Volume 8, Issue 3, 2018, 3306 - 3313

# **Biointerface Research in Applied Chemistry**

www.BiointerfaceResearch.com

# **Original Research Article**

**Open Access Journal** 

Received: 20.06.2018 / Revised: 10.06.2018 / Accepted: 11.06.2018 / Published on-line: 15.06.2018

Synthesis and evaluation of biological activity from two steroid-diazacyclododecin derivatives on left ventricular pressure

Rosas-Nexticapa Marcela<sup>1</sup>, Figueroa-Valverde Lauro<sup>2,\*</sup>, Diaz-Cedillo Francisco<sup>3</sup>, Mateu-Armand Virginia<sup>2</sup>, Lopez-Ramos Maria<sup>2</sup>, García-Cervera Elodia<sup>2</sup>, Pool Gómez Eduardo<sup>2</sup>, García-Martínez Rolando<sup>2</sup>, Parra-Galindo Perla<sup>1</sup>, Cauich-Carrillo Regina<sup>2</sup>, Alfonso-Jimenez Alondra<sup>2</sup>, Cabrera-Tuz Jhair<sup>2</sup>

<sup>1</sup>Facultad de Nutrición, Universidad Veracruzana. Médicos y Odontólogos s/n, 91010, Xalapa, Veracruz. México

<sup>2</sup>Laboratory of Pharmaco-Chemistry at the Faculty of Chemical Biological Sciences of the University Autonomous of Campeche, Av. Agustín Melgar s/n, Col Buenavista C.P.24039 Campeche Cam., México.

<sup>3</sup>Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional, Col. Santo Tomás, México, D.F., C.P. 11340

\*corresponding author e-mail address: lauro\_1999@yahoo.com

### ABSTRACT

There are several studies which indicate that some cardiovascular diseases are relationship with biological activity of estrogens through of its receptors activation. The objective of this work was synthesizing two new steroid-diazacyclododecin derivatives (compounds 11 or 12) to evaluate their effect on left ventricular pressure in Langendorff model using as control to  $17\beta$ -estradiol. In addition, a theoretical study was carried out to evaluate the interaction of compound 11 or 12 with the estrogen receptor (30s8 protein) using a docking model. The experimental results showed that only the compound 12 decreased left ventricular pressure in a similar form to  $17\beta$ -estradiol. In addition, other experimental data showed that effect exerted by  $17\beta$ -estradiol was inhibited in presence of 12. Finally, theoretical results indicated that interaction of 12 with 30s8 protein involved some aminoacid residues such as  $Pro_{224}$ ,  $Leu_{319}$ ,  $Leu_{320}$ ,  $Leu_{327}$ ,  $Asp_{321}$ ,  $Ala_{322}$ ,  $Glu_{323}$ ,  $Pro_{324}$ ,  $Pro_{325}$ ,  $Ile_{326}$  and  $Leu_{327}$ . All these data suggest that compound 12 may act as a selective agonist of estrogen receptor which translated as changes on left ventricular pressure.

**Keywords:** *Steroid-diazacyclododeci; derivatives; 17β-estradiol; left ventricular pressure; docking.* 

### 1. INTRODUCTION

Cardiovascular diseases are one of the main health problems in the world; several reports suggest that exogenous sex hormones have an influence on these clinical pathologies. For example, one study showed that estrogens can reduce the development of coronary heart diseases [1]. Another data indicates that 17β-estradiol; protect blood vessels from atherosclerotic lesion formation [2, 3]. It is noteworthy that the cellular effects exerted by estrogen can be controlled by the expression of nuclear receptors [4]. In this sense, several studies carried out in patients with cardiovascular diseases (coronary artery disease and atherosclerosis) and estrogen therapy showed an association between these clinical pathologies and estrogen receptor activation [5-10]. In addition to evaluate this association, a series of studies have been carried out using various estrogen receptor agonists or antagonist. A study showed that 16a-lactone-estradiol activates estrogen receptor which results as decrease cardiac hypertrophy using an animal model [11]. Other data indicated that propyl

### **2. EXPERIMENTAL SECTION**

*Chemical synthesis*. The compounds 2-nitro-estrone and 2-nitro-estradiol were prepared using previously methods reported [16, 17]. Additionally, all the reagents used in this study were purchased from Sigma-Aldrich Sigma-Aldrich Co., Ltd. The melting point for compounds was evaluated on an Electrothermal (900 model). Infrared spectra (IR) were determined using KBr pellets on a Perkin Elmer Lambda 40 spectrometer.<sup>1</sup>H and <sup>13</sup>C

pyrazole and diarylpropiolnitrile exert cardioprotection through estrogen receptor activation in a hemorrhage injury model [12]. Also, a study showed that raloxifene relaxes coronary arteries via estrogen receptor activation in vitro [13]. However, other studies indicate that 27-hydroxycholesterol (endogenous oxysterol) inhibits the cardiovascular effects of estrogen [14]. Also, a study showed that tamoxifen and fulvestrant induce changes on cell growth via estrogen receptor [15]. All these data indicate that some drugs exert their biological activity as agonist or antagonist of estrogen receptors; this phenomenon can be due to the different functional groups involved in the chemical structure of each compound or to different protocols used. Analyzing this hypothesis, in this study two steroid-diazacyclododecin derivatives were synthesized to evaluate their biological activity using a Langendorff model. In addition, a theoretical study was conducted to characterize its interaction with estrogen receptor.

NMR (nuclear magnetic resonance) spectra were recorded on a Varian VXR300/5 FT NMR spectrometer at 300 and 75.4 MHz (megahertz) in  $CDCl_3$  (deuterated chloroform) using TMS (tetramethylsilane) as an internal standard. EIMS (electron impact mass spectroscopy) spectra were determined using a Finnigan Trace Gas Chromatography Polaris Q-Spectrometer. Elementary

analysis data were determined from a Perkin Elmer Ser. II CHNS/02400 elemental analyzer.

### General method for preparation of tert-Butyl-dimethylsilanyloxy-nitro-steroid derivatives (3 or 4).

A solution of compound 1 or 2 (0.50 mmol), *tert*butyldimethylsilyl chloride (200  $\mu$ l, 1.07 mmol) in 3 ml of chloroform was stirring for 12 h at room temperature. The mixture obtained was dried under reduced pressure and purified by crystallization using the methanol:water (4:1) system.

