolution transmission electron microscope (HR-TEM). The microscope was operated in transmission mode at 300kV with TEM point resolution of 2 Å and line resolution of 1 Å [18].

**2.2. Preparation of magnetic MWCNTs/Cephalosporins hybrid materials.** The following cephalosporins have been selected for adsorption on the hybrid material: Cefaclor, Cefoperazone, Ceftriaxone, Cefpirome, Cefotaxime and Cefepime. The amount of the antibiotic adsorbed on the nanostructured support was 3 %. In a grinding mortar equipped with a 100 Kgf Nd-Fe-B magnet at the bottom side, the nanostructured material and antibiotic which is to be adsorbed are introduced. The mix is ground with 2 ml of chloroform until the latter completely evaporates.



**Figure 1.** Illustrative representation of cephalosporins adsorbed on the surface of magnetic MWCNTs

### 2.3. Evaluation of the antimicrobial activity of cephalosporins and MWCNTs/cephalosporins

**2.3.1. Bacterial strains.** Bacterial strains used in the study were selected from the collection of the Laboratory of Microbiology-Botany Department, and belong to the following species: *Klebsiella pneumoniae*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Bacillus subtilis*.

**2.3.2. Qualitative screening of the antimicrobial activity.** Qualitative screening of the susceptibility of different microbial strains to magnetic MWCNTs/cephalosporins has been accomplished through a adapted diffusion method, on Mueller Hinton solid medium [19, 20]. In this purpose, 5 µl from a stock solution of the tested product, containing 30 µl of antibiotic were distributed in spots on Petri plates. The results' reading was performed by measuring the bacterial growth inhibition zones' diameters around the spots. The used solvent, dimethyl sulfoxide (DMSO) [21], was comparatively tested for its potential antimicrobial activity.

**2.3.3. Quantitative assay of the antimicrobial activity.** Quantitative testing of antimicrobial activity of hybrid nanosystems and the establishment of MIC (minimum inhibitory concentration) was determined by microdilution technique in liquid medium (Mueller Hinton broth), using 96 multiwell plates [22]. Twofold serial microdilutions were achieved in 200 µl medium, the dilution

range varying, depending on the tested antibiotic and the bacterial strain, in accordance with CLSI breakpoints (CLSI, 2010). Subsequently, the wells were seeded with 50 ml of each bacterial suspension, adjusted at MacFarland standard 0,5 [23]. Positive and negative controls were used. After incubating the plates at 37°C for 24 hours, the results were macroscopically assessed for bacterial growth, MIC corresponding to the well with clear content, thus without any visible microbial growth.

### 3. Results section

**3.1. HR-TEM analysis.** The multiple graphitic walls of the CNTs were clearly visible in TEM image, which showed that diameter of the tube not exceeding 15 nm (Fig. 2).