## **3**-(tert-Butyl-dimethyl-silanyloxy)-13-methyl-17-(1-methyl-1trimethylsilanyl-ethyl)-2-nitro-7,8,9,11,12,13,14,15,16,17-decahydro-6H-cyclopenta[a]phenanthrene (3)

Yielding 68 % of product, m.p. 66-68 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 1352 and 1080: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm H}$ : 0.08 (s, 6H), 0.22 (s, 6H), 0.80 (s, 3H), 0.86 (s, 9H), 0.98 (s, 9H), 1.06-1.92 (m, 10H), 2.10-7.80 (m, 7H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm C}$ : -4.50, -4.40, 15.22, 17.80, 18.22, 25.34, 25.62, 25.70, 25.74, 27.76, 29.84, 32.97, 35.07, 37.28, 43.58, 43.74, 51.49, 82.62, 115.62, 122.73, 132.52, 133.34, 144.24, 151.72 ppm. EI-MS m/z: 545.33 Anal. Calcd. for C<sub>30</sub>H<sub>51</sub>NO<sub>4</sub>Si<sub>2</sub>: C, 66.10; H, 9.42; N, 2.57; O, 11.72; Si, 10.29. Found: C, 66.02; H, 9.36.

# 3-(tert-Butyl-dimethyl-silanyloxy)-13-methyl-2-nitro-6,7,8,9, 11,12,13,14,15,16-deca- hydro-cyclopenta[a]phenanthren-17one (4)

Yielding 68 % of product, m.p. 50-52 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 1350 and 1082: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 0.22 (s, 6H), 0.92 (s, 3H), 1.00 (s, 9H), 120-1.90 (m, 7H), 2.10-7.90 (m, 10H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{C}$ : -4.44, 13.82, 18.22, 21.70, 25.51, 25.74, 26.40, 29.84, 31.06, 35.00, 37.24, 46.40, 48.34, 50.12, 115.62, 122.34, 132.00, 133.34, 143.94, 151.72, 219.70 ppm. EI-MS m/z: 429.23 Anal. Calcd. for C<sub>24</sub>H<sub>35</sub>NO<sub>4</sub>Si: C, 67.10; H, 8.21; N, 3.26; O, 14.90; Si, 6.54. Found: C, 67.02; H, 8.16.

### Synthesis of two tert-Butyl-dimethyl-silanyloxy-steroidcarbaldehyde derivatives (5 or 6)

In a round bottom flask (10 ml), the compound 3 or 4 (0.50 mmol), 2-hydroxy-1-naphthaldehyde (100 mg, 0.58 mmol) and potassium carbonate anhydrous (50 mg, 0.36 mmol) in 5 ml of dimethyl sulfoxide were stirring for 72 h at room temperature. The reaction mixture was evaporated to dryness under reduced pressure. After, the residue was purified by crystallization from methanol:hexane:water (4:2:1)

### 8-[3,17-Bis-(tert-butyl-dimethyl-silanyloxy)-13-methyl-7,8,9, 11,12,13,14,15,16,17-decahydro-6H-cyclopenta[a]phenanthren-2-yloxy]-naphthalene-1-carbaldehyde (5)

Yielding 54 % of product, m.p. 140-142 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 1724, 1350 and 1082: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm H}$ : 0.06 (s, 6H), 0.18 (s, 6H), 0.80 (s, 3H), 0.86 (s, 9H), 0.98 (m, 1H), 1.00 (s, 9H), 110-1.90 (m, 10H), 2.10-6.52 (m, 7H), 7.10-8.40 (m, 6H), 9.96 (broad, 1H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm C}$ : 9.96 (broad, 1H) ppm. <sup>-4.55</sup>, -4.50, 15.22, 17.84, 18.22, 25.32, 25.56, 25.74, 25.92, 27.76, 29.65, 32.97, 35.07, 37.24, 43.74, 44.00, 51.46, 82.60, 112.50, 114.94, 118.12, 120.92, 121.14, 126.72, 126.94, 128.44, 132.74, 135.62, 136.84, 137.62, 138.44, 143.64, 144.33, 147.92, 197.82 ppm. EI-MS m/z: 670.38 Anal.

Calcd. for  $C_{41}H_{58}O_4Si_2$ : C, 73.28; H, 8.71; O, 9.54; Si, 6.54. Found: C, 73.20; H, 8.66.

### 8-(((13S)-3-((tert-butyldimethylsilyl)oxy)-13-methyl-17-oxo-7,8,9,11,12,13,14,15,16,17-decahydro-6H-

### cyclopenta[a]phenanthren-2-yl)oxy)-1-naphthaldehyde (6)

yielding 54 % of product, m.p. 140-142 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 3466, 3402, 1712 and 1620: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm H}$ : 0.18 (s, 6H), 0.90 (s, 3H), 1.00 (s, 9H), 120-1.90 (m, 7H), 2.10-6.52 (m, 10H), 7.10-8.40 (m, 6H), 9.96 (broad, 1H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm C}$ : 9.96 (broad, 1H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm C}$ : -4.54, 13.80, 18.22, 21.78, 25.84, 25.92, 26.40, 29.64, 31.50, 35.40, 37.52, 46.84, 48.12, 50.40, 112.24, 114.94, 118.12, 120.93, 121.14, 126.60, 126.74, 128.40, 132.74, 135.62, 136.82, 137.12, 138.50, 143.64, 144.38, 147.90, 197.80, 220.70 ppm. EI-MS m/z: 554.28 Anal. Calcd. for  $C_{35}H_{42}O_4$ Si: C, 75.77; H, 7.63; O, 11.54; Si, 5.06. Found: C, 75.66; H, 7.58.

**Deprotection of hydroxyl group.** In a round bottom flask (10 ml), the compound 5 or **6** (0.50 mmol), 10 ml of Hydrofluoric acid were stirring for 72 h at room temperature. The product obtained was dried under reduced pressure. Then, the mixture was purified through of crystallization with methanol:benzene:water (4:2:1) system

# 8-(3,17-Dihydroxy-13-methyl-7,8,9,11,12,13,14,15,16,17-decahydro-6H-cyclopenta[a]phenanthren-2-yloxy)-naphthalene-1carbaldehyde (7)

yielding 54 % of product, m.p. 308-310 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 3400, 1724, 1222 and 1080: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 0.76 (s, 3H), 0.82-1.90 (m, 11H), 2.10-3.64 (m, 5H), 5.90 (broad, 2H), 6.04-6.70 (m, 2H), 7.12-8.40 (m, 6H), 9.96 (d, 2H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{C}$ : 15.76, 24.22, 25.24, 27.76, 29.64, 32.74, 33.68, 37.24, 44.00, 44.42, 50.78, 82.42, 113.50, 115.00, 116.24, 120.92, 121.08, 126.74, 128.00, 128.40, 132.74, 136.80, 137.94, 138.52, 143.44, 143.94, 144.38, 151.60, 197.82 ppm. EI-MS m/z: 442.21 Anal. Calcd. for C<sub>29</sub>H<sub>30</sub>O<sub>4</sub>: C, 78.71; H, 6.83; O, 14.46. Found: C, 78.64; H, 6.76.