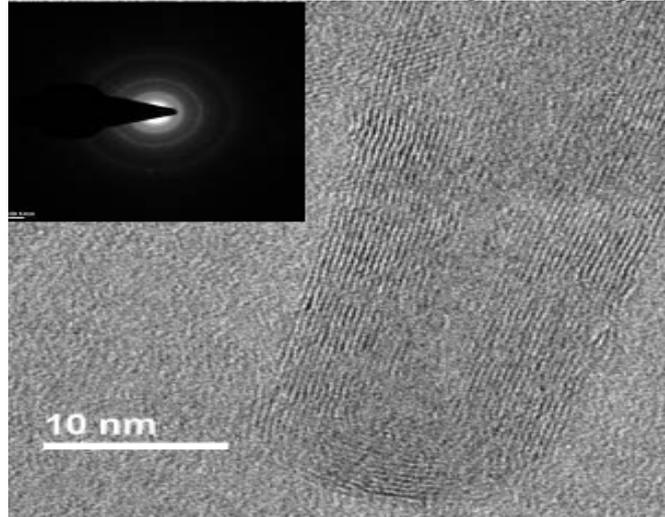
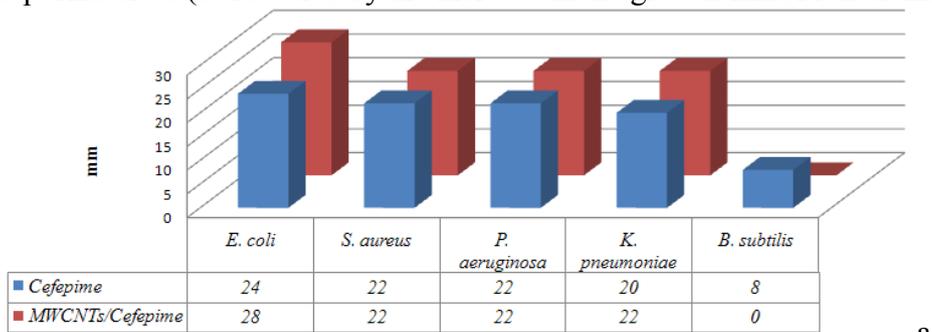
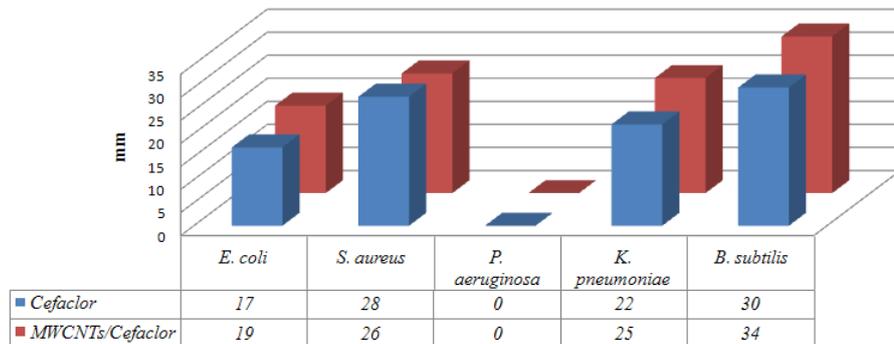


Figure 2. HR-TEM images of magnetic MWCNTs

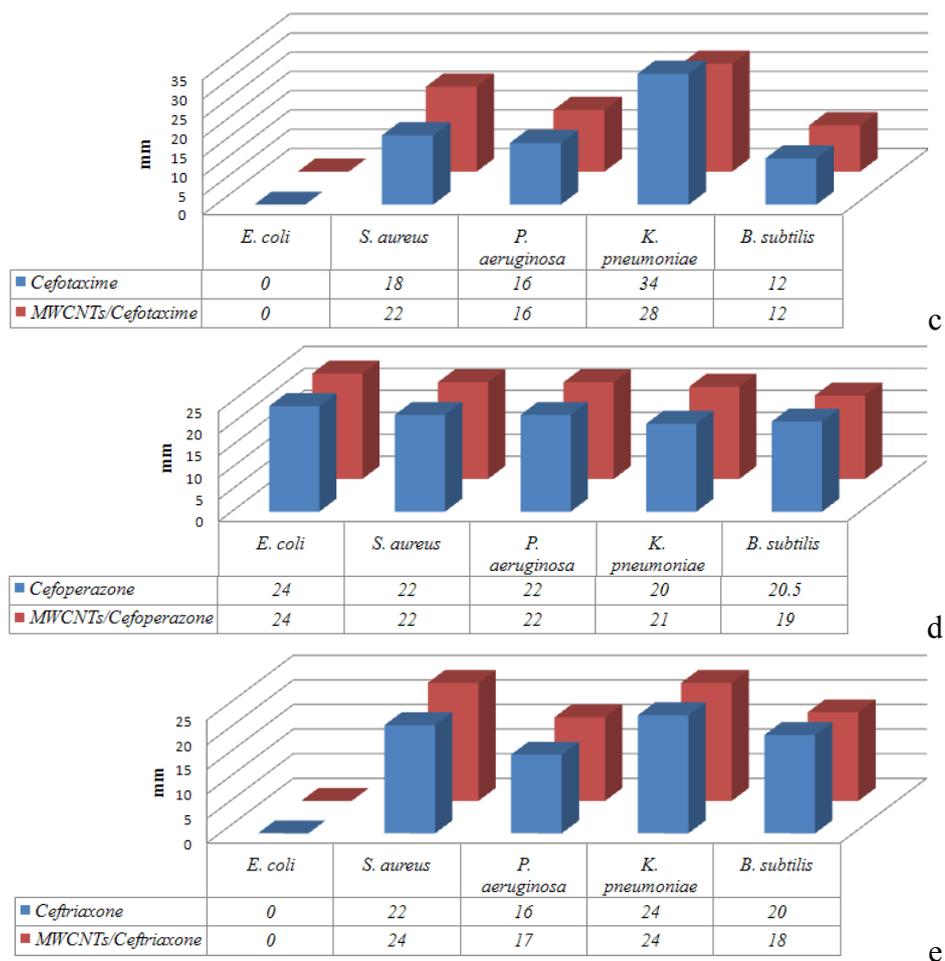
**3.2. Qualitative testing results of susceptibility of bacterial strains to the tested cephalosporins, as well as embedded in NTs.** The qualitative screening revealed that the antibacterial activity of hybrid nanosystems was improved or at least similar on Gram-negative strains comparing with the standard cephalosporine discs (as revealed by the increase in the growth inhibition diameter).