## 8-(3-Hydroxy-13-methyl-17-oxo-7,8,9,11,12,13,14,15,16,17decahydro-6H-cyclopenta[a]phenanthren-2-yloxy)-naphthalene-1-carbaldehyde (8)

yielding 54 % of product, m.p. 234-236 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 1728, 1725 and 1220: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 0.90 (s, 3H), 120-1.90 (m, 7H), 2.10-2.80 (m, 8H), 5.36 (broad, 1H), 6.14-6.70 (m, 2H), 7.12-8.40 (m, 6H), 9.96 (broad, 1H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 13.80, 21.74, 25.84, 26.44, 29.64, 31.48, 35.42, 37.54, 46.87, 48.11, 50.41, 113.14,115.00, 16.28, 120.92, 121.11, 126.74, 127.60, 128.41, 132.77, 136.84, 137.50, 138.50, 143.48, 143.94, 144.38, 151.54, 197.80, 220.70 ppm. EI-MS m/z: 440.19 Anal. Calcd. for C<sub>29</sub>H<sub>28</sub>O<sub>4</sub>: C, 79.07; H, 6.41; O, 14.53. Found: C, 79.00; H, 6.36.

**Preparation of formyl-steroid-carbaldehyde complex**. In a round bottom flask (50 ml), the compound **7** or **8** (0.50 mmol) and 10 ml of dimethyl sulfoxide were stirring for 72 h at room temperature. The product was dried under reduced pressure. Then, the mixture was purified by crystallization with the methanol:bencene:water (3:1) system

Rosas-Nexticapa Marcela, Figueroa-Valverde Lauro, Diaz-Cedillo Francisco, Mateu-Armand Virginia, Lopez-Ramos Maria, García-Cervera Elodia, Pool Gómez Eduardo, García-Martínez Rolando, Parra-Galindo Perla, Cauich-Carrillo Regina, Alfonso-Jimenez Alondra, Cabrera-Tuz Jhair

# 2-(8-Formyl-naphthalen-1-yloxy)-17-hydroxy-13-methyl-7,8,9, 11,12,13,14,15,16,17-decahydro-6H-cyclopenta[a]phenanthrene-3-carbaldehyde (9)

Yielding 54 % of product, m.p. 132-134 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 3400, 1724 and 1220: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm H}$ : 0.76 (s, 3H), 0.80-3.62 (m, 16H), 6.40 (broad, 1H), 7.10 (m, 1H), 7.24-7.62 (m, 3H), 7.66 (m, 1H), 7.70-8.40 (m,3H), 10.00-10.44 (m, 2H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm C}$ : 15.80, 24.22, 25.34, 28.00, 29.61, 32.78, 33.71, 37.28, 44.02, 44.39, 50.76, 82.46, 107.02, 115.20, 119.92, 120.52, 121.12, 126.74, 127.17, 128.50, 129.24, 129.54, 135.62, 138.49, 144.36, 154.60, 154.76, 164.64, 189.50, 197.80 ppm. EI-MS m/z: 454.21 Anal. Calcd. for C<sub>30</sub>H<sub>30</sub>O<sub>4</sub>: C, 79.27; H, 6.65; 0, 14.08. Found: C, 79.18; H, 6.56.

## 2-(8-Formyl-naphthalen-1-yloxy)-13-methyl-17-oxo-7,8,9,11, 12,13,14,15,16,17-decahydro-6H-cyclopenta[a]phenanthrene-3-carbaldehyde (10)

Yielding 54 % of product, m.p. 118-120 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 1728, 1724 and 1220: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : s (0.88), 1.20-7.16 (m, 16H), 7.26-7.62 (m, 3H), 7.66 (m, 1H), 7.68-8.40 (m, 3H), 10.00-10.44 (m, 2H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{C}$ : 13.82, 21.74, 25.87, 27.55, 29.61, 31.51, 35.43, 37.56, 46.87, 48.11, 50.41, 106.70, 115.24, 119.92, 120.52, 121.08, 126.74, 127.17, 128.14, 129.28, 129.54, 135.62, 138.49, 144.38, 154.12, 154.76, 164.66, 189.50, 197.80, 220.70 ppm. EI-MS m/z: 452.19 Anal. Calcd. for  $C_{30}H_{28}O_4$ : C, 79.62; H, 6.24; 0, 14.14. Found: C, 79.54; H, 6.18.

**Preparation of imino-steroid derivatives.** In a round bottom flask (50 ml), the compound **11** or **12** (0.50 mmol), ethylenediamine (50  $\mu$ l, 0.74 mmol) and boric acid (26 mg, 0.42 mmol) 10 ml of methanol were stirring for 72 h to room temperature. The product obtained was dried under reduced pressure. Then, the residue was purified through the crystallization with the methanol:hexane:water (3:1:1) system

## (4E,8E,15aS)-15a-methyl-7,11,12,12a,12b,13,14,15,15a,16,17, 17a-dodecahydro-6H-cyclopenta[7,8]phenanthro[3,2-b]naphtha[1,8-jk][1]oxa[5,8]diazacyclododecin-15-ol (11)

yielding 54 % of product, m.p. 140-142 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 3380, 3322 and 1220: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 0.76 (s, 3H), 0.80-3.62 (m, 16H), 4.18 (m, 4H), 6.40 (broad, 2H), 6.88 (m, 1H), 7.18 (m, 1H), 7.46 (m, 1H), 7.50-7.70 (m, 5H), 8.30-8.50 (m, 2H) ppm. <sup>13</sup>C NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 12.74, 23.87, 25.82, 27.62, 29.33, 31.16, 35.66, 38.90, 44.06, 44.15, 49.82, 58.96, 81.24, 111.00, 113.00, 120.16, 123.17, 126.02, 126.68, 128.15, 128.72, 128.95, 129.54, 130.63, 133.50, 136.55, 138.44, 144.63, 147.02, 158.64, 165.54 ppm. EI-MS m/z: 544.35 Anal. Calcd. for  $C_{37}H_{44}N_4$ : C, 81.57; H, 8.14; N, 10.28. Found: C, 81.50; H, 8.08.

# 2-(((4E,8E,15E,15aS)-15a-methyl-6,7,11,12,12a,12b,13,14,15a, 16,17,17a-dodecahydro-15H-cyclopenta[7,8]phenanthro[3,2-b] naphtho[1,8-jk][1]oxa[5,8]diazacyclododecin-15-ylidene)amino) ethan-1-amine (12)

Yielding 54 % of product, m.p. 150-152 °C; IR ( $V_{max}$ , cm<sup>-1</sup>) 3380, 3320 and 1222: <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta_{H}$ : 1.00 (s,

3H), 1.22-2.98 (m, 15H), 3.10-3.50 (m, 4H), 4.18 (m, 4H), 4.34 (broad, 2H), 7.10 (m, 1H), 7.18 (m, 1H), 7.44 (m, 1H), 7.52- 7.72- (m, 5H) 8.30-8.50 (m, 2H) ppm.  $^{13}$ C NMR (500 MHz, Chloroform-*d*)  $\delta_{\rm H}$ : 15.96, 21.98, 26.00, 27.00, 27.53, 29.30, 32.42, 37.60, 41.02, 41.05, 44.56, 54.12, 54.29, 58.96, 111.00, 113.94, 120.18, 123.14, 126.00, 126.64, 128.19, 128.95, 129.56, 129.60, 130.63, 133.54, 136.55, 138.43, 144.62, 148.03, 158.68, 165.52, 176.82 ppm. EI-MS m/z: 518.30 Anal. Calcd. for  $C_{34}H_{38}N_4$ O: C, 78.73; H, 7.38; N, 10.80; O, 3.08. Found: C, 78.68; H, 7.30.