a



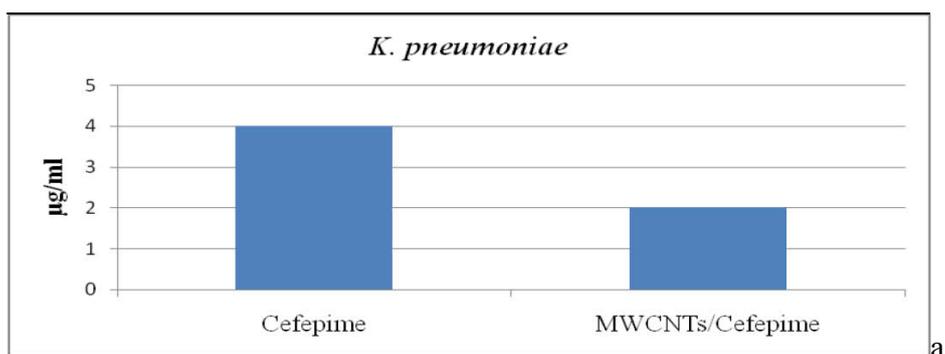
b

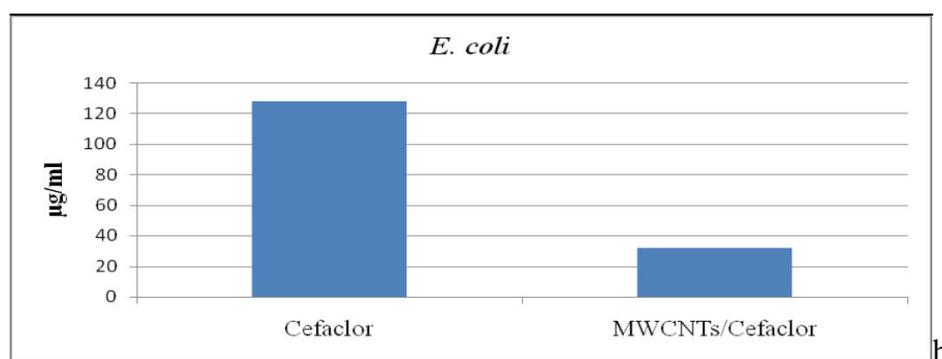


**Figure 3 (a-e).** The comparative graphic representation of the growth inhibition zone diameter values obtained for cefepime (a), cefaclor (b), cefotaxime (c), cefoperazone (d) and ceftriaxone (e) as well and respectively, embedded in magnetic MWCNTs on different bacterial strains

Improved activity was obtained for cefepime and cefaclor against *E. coli* and *K. pneumoniae*, for ceftriaxone against *K. pneumoniae* and for cefoperazone on *P. aeruginosa*. Concerning the Gram-positive strains, cefotaxime activity on *S. aureus* and cefaclor's on *B. subtilis* were enhanced by the hybrid nanosystems (Fig. 3 a-e).

**3.3. Quantitative analysis (MIC) of antimicrobial activity.** The quantitative assay confirmed the qualitative screening results, significantly decreased MIC values being achieved for cefaclor loaded hybrid nanosystem on *E. coli* strain, from 128 $\mu$ g/mL to 32 $\mu$ g/mL. The MIC value slightly decreased for cefepime charged nanosamples, from 4 $\mu$ g/mL to 2 $\mu$ g/mL (Fig. 4 a, b).





**Figure 4.** The comparative graphic representation of the MIC values obtained for cephalosporins as well and respectively, embedded magnetic MWCNTs on *K. pneumoniae* (a) and *E. coli* (b) bacterial strains

## 4. Conclusions

Magnetic MWCNTs significantly improved the antimicrobial activity of cephalosporin antibiotics adsorbed onto their surface in a specific manner, dependent on the tested microbial strain and the tested antibiotic, demonstrating that this type of nanosystems could be used not only as drug delivery systems, but also as active potentiators of the antimicrobial activity of different substances. However, in-depth studies are required in order to confirm these results on a large number of strains and to elucidate the interactions established between the active substances and the carrier system on one side, and between the hybrid nanosystem and the target cell on the other one.

## 5. References

- [1] Sivakumar V. M., Abdullah A.Z., Mohamed A.R., Chai S.P., Studies on carbon nanotube synthesis via methane cvd process using  $\text{CoO}_x$  as catalyst on carbon supports, *Digest J. of Nanom. and Biostruct.*, 5, 3, 691–698, **2010**
- [2] Ghoreishi S. M., Behpour M., Khayatkashani M., Motaghedifard M. H., New applied method for simultaneous determination of ellagic and tannic acid by multi-wall carbon nanotube paste electrode: application in quantification punica granatum and quercus infectoria, *Digest J. of Nanom. and Biostruct.*, 6, 2, 625-635, **2011**
- [3] Dima D., Murescu M., Andrei G., Dispersion of carbon nanotubes coated with iron (iii) oxide into polymer composite under oscillating magnetic field, *Digest J. of Nanom. and Biostruct.*, 5, 4, 1009-1014, **2010**
- [4] Soleimani B., Rostamnia S., Ahmadi A., mathematical modeling for the simulation of heavy metal ions adsorption by single wall carbon nanotubes (SWCNTs) based on computational calculation, *Digest J. of Nanom. and Biostruct.*, 5, 1, 153 – 159, **2010**
- [5] Haegeun Chung A.N., Yowhanson A., Taekyungyoon A., Seungwookkim B., Woongkim B.N., The effect of multi-walled carbon nanotubes on soil microbial activity, *Ecotoxicology And Environmental Safety*, 74, 569–575, **2011**
- [6] Şimăndan I. D., Popescu M., Lőrinczi A., Velea A., Fagadar-Cosma E., Preparation and properties of barium stearate multilayers with carbon nanotubes, manganese porphyrin and silver nitrate, *Digest J. of Nanom. and Biostruct.*, 5, 4, 1029 – 1033, **2010**
- [7] Popescu M., Velea A., Lőrinczi A., Biogenic production of nanoparticles, *Digest J. of Nanom. and Biostruct.*, 5, 4, 1035, **2010**
- [8] Matei A., Dinescu M., Buruiana E.C., Buruiana T., Petcu I., Mustaciosu C., Ormosils scaffolds produced by laser processing for fibroblast cell growth, *Digest J. of Nanom. and Biostruct.*, 6, 1, 29, **2011**