*Physicochemical properties of compounds 9 and 10.* Theoretical electronic properties, such as HOMO (Highest Occupied Molecular Orbital), LUMO (Lowest Unoccupied Molecular Orbital) energy, orbital coefficients distribution, molecular dipole moment and HBD (hydrogen bond donor groups) and HBA (hydrogen bond acceptor groups) and TPSA (topological polar surface area) were evaluated using the SPARTAN'06 software [18].

*Evaluation of biological activity.* The rats male (Wistar; weighing 200-250 g) used were obtained from the pharmacochemical laboratory of the Autonomous University of Campeche. It is important to mention that animals were handled in accordance with international standards for the care of laboratory animals [19].

*Reagents.* The different compounds were dissolved in methanol; in addition, the dilutions were obtained using Krebs-Henseleit.

*Experimental design.* The animals were anesthetized with pentobarbital (50 mg/Kg body weight) via intraperitoneal injecting. After, the chest was opened to expose the heart, following descending aorta was cut and the heart was immediately flushed with Krebs-Henseleit solution. Then, the heart was removed and perfused with Krebs-Henseleit\* solution via retrograde at a constant flow rate of 10 ml/min.

\*Krebs-Henseleit solution (pH = 7.4; 35-37  $^{0}$ C) bubbled with gas mixture (CO2, 5% and O2, 95%). Experimental data were done after of an equilibration period (10 min).

**Perfusion pressure.** The perfusion pressure and left ventricular pressure produced by the administration of each compound were determined using a pressure transducer that was bound to a chamber (where was inserted the heart). In addition, the signals were obtained using a computerized data capture system (MP-100).

*Inotropic activity.* To evaluate the inotropic effect, a latex balloon filled with saline solution (0.01 mm, diameter) was inserted into the left ventricle through the left atrium. It is noteworthy, that latex balloon was bound to pressure transducer which was connected to a computerized data capture system (MP-100). After, inotropic effect produced by compounds involved in this study was evaluated by determining left ventricular developed pressure (LV/dP) [19].

# Biological evaluation

First stage

Biological activity exerted by 17β-estradiol and the compounds 6 or 7 against perfusion pressure: Effect produced by the

### Synthesis and evaluation of biological activity from two steroid-diazacyclododecin derivatives on left ventricular pressure

compounds  $17\beta$ -estradiol, 11 or 12 (0.001 nM) and the conditions control on perfusion pressure through of time (3 to 18 min) was determined.

### Second stage

Effects exerted by the compound 12 on left ventricular pressure through estrogen-receptor. 50  $\mu$ l of compound 12 at dose of 0.001 to 100 nM were administered and their effect induced on left ventricular pressure in absence or presence of tamoxifen\* (1 nM) was determined.

\*Duration of preincubation with tamoxifen was by a 10 min equilibration period.

Biological activity produced by  $17\beta$ -estradiol via estrogen receptor. 50 µl of  $17\beta$ -estradiol at dose of 0.001 to 100 nM were

### **3. RESULTS SECTION**

There are several methods for preparation of azabicyclic; however, some reagents used require special conditions [23-25]; analyzing this data, in this report, a new azabicyclic derivative was prepared using different chemical strategies.

**Praparation of tert-butyldimethylsilanyloxy steroid-derivatives (3** or 4). The first stage was achieved by protecting hydroxyl group of estradiol estrone using a previous method reported. This process was carried out in order to avoid a possible reaction of the hydroxyl group with any substance involved in the following reactions. It is noteworthy, that several organosilyl groups have been used as protectors of hydroxyl groups such as tertbutyldimethylsilyl and tert-butyldiphenylsilyl [26]. In this study, hydroxyl groups of  $17\beta$ - estradiol or estrone were protecting with tert-butyldimethylsilyl chloride to form **3** or **4** (Figure 1).

The <sup>1</sup>H NMR spectrum of the compound 3 showed signals at 0.08-0.22 and 0.86 ppm for terbuthyldimethylsylane fragment; at 0.80 ppm for methyl group bound to steroid nucleus; at 1.00 and 1.06-7.80 ppm for steroid moiety. The <sup>13</sup>C NMR spectra display chemical shifts at -4.50, -4.40, 17.80-18.22, 25.70-25.74 ppm for terbuthyldimethylsylane fragment; at 25.34-25.62, 27.76-132.52 and 144.24 ppm for steroid moiety; at 133.34 ppm for carbon bound to nitro group; at 151.72 ppm for ether group. Finally, the mass spectrum from 3 showed a molecular ion (m/z) 545.33.

Other result showed the <sup>1</sup>H NMR spectrum of the compound 4 at 0.22 and 1.00 ppm for terbuthyldimethylsylane fragment; at 0.92 ppm for methyl group bound to steroid nucleus; at 1.20-7.90 ppm for steroid moiety. The <sup>13</sup>C NMR spectra displays chemical shifts at -4.44, 18.22 and 25.74 ppm for terbuthyldimethylsylane fragment; at 13.82 ppm for methyl group bound to steroid nucleus; at 21.70-25.51, 26.40-132.00 and 143.94 ppm for steroid moiety; at 133.34 ppm for carbon bound to nitro group; at 151.72 ppm for ether group; at 219.70 for ketone group. In addition, the mass spectrum from 4 showed a molecular ion (m/z) 429.23.

administered and biological activity induce on left ventricular pressure in absence or presence of compound  $12^{**}$  (0.001 nM) was determined. \*\*Duration of preincubation with compound 12 was by a 10 min equilibration period.

### Statistical analysis

Experimental data are display as average  $\pm$  SE, using each heart as its own control. The results were analyzed with a statistical package (ANOVA) with p = 0.05 [20].

# Theoretical evaluation of the interaction between compounds 6 or 7 with phosphodiesterase-4B.

A theoretical analysis was carried out using a docking program (DockingServer) [21]. Estrogen receptor (3008 protein) was used to determine the interaction of compounds 11 or 12 with the enzyme [22].



**Figure 1**. Preparation of two butyl-dimethyl-silanyloxy-nitro-steroid derivatives (3 or 4). Reaction of estradiol (1) or estrone (2) with *tert*-Butyldimethylsilyl chloride to form 3 or 4.