- [9]Prodana M., Ionita D., Ungureanu C., Bojin D., Demetrescu I., Enhancing antibacterial effect of multiwalled carbon nanotubes using silver nanoparticles, *Digest J. of Nanom. and Biostruct.*, 6, 2, 549 – 556, **2011**
- [10]Foldvari M., Bagonluri M., Carbon nanotubes as functional excipients for nanomedicines: II. Drug delivery and biocompatibility issues, *Nanomedicine: Nanotechnology, Biology, And Medicine*, 4, 3, 183–200, **2008**
- [11]Wu W., Wieckowski S., Pastorin G., Benincasa M., Klumpp C., Briand Jp., Et Al., Targeted delivery of amphotericin B to cells by using functionalized carbon nanotubes, *Angew Chem Int Edn Engl*, 44, 6358-62, **2005**
- [12]Singh R., Pantarotto D., Mccarthy D., Chaloin O., Hoebeke J., Partidos Cd., et al., Binding and Condensation of Plasmid DNA onto Functionalized Carbon Nanotubes: Toward the Construction of Nanotube-Based Gene Delivery Vectors, *J Am Chem Soc*, 127, 4388-96, **2005**
- [13]Pantarotto D., Partidos Cd., Hoebeke J., Brown F., Kramer E., Briand Jp., et al., Immunization with peptide-functionalized carbon nanotubes enhances virus-specific neutralizing antibody responses, *Chem Biol*, 10, 961-6, **2003**
- [14] Bianco A., Hoebeke J., Godefroy S., Chaloin O., Pantarotto D., Briand Jp., et al., Cationic carbon nanotubes bind to CpG oligodeoxynucleotides and enhance their immunostimulatory properties, *J Am Chem Soc*, 127, 58-9, **2005**
- [15]Pantarotto D., Briand Jp., Prato M., Bianco A., Translocation of bioactive peptides across cell membranes by carbon nanotubes, *Chem Commun (Camb)* 10, 16-7, **2004**
- [16]Mihaiescu D.E., Grumezescu A.M., Mogosanu D.E., Traistaru V., Balaure P.C., Buteica A.S., Hybrid inorganic/organic nanomaterial for controlled cephalosporins release, *Biointerface Res. in App. Chem.*, 1, 2, 041-047, **2011**
- [17] Buteică A. S., Mihaiescu D. E., Grumezescu A. M., Vasile B. S., Popescu A., Călina D., Mihaiescu O. M., The cytotoxicity of (non)magnetic nanoparticles tested on *Escherichia coli* and *Staphylococcus aureus*, *Digest J. of Nanom. and Biostructures*, 5, 3, 651, **2010**
- [18]Ficai D., Ficai A., Vasile B. S., Ficai M., Oprea O., Guran C., Andronescu E., Synthesis of rod-like magnetite by using low magnetic field, *Digest J. Of Nanomat. And Biostruct.*, 6, 3, 943 – 951, **2011**
- [19]Saviuc C., Grumezescu A. M., Holban A., Chifiriuc C., Mihaiescu D., Lazar V., Hybrid nanostructured material for biomedical applications, *Biointerface Res. in App. Chem.*, 1, 2, 064-071, **2011**
- [20]Saviuc C., Grumezescu A. M., Holban A., Bleotu C., Chifiriuc C., Balaure P., Lazar V., Phenotypical studies of raw and nanosystem embedded *Eugenia carryophyllata* buds essential oil antibacterial activity on *Pseudomonas aeruginosa* and *Staphylococcus aureus* strains, *Biointerface Res. in App. Chem.*, 1, 3, 111-118, **2011**
- [21]Saviuc C., Grumezescu A. M., Oprea E., Radulescu V., Dascalu L., Chifiriuc M. C., Bucur M., Banu O., Lazar V., Antifungal activity of some vegetal extracts on Candidabiofilms developed on inert substratum, *Biointerface Res. in App. Chem.*, 1, 1, 015-023, **2011**
- [22]Banu O., Bleotu C., Chifiriuc M. C., Savu B., Stanciu G., Antal C., Alexandrescu M., Lazăr V., Virulence factors of *Staphylococcus aureus* and *Pseudomonas aeruginosa* strains involved in the etiology of cardiovascular infections, *Biointerface Res. in App. Chem.*, 1, 2, 72-77, **2011**
- [23]Panus E., Chifiriuc C.M., Banu O., Mitache M., Bleotu C., Rosoiu N., Lazar V., Comparative study of resistance and virulence markers in *Escherichia coli* strains isolated from hospital surfaces, clinical specimens and drinking/ marine waters, *Biointerface Res. in App. Chem.*, 1, 1, 24-30, **2011**

**Acknowledgements:** The results presented in this paper have been obtained with the financial support obtained from the Human Resources Project no. 135/2010 (Contract no. 76/2010) and PN2 42150/2008.