### Synthesis of two steroid-naphtalen-carbaldehydederivatives-tertbutyl-dimethyl-silanyloxy complex (5 or 6).

Several carbaldehyde derivatives have been prepared using some reagent such as POCl<sub>3</sub> [27], nBuLi/THF [28], Cu/Fe [29], difluoro(phenylsulfanyl) methane [30], RhCl<sub>2</sub> [31] and others. However, some of these reagents are expensive and difficult to handle; therefore, in this study **5** or **6** were prepared using a previously method reported [32]. The compounds 3 or 4 were reacted with DMSO to form **5** or **6** (Figure 2). The <sup>1</sup>H NMR spectrum of the compound **5** showed signals at 0.06-0.18, 0.86 and 1.00 ppm for terbuthyldimethylsylane fragment; at 0.80 ppm for methyl group bound to steroid nucleus; at 0.98 and 1.10-6.52 ppm for steroid moiety; at 7.08-8.40 ppm for phenyl groups bound to both aldehyde and ether groups; at 9.96 ppm for aldehyde group.

Rosas-Nexticapa Marcela, Figueroa-Valverde Lauro, Diaz-Cedillo Francisco, Mateu-Armand Virginia, Lopez-Ramos Maria, García-Cervera Elodia, Pool Gómez Eduardo, García-Martínez Rolando, Parra-Galindo Perla, Cauich-Carrillo Regina, Alfonso-Jimenez Alondra, Cabrera-Tuz Jhair



**Figure 2**. Synthesis of two steroid-naphtalen-carbaldehydederivatives (5 or 6). Reaction of 3 or 4 with 2-hydroxy-1-naphthaldehyde (ii) to form 5 or 6. Then 5 or 6 reacted with hydrofluoric acid (ii) to synthesis of 5 or 6.

The  ${}^{13}$ C NMR spectra displays chemical shifts at -4.55, -4.50, 17.84-18.22 and 25.74-25.92 ppm for terbuthyldimethylsylane fragment; at 15.22 ppm for methyl group; at 25.32-25.56, 27.76-112.50, 118.12, 126.94, 135.62 and 137.62 ppm for steroid moiety; at 114.94, 129.92-126.72. 128.44-132.74, 136.84 and 138.44-144.33 ppm for phenyl groups bound to both aldehyde and ether groups; at 147.92 for ether group bound to diphenyl group; at 197.82 ppm for aldehyde group. Finally, the mass spectrum from **5** showed a molecular ion (m/z) 670.38.

On the other hand, the <sup>1</sup>H NMR spectrum of the compound **6** at 0.18 and 1.00 ppm for terbuthyldimethylsylane fragment; at 0.90 ppm for methyl group; at 1.20-6.52 ppm for steroid moiety; at 7.10-8.40 ppm for phenyl groups bound to both aldehyde and ether groups; at 9.96 ppm for aldehyde group. The <sup>13</sup>C NMR spectra displays chemical shifts at -4.54, 18.22 and 25.92 ppm for terbuthyldi- methylsylane fragment; at 13.80 ppm for methyl group bound to steroid moiety; at 21.78-25.84, 26.40-112.24, 118.12, 126.60 and 137.12 ppm for steroid moiety; at 114.94, 120.93-121.14, 126.74-132.74, 135.62-136.82, 138.50 and 144.38 ppm for phenyl groups bound to both aldehyde and ether groups; at 147.90 ppm for ether group bound to diphenyl groups; at 197.80 ppm for aldehyde group. In addition, the mass spectrum from 6 showed a molecular ion (m/z) 554.28.

### Removal of tert-butyldimethylsilyl protecting groups.

Some studies showed that hydrofluoric acid can be used to the removal of the *tert*-butyldimethylsilyl protecting groups [32, 33]. In this investigation, the compounds 5 or 6 were reacted with a hydrofluoric acid to form the compounds 7 or 8 (Figure 2). The <sup>1</sup>H NMR spectrum of the compound 7 at 0.76 ppm for methyl group bound to steroid nucleus; at 0.82-3.64 and 6.04-5.70 ppm for steroid nucleus; at 5.90 for hydroxyl groups; at 7.12-8.40 ppm for phenyl groups bound to both aldehyde and ether groups; at 9.96 ppm for aldehyde group. The <sup>13</sup>C NMR spectra displays chemical shifts at 15.76 ppm for methyl group; at 113.50, 116.24, 128.00, 137.94 and 143.44 for steroid moiety; at 115.00, 120.92-126.74, 128.40-136.80, 138.52 and 143.94-144.38 ppm for phenyl groups bound to both aldehyde and ether groups; at 151.60 for ether group; at 197.82 ppm for aldehyde group. Finally, the mass spectrum from 7 showed a molecular ion (m/z) 442.21.

The <sup>1</sup>H NMR spectrum of the compound **8** at 0.90 ppm for methyl group; at 1.202.80 and 6.16-6.70 ppm for steroid moiety; at 5.30

ppm for hydroxyl group; at 7.12-8.40 ppm for phenyl groups bound to both aldehyde and ether groups; at 9.16 ppm for aldehyde group. The <sup>13</sup>C NMR spectra displays chemical shifts at 13.80 ppm for methyl group; at 21.74-113.14, 116.28, 127.60, 137.50, 143.48 and 156.54 ppm for steroid moiety; at 115.00, 120.92-126.74, 128.41-138.84, 138.50 and 143.94-144.38 ppm for phenyl groups bound to both aldehyde and ether groups; at 197.80 for aldehyde group; at 220.70 ppm for ketone group. In addition, the mass spectrum from **8** showed a molecular ion (m/z) 440.19.

### Preparation of formyl-steroid-carbaldehyde complex.

The compounds 7 or 8 were reacted (Figure 3) with dimethyl sulfoxide for the synthesis of new steroid-carbaldehyde derivatives (9 or 10).



Figure 3. Preparation of di-carbaldehyde-steroid derivatives (9 or 10). Reaction of steroid-naphtalen-carbaldehydederivatives (7 or 8) with dimethylsulfoxide (iii) to form 9 or 10.

The spectrum of the compound **9** at 0.76 ppm for methyl group; at 0.80-3.62, 7.10 and 7.66 ppm for steroid moiety; at 7.24-7.62 and 7.70-8.40 ppm for phenyl groups bound to ether group; at 10.00-10.44 for both aldehyde groups. The <sup>13</sup>C NMR spectra display chemical shifts at 15.80 ppm for methyl group; at 24.22-107.02, 120.52, 128.54, 129.54, 154.60 and 164.64 ppm for steroid moiety; at 115.20-119.92, 121.12-127.17, 129.24, 135.62-144.36 and 154.76 ppm for phenyl groups bound to ether group; at 189.50-197.80 ppm for both aldehyde groups. Finally, the mass spectrum from **9** showed a molecular ion (m/z) 454.21.

Other results showed the spectrum of the compound **10** at 0.88 ppm for methyl group; at 1.20-7.16 and 7.66 ppm for steroid moiety; at 7.26-7.62 and 7.68-8.40 ppm for phenyl groups; at 10.00-10.44 ppm for both aldehyde groups. The <sup>13</sup>C NMR spectra displays chemical shifts at 13.82 ppm for methyl group; at 21.74-106.70, 120.52, 128.14, 129.54, 154.12 and 164.66 ppm for steroid moiety; at 115.24, 121.08-127.17, 129.28, 135.62-144.38 and 154.76 ppm for phenyl groups bound to ether group; at 189.50-197.80 ppm for both aldehyde groups; at 220.70 ppm for ketone group. Additionally, the mass spectrum from **10** showed a molecular ion (m/z) 452.19.

### Preparation of imino-steroid derivatives.

Several protocol have been used use some reagents for preparation of imino derivatives [34, 35]; however, some methods require special conditions; therefore, in this study, the compounds **9** or **10** were reacted with ethylenediamine using boric acid as catalyst [36] to form **11** or **12** (Figure 4).



**Figure 4**. Preparation of amino-steroid derivative (11 or 12). Reaction dicarbaldehyde-steroid derivatives (9 or 10) with ethylenediamine (iv) to form 11 or 12.

The <sup>1</sup>H NMR spectrum of the compound 11 at 0.76 ppm for methyl group bound to steroid nucleus; at 0.80-3.62, 6.88 and 7.46 ppm for steroid moiety; at 4.18 ppm for methylene groups bound to both imino groups; at 7.18 and 7.50-7.70 for phenyl groups; at 8.30-8.50 ppm for both imino groups. The <sup>13</sup>C NMR spectra displays chemical shifts at 13.20 ppm for methyl group bound to steroid nucleus; at 12.74 ppm for methyl group at 23.87-49.82, 81.24, 113.00, 128.72, 130.63-133.50, 147.82 and 150.64 ppm for steroid moiety; at 111.00, 120.16-128.15, 128.95-129.54. 136.55 and 165.54 ppm for phenyl groups bound to ether group; at 138.44-144.63 for both imino groups. Finally, the mass spectrum from **11** showed a molecular ion (m/z) 544.35.

The <sup>1</sup>H NMR spectrum of the compound **12** at 1.00 ppm for methyl group bound to steroid nucleus; at 1.22-2.98, 7.10 and 7.44 ppm for steroid moiety; at 3.10-3.50 ppm for methylene groups bound to both imino and amino groups; at 4.18 ppm for methylene groups bound to both imino groups; at 4.34 ppm for amino group 7.18 and 7.52-7.72 ppm for both phenyl groups bound to ether group; at 8.30-8.50 ppm for both imino groups. The <sup>13</sup>C NMR spectra displays chemical shifts at 15.96 ppm for methyl group bound to steroid nucleus; at 21.98-37.60, 41.05-44.56, 54.29, 113.84, 129.60-133.54 and 148.03-158.68 ppm for steroid moiety; at 41.00 and 52.12 ppm for methylene groups bound to both amino and imino groups; at 58.96 ppm for methylene groups bound to both imino groups; at 111.00, 112.18-129.56, 136.55 and 165.52 ppm for phenyl groups bound to ether group; at 135.43-144.62 and 176.82 ppm for imino groups. Additionally, the mass spectrum from 12 showed a molecular ion (m/z) 518.30.

#### Electronic parameters evaluation (HOMO and LUMO).

The molecular orbitals HOMO and LUMO (Figure 5, Table 1) for the compounds **11** and **12** were theoretically evaluated with SPARTAN'06 software, using Hartree-Fock method at 321-G level [37].

The results showed in Figure 5 indicated that LUMO values were lower for the compound 12 compared with 11; in addition, HBD and HBA values were different for two compounds (table 1), these data indicate that 12 have a different electron donation ability compared to 11.



**Figure 5**. Molecular Orbitals (HOMO and LUMO) involved in the compounds 11 (A) and 12 (B). Visualized with SPARTAN'06 software.

Table 1. Structural properties						
Parameter	Compound 11	Compound 12				
logP	1.37	1.42				
HBD count	1	0				
HBA count	4	5				
PSA	38.437 Å <sup>2</sup>	52.188 Å <sup>2</sup>				
Polarizability	79.98	83.84				

### **Biological activity**

Effect induced by both compound 11 and 12 against perfusion pressure. The effect induced by the compounds 11 or 12 and  $\beta$ -estradiol against left ventricular pressure was evaluated in a Langendorff model. The experimental data showed that compound 12 decreased the left ventricular pressure in a similar form that estradiol.



**Figure 6.** Effects induced by  $17\beta$ -estradiol, compounds **11** and **12** on left ventricular pressure (LVP).  $50\mu$ L of  $17\beta$ -estradiol, **11** and **12** at dose of 0.001 to 100 nM were administered and the biological activity against LVP was evaluated. Experimental data obtained shown that compound **12** decreased LVP (P = 0.06) in a similar form that  $17\beta$ -estradiol. Each bar represents the mean ± SE of 9 experiments.

To evaluate the possibility that compound **12** could exert its activity through the activation of the estrogen receptor, in this study the effect induced by compound **12** on left ventricular pressure (LVP) in the absence or in presence of an estrogen antagonist (tamoxifen) was evaluated (Figure 7). The results indicated that compound **12** decrease the **LVP** in a dose-dependent

manner and their biological activity was blocked by tamoxifen; this phenomenon suggests that molecular mechanism involved in produced by compound 12 was via estrogen receptor activation.



**Figure 7**. Effect produced by compounds 12 on left ventricular pressure (LVP) in absence or presence of tamoxifen. Intracoronary boluses  $(50\mu L)$  of **12** [0.001 to 100nM] were administered in absence or presence of tamoxifen and their biological activity against LVP was evaluated. The results showed that compound **12** decreased LVP (P = 0.06) and this effect was inhibited with tamoxifen. Each bar represents the mean  $\pm$  SE of 9 experiments.

Analyzing this data and other reports, which showed that fulvestrant (steroid derivative) inhibit the biological activity of 17 $\beta$ -estradiol exerted on left ventricular pressure [15]; therefore, in this investigation the effect induced by 17 $\beta$ -estradiol against left ventricular pressure was evaluated in presence or absence of compound **12** (Figure 8).



**Figure 8.** Biological activity induced by 17β-estradiol on left ventricular pressure (LVP). Intracoronary boluses (50 $\mu$ L) of 17β-estradiol [0.001 to 100nM] were administered in absence or presence of compound **12** [0.001 nM] and their biological effect against LVP was evaluated. The results showed that 17β- estradiol decreased LVP (p = 0.05) and this effect was inhibited by the compound **12**. Each bar represents the mean  $\pm$  SE of 9 experiments.

## 4. CONCLUSIONS

The experimental results showed that; 1) compound 12 decreased left ventricular pressure via estrogen receptor; 2) the effect exerted by estradiol was inhibited in presence of 12; 3) the

### **5. REFERENCES**

[1] Von-Schacky C., Associations of Omega-3 fatty acid supplement use with cardiovascular disease risks: meta-analysis of 10 trials involving 77 917 individuals, *Altern. Ther. Health Med.*, 24, 2, 8-9, **2018**.

[2] Zhao D., Guallar E., Ouyang P., Subramanya V., Vaidya D., Ndumele C., Lima J., Allison M., Shah S., Bertoni A., Budoff M., Post W., Michos E., Endogenous Sex Hormones and Incident Cardiovascular Disease in Post-Menopausal Women, *J. Am. Coll. Cardiol.*, 71, 22, 2555-2566, **2018**.

The results indicated that  $17\beta$ -estradiol decreased the left ventricular pressure; however, this phenomenon was inhibited by the compound 12. These results suggest that compound 12 could be an estrogen selective agonist. It is noteworthy, that there are several theoretical studies which have been used to predict the biological activity of several substances as estrogen agonist [38-40].

### Theoretical analysis.

To evaluate the interaction of compound 12 with estrogen receptor (30s8 protein) [22] a Docking model was used [21]. The results showed (Figure 9 and Table 2) the interaction of compound 12 with the amino acid residues of 30s8 protein such as  $Pro_{224}$ ,  $Leu_{319}$ ,  $Leu_{320}$ ,  $Leu_{327}$ ,  $Asp_{321}$ ,  $Ala_{322}$ ,  $Glu_{323}$ ,  $Pro_{324}$ ,  $Pro_{325}$ ,  $Ile_{326}$ , and  $Leu_{327}$ . It is important to mention of amino group free of compound **12** could bound to  $Leu_{327}$ ,  $Ile_{326}$  and  $Pro_{324}$  aminoacid residues.



Figure 9. Distance between compound 10 and aminoacid residues of estrogen receptor (30s8 protein).

Table 2. Distanc	e between the	e aminoacid	residues	of 3os8	protein	and
	both nitrog	en and oxy	gen atoms	5		

Aminoacid Residues	Nitrogen (Å)	Oxygen (Å)
Pro <sub>224</sub>	-	26.66
Leu <sub>319</sub>	22.04	-
Leu <sub>320</sub>	29.56	-
Asp <sub>321</sub>	24.99	-
Ala <sub>322</sub>	24.93	26.01
Glu <sub>323</sub>	26.01	25.51
Pro <sub>324</sub>	24.07	-
Pro <sub>325</sub>	26.07	-
Ile <sub>326</sub>	20.07	-
Leu <sub>327</sub>	22.93	-

data indicate that compound 12 could exert their biological activity which the interaction with could involves some aminoacids residue of the estrogen receptor.

[3] Escalante C., Mora S., Bolaños L., Hormone replacement therapy reduces lipid oxidation directly at the arterial wall: A possible link to estrogens' cardioprotective effect through atherosclerosis prevention, *J. Midlife Health.*, 8, 1,11-16, **2017**.

[4] Gourdy P., Guillaume M., Fontaine C., Adlanmerini M., Montagner A., Laurell H., Lenfant F., Arnal J., Estrogen receptor subcellular localization and cardiometabolism, *Mol. Metab.*, **2018**.

[5] Sharma G, Mauvais F, Prossnitz E., Roles of G proteincoupled estrogen receptor GPER in metabolic regulation. *J. Steroid Biochem. Mol. Biol.*, 176, 31-37, **2018**.

[6] Kunnas T., Laippala P., Penttila A., Lehtimaki T., Karhunen P., Association of polymorphism of human alpha oestrogen receptor gene with coronary artery disease in men: a necropsy study, *Br. Med. J.*, 321, 273-274, **2000**.

[7] Lu H., Higashikata T., Inazu A., Association of estrogen receptoralpha gene polymorphisms with coronary artery disease in patients with familial hypercholesterolemia, *Arterioscler. Thromb. Vasc. Biol.*, 22, 817-823, **2002**.

[8] Evangelopoulos D., Alevizaki M., Lekakis J., Molecular analysis of the estrogen receptor alpha gene in men with coronary artery disease: association with disease status, *Clin. Chim. Acta.*, 331, 37-44, **2003**.

[9] Petrovic D., Peterlin B., Estrogen receptor dinucleotide (ta) polymorphism does not predict premature myocardial infarction in Caucasian women, *Cardiology.*, 99, 163-165, **2003**.

[10] Lehtimaki T., Kunnas T.A., Mattila K.M., Coronary artery wall atherosclerosis in relation to the estrogen receptor 1 gene polymorphism: an autopsy study, *J. Mol. Med.*, 80, 176-180, **2002**.

[11] Pelzer T., Jazbutyte V., Hu K., Segerer S., Nahrendorf M., Nordbeck P., Bonz A., Muck J., Heinrich K., Hegele C., The estrogen receptor- $\alpha$  agonist 16 $\alpha$ -LE2 inhibits cardiac hypertrophy and improves hemodynamic function in estrogen-deficient spontaneously hypertensive rats, *Cardiov. Res.*, 67, 4, 604-612, **2005**.

[12] Huang Y., Tomoharu S., Mashkoor A., Ya H., Takao S., Kirby I., Mechanism of cardioprotection following trauma-hemorrhagic shock by a selective estrogen receptor- $\beta$  agonist: up-regulation of cardiac heat shock factor-1 and heat shock proteins, *J. Mol. Cell. Cardiol.*, 40, 1, 185-194, **2006**.

[13] Figtree A., Lu Y., Webb C., Collins P., Raloxifene Acutely Relaxes Rabbit Coronary Arteries In Vitro by an Estrogen Receptor-Dependent and Nitric Oxide–Dependent Mechanism, *Circulation.*, 100, 1095-1101, **1999**.

[14] Umetani M., Domoto H., Gormley A., Yuhanna I., Cummins C., Javitt N., Korach K., Shaul P., Mangelsdorf D., 27-Hydroxycholesterol is an endogenous SERM that inhibits the cardiovascular effects of estrogen, *Nature Med.*, 13, 1185-1192, **2007**.

[15] Mercier I., Mader S., Calderone A., Tamoxifen and ICI 182,780 negatively influenced cardiac cell growth via an estrogen receptor-independent mechanism, *Cardiov. Res.*, 59, 4, 883-892, **2003**.

[16] Tomson A., Horwitz J., Some 2-and 4-Substituted Estrone 3-Methyl Ethers, J. Org. Chem., 24, 2056-2059, **1959**.

[17] Pezzella A., Manini P., Di Donato P., Boni R., Napolitano A., Palumbo A., 17-Estradiol nitration by peroxidase/H2O2/NO2<sup>-</sup>: a chemical assessment, *Bioorg. Med. Chem.*, 12, 2927–236, **2004**.

[18] Sahin A, Sacan A., Understanding the toxic potencies of xenobiotics inducing TCDD/TCDF-like effects. *J. SAR and OSAR Envir. Res.*, 2, 117-131, **2018**,

[19] Figueroa-Valverde L., Díaz-Cedillo F., García-Cervera E., Rosas-Nexticapa M., Pool-Gómez E., Lopéz-Ramos M., Rodriguez-Hurtado F., Chan-Salvador M., Evaluation of activity of an estrogen-derivative as cardioprotector drug using an ischemia-reperfusion injury model, *Int. J. Clin. Exp. Med.*, 8, 8, 12041-12055, **2015**.

[20] Hocht C., Opezzo J., Gorzalczany S., Una Aproxi-mación Cinética y Dinámica de Metildopa en Ratas con Coartación Aórtica Mediante Microdiálisis, *Revista Arg. Cardiol.*, 67, 769-773, **1999**.

[21] Réau M., Langenfeld F., Zagury F., Montes., Predicting the affinity of Farnesoid X Receptor ligands through a hierarchical ranking protocol: a D3R Grand Challenge 2 case study, *J. Com. Mol. Des.*, 32, 1, 231–238, **2018**.

[22] Sherin D., Manojkumar T., Flavanoids from Saraca asoca- Ideal Medication for Breast Cancer: A Molecular Simulation Approach, *Biomed J. Sci. Tech. Res.*, 1, 6, 1-3, **2017**.

[23] Neipp C., Martin S., Synthesis of Bridged Azabicyclic Structures via Ring-Closing Olefin Metathesis, *J. Org. Chem.*, 68, 23, 8867-8878, 2003.
[24] Solé D., Peidró E., Bonjoch J., Palladium-Catalyzed Intramolecular Coupling of Vinyl Halides and Ketone Enolates. Synthesis of Bridged Azabicyclic Compounds. *Org. Lett.*, 2, 15, 2225-2228, 2000.

[25] Arjona O., Čsákÿ A., Medel R., Plumet J., Domino Metathesis of 2-Azanorbornenones: A New Strategy for the Enatioselective Synthesis of 1-Azabicyclic Compounds, *J. Org. Chem.*, 67, 4, 380-1383, **2002**.

[26] Corey E., Venkateswarlu A., Protection of hydroxyl groups as tertbutyldimethylsilyl derivatives, *J. Am. Chem. Soc.*, 94, 17, 6190-6191, **1972**.

[27] Zeng L., Bao W., Zheng L., Dong Q., Tian L., Synthesis, crystal structure, DNA interaction and antioxidant activities of two novel water-soluble Cu(2+) complexes derivated from 2-oxo-quinoline-3-carbaldehyde Schiff-bases, *Eur. J. of Med. Chem.*, 44, 11, 4477-4484, **2009**.

[28] Journet M., Cai D., Kowal J., Larsen R., Highly efficient and mild synthesis of variously 5-substituted-4-carbaldehyde-1,2,3-triazole derivatives, *Tetrahedron Lett.*, 42, 52, 9117-9118, **2001**.

[29] Chen M., Peng J., Mao T., Huang J., Cu/Fe-Cocatalyzed Meyer–Schuster-like Rearrangement of Propargylic Amines: Direct Access to *E*- $\beta$ -Aminoacryaldehydes, *Org. Lett.*, 16, 24, 6286-6289, **2014**.

[30] Betterley N., Kongsriprapan S., Chaturonrutsamee S., Chutima Kuhakarn., Electrophilic Aromatic Formylation with Difluoro(phenylsulfanyl) methane, *Synthesis.*, 50, 10, 2033-2040, **2018**.

[31] Zou M., Liu J., Tang C., Jiao N., Rh-Catalyzed N-O Bond Cleavage of Anthranil: A C–H Amination Reagent for Simultaneous Incorporation of Amine and a Functional Group, *Org. Lett.*, 18, 12, 3030-3033, **2016**.

[32] Figueroa-Valverde L., Díaz-Cedillo F., García-Cervera E., Pool-Gómez E., Rosas-Nexticapa M., López-Ramos M., Vera-Escobedo I., Design and synthesis of two triazonine-carbaldehyde derivatives using several chemical tools, *J. Saudi Chem. Soc.*, 22, 2, 183-197, **2018**.

[33] Paat J., Figueroa L., Lopez M., Hau L., Diaz F., Garcia E., Pool E., Rosas M., Mateu V., Preparation of an oxetanphenyltetrahydropyridazine-3,6-dione derivative using some chemistry tools, *Iran Chem. Comm.*, 6, 3, 218-311, **2018**.

[34] Pinheiro A., Da-Silva S., Roisnel T., Kirillov E., Carpentier J., Osvaldo Casagrande L., Synthesis and structural characterization of zirconium complexes supported by tridentate pyrrolide-imino ligands with pendant *N*-, *O*- and *S*-donor groups and their application in ethylene polymerization, *New J. Chem.*, 42, 1477-1483, **2018**.

[35] Wang Z., Solan G., Mahmood Q., Liu Q., Ma Y., Hao X., Sun W., Bis(imino)pyridines Incorporating Doubly Fused Eight-Membered Rings as Conformationally Flexible Supports for Cobalt Ethylene Polymerization Catalysts, *Organometallics.*, 37, 3, 380-389, **2018**.

[36] Figueroa-Valverde L., Díaz-Cedillo F., García-Cervera E., Pool-Gómez E., López-Ramos M., Rosas-Nexticapa M., Hau-Heredia L., Sarabia-Alcocer B., Design and synthesis of two azetidin-haloperidol derivatives using some strategies, *Oriental J. Chem.*, 30, 3, 947-952, **2014**.

[37] Xue W., Lai W., Observation of negative electron-binding energy in a molecule. *Nature.*, 400, 245-248, **1999**.

[38] Brogi S., Kladi M., Vagias C., Papazafiri P., Roussis V., Tafi A., Pharmacophore Modeling for Qualitative Prediction of Antiestrogenic Activity, *J. Chem. Inf. Model.*, 49, 11, 2489-2497, **2009**.

[39] Tilghman B., Burow G., A novel class of antiestrogenic phytoalexins, *Mol. Cel. Pharm.*, 2, 4, 155-160, **2010**.

[40] Nose T., Tokunaga T., Shimohigashi Y., Exploration of endocrinedisrupting chemicals on estrogen receptor  $\alpha$  by the agonist/antagonist differential-docking screening (AADS) method: 4-(1-Adamantyl)phenol as a potent endocrine disruptor candidate, *Tox. Lett.*, 19, 1, 33-39, **2009**.

[41] Klotz I., Physiochemical aspects of drug-protein interactions: a general perspective, *Ann. N. Y. Acad. Sci.*, 26, 226, 18-35, **1973**.

© 2018 by the authors. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/4.0/